

5:9 Inhibition by a *Lactobacillus* of the growth of *Brochothrix thermosphacta* in mixed culture.

B.J. SHAY, A.F. EGAN AND P.J. ROGERS*

CSIRO Division of Food Research, Meat Research Laboratory, Cannon Hill, Queensland, 4170, Australia.

* School of Science, Griffith University, Nathan, Queensland, 4111, Australia.

Introduction

Whilst lactic acid bacteria dominate the flora of vacuum-packaged beef, *Brochothrix thermosphacta* may also be present (Pierson, Collins-Thompson and Ordal, 1970; Gill and Newton, 1978; Dainty et al., 1979). However there are a number of apparently conflicting reports concerning the ability of *B.thermosphacta* to grow on vacuum-packaged beef. These studies are discussed by Campbell et al. (1979) who showed that the amount of growth of this organism is controlled by a combination of two factors, the pH of the meat and the availability of oxygen. These results were obtained in pure culture.

In contrast, other studies have suggested that poor growth of *B.thermosphacta* in mixed culture is caused by the other bacteria present. Roth and Clark (1975) observed that the growth of *B.thermosphacta* on vacuum-packaged beef was restricted by *Lactobacilli* and Newton and Gill (1978) showed that a *Lactobacillus* sp. inhibited the growth of *B.thermosphacta* on beef stored under anaerobic conditions.

Previously, Gill (1976) had shown that the amount of growth of a *Lactobacillus* sp. on beef was limited by the rate of diffusion of fermentable substrates from within the meat to the surface and this led Newton and Gill (1978) to suggest that *Lactobacillus* inhibited the growth of *B.thermosphacta* by competition for glucose. However when grown together under anaerobic conditions in glucose-limited continuous culture *B.thermosphacta* displaced the *Lactobacillus* from the chemostat, presumably because it had a higher affinity for glucose. This result led those workers to conclude that the *Lactobacillus* sp. inhibited the growth of *B.thermosphacta* on meat by producing an antibacterial agent, however they were unable to demonstrate inhibition in liquid culture.

We have now examined in more detail the growth of *B.thermosphacta* and a *Lactobacillus* in pure and mixed culture both on beef of high pH and in glucose-limited continuous culture in a chemostat. Under aerobic conditions no significant interactions between the organisms were observed, but under anaerobic conditions the growth of *B.thermosphacta* was inhibited in the presence of the *Lactobacillus*. These results may be explained in terms of competition for glucose.

Materials and Methods

B.thermosphacta was maintained on nutrient agar slopes and *Lactobacillus* L13 (Shay and Egan, 1981) in cooked meat medium. Cultures were stored at 0°C. Growth of inocula, measurement of meat pH, preparation of meat samples, conditions of incubation and preparation of samples for viability determination have been described previously (Campbell et al., 1979). To

determine the viable count of *Lactobacillus* L13 in pure culture, samples were plated on MRS Agar (Oxoid) and Tryptone Soya Agar (Oxoid) supplemented with 0.5% (w/v) yeast extract and 0.2% (w/v) glucose (TSYG agar). For *B.thermosphacta* in pure culture TSYG agar and STAA agar (Gardner, 1966) were used. For mixed cultures all three media were used.

A New Brunswick Bioflo C30 apparatus was used for continuous culture experiments as previously described (Hitchener, Egan and Rogers, 1979). The culture medium consisted of a mineral-salts basal medium (medium 56 of Monod, Cohen-Bazire and Cohn, 1951) used half-strength and supplemented with 0.3% (w/v) yeast extract and 0.1% (v/v) Tween 80. The medium was sterilized by autoclaving and after cooling sterile solutions of manganese sulphate (final concentration 0.3 mM) and glucose were added. The temperature for all continuous culture experiments was 25°C and that for the meat experiments was 5°C.

Results

Pieces of beef semitendinosus muscle, pH 6.5-6.7, inoculated with aerobically grown cells of *B.thermosphacta* and/or *Lactobacillus* L13, were incubated in Thunberg tubes at 5°C. Fig. 1a shows the results of a typical experiment where conditions of growth were aerobic. Starting from an initial population of ca. 10⁴ cells/g of meat, *B.thermosphacta* and the *Lactobacillus* reached maximum populations in excess of 10⁹/g and 10⁸/g respectively. In all experiments, *B.thermosphacta* reached the higher population. In an experiment in which the initial population of *B.thermosphacta* was 10⁵/g and that of the *Lactobacillus* only 10⁴/g, there appeared to be a slight inhibition of the growth rate of the *Lactobacillus* but it still reached the same maximum population.

For similar experiments using anaerobic conditions, cells for use as inocula were also grown anaerobically. Under such conditions the *Lactobacillus* reached populations of 2 x 10⁸ to 10⁹/g in both pure and mixed culture (Fig. 1b). In contrast whilst the population of *B.thermosphacta* exceeded 10⁸/g in pure culture its growth in mixed culture was inhibited. Starting from an initial population of ca. 10⁴/g it reached a maximum of ca. 6 x 10⁶/g and then tended to decline on further storage. In an experiment where the *Lactobacillus* started at an initial count of 10⁴/g and *B.thermosphacta* at 10⁵/g, less inhibition was observed. In this experiment the *Lactobacillus* reached a maximum population of 2-3 x 10⁸/g and *B.thermosphacta* reached ca. 5 x 10⁷/g.

Lactobacillus L13 was grown in continuous culture in the chemostat at 25°C. With 3.7 mM glucose in the input medium the culture was glucose limited over a range of dilution rates from 0.1 to 0.5 h⁻¹ and the population of cells was between 10⁸ and 10⁹/ml. Similar results were obtained under both aerobic and anaerobic conditions of growth, however most pure-culture experiments were done using anaerobic conditions.

When *B.thermosphacta* was grown in glucose-limited continuous culture in the chemostat at 25°C the growth characteristics were similar to those observed previously in a more detailed study (Hitchener, Egan and Rogers, 1979). In the present experiments it reached a maximum population in excess of 10⁸/ml under both aerobic and anaerobic conditions.

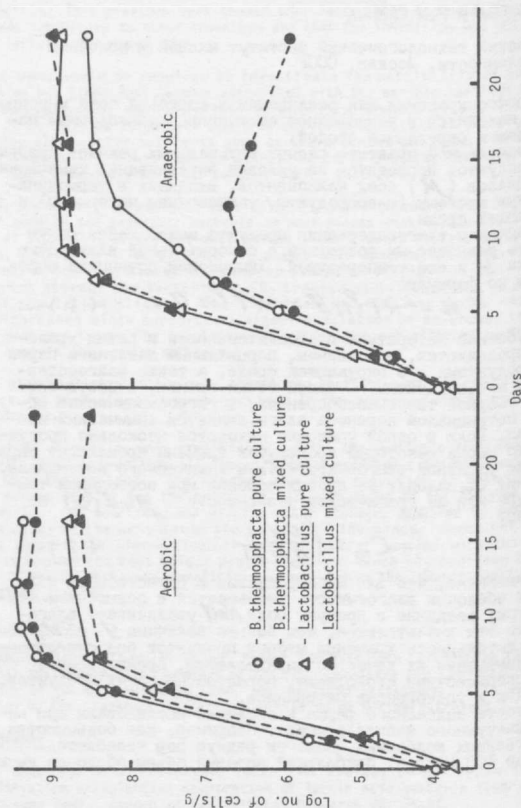


Fig. 1. Growth of *Brochothrix thermosphacta* and *Lactobacillus* L13 in pure and mixed culture at 5°C on beef of pH 6.5-6.7.

The results of a typical experiment in which the two organisms were grown together in mixed culture in the chemostat under conditions of glucose limitation are shown in Fig. 2. Under aerobic conditions, *B.thermosphacta* grew from an initial inoculum of ca. 10⁶ cells/ml to a population in excess of 10⁹/ml, in spite of the presence of a high population (ca. 2 x 10⁸/ml) of the *Lactobacillus*. However when the culture was made anaerobic, the population of *B.thermosphacta* declined to between 10⁶ and 10⁷/ml. Re-establishment of aerobic conditions resulted in its population increasing to the previous level. Under anaerobic conditions, *B.thermosphacta* was never completely eliminated from the chemostat but it continued to co-exist with a much greater population of the *Lactobacillus*.

When the order of inoculation was reversed, similar results were obtained. The *Lactobacillus* grew from an initial population of ca. 10⁶/ml to greater than 10⁸/ml in spite of the presence of a high population of *B.thermosphacta*. Under aerobic conditions the increase in population of the *Lactobacillus* had no significant effect on the pre-existing population of *B.thermosphacta* and high populations (>10⁸/ml) of each organism co-existed together. However if conditions were anaerobic at the time of introduction of the *Lactobacillus* then the population of *B.thermosphacta* declined to ca. 10⁵/ml concomitant with the increase in the population of the *Lactobacillus*.

In further experiments the affinity of each organism for the growth-limiting substrate (glucose) was determined. The steady-state glucose concentration was measured over a range of dilution rates for pure cultures of each organism. The residual glucose concentration increased as the dilution rate was increased, until the maximum growth rate was reached at which point the culture commenced to "wash-out". The data so obtained was plotted in double reciprocal form (Lineweaver-Burk plot) in order to determine K_s for glucose. For *B.thermosphacta* it was found to be ca. 0.1 mM under aerobic conditions and 0.6 mM under anaerobic conditions. For *Lactobacillus* L13 it was ca. 0.1 mM in both cases.

Discussion

The continuous culture experiments show that the composition of a mixed culture of the *B.thermosphacta* and *Lactobacillus* strains used was dependent upon the availability of oxygen. These results are consistent with competition for glucose being the major factor controlling the composition of the population. Under aerobic conditions, K_s for glucose was the same for both bacteria, but under anaerobic conditions the affinity of the facultative *B.thermosphacta* for glucose was reduced, allowing the *Lactobacillus* to dominate in mixed culture. *B.thermosphacta* was not completely eliminated from the culture under anaerobic conditions presumably because it was able to utilize a component of the yeast extract.

Our continuous culture experiments gave different results from those of Newton and Gill (1978). The reason for this is not known as those workers did not determine K_s values.

The fact that the population dynamics of the two organisms growing in mixed culture on meat was similar to those in continuous culture, suggests that competition for substrate may have been a controlling factor in that situation also. Although there is considerable evidence that *Lactobacilli* inhibit the

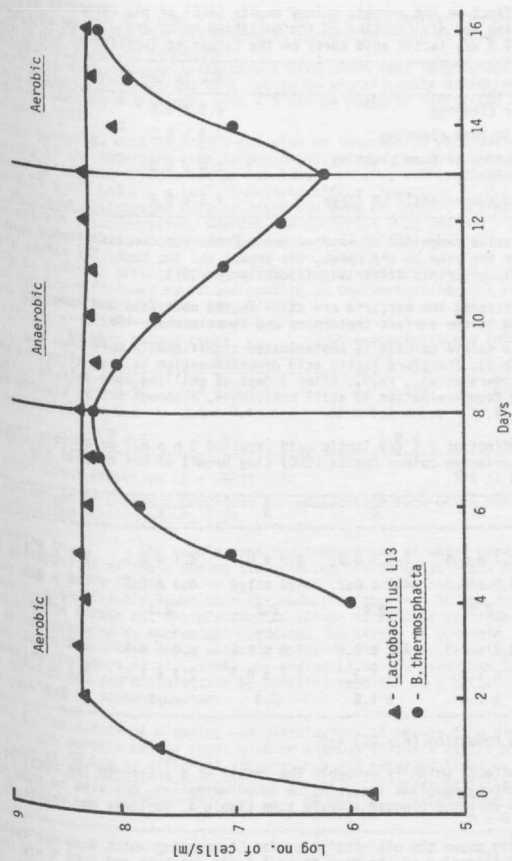


Fig. 2: Growth of *Brochothrix thermosphacta* and *Lactobacillus* L13 at 25°C in glucose-limited continuous culture in the chemostat.

Growth of *B. thermosphacta* on both fresh (Roth and Clark, 1975; Newton and Gill, 1978) and processed meats (Collins-Thompson and Lopez, 1982), there appears to be no direct evidence for antibiotic production in these situations. In some studies it appears that environmental factors may not have been sufficiently closely controlled (discussed by Campbell et al., 1979) and the results presented here demonstrate that competition for substrate must also be considered as a mechanism of inhibition.

Further studies of substrate utilization by bacteria growing on meats are needed as part of wider studies on the interactions between organisms growing in mixed populations on meats. It seems likely that certain lactic acid bacteria may produce bacteriocins during growth on meat. The isolation of such strains would be of considerable importance in view of their potential for future use.

References

- Campbell, R.J., Egan, A.F., Grau, F.H. & Shay, B.J. 1979. *J. Appl. Bacteriol.* **47**, 505-509.
- Collins-Thompson, D.L. & Rodriguez Lopez, G. 1982. *Can. Inst. Food Sci. Technol. J.* **15**, 307-309.
- Dainty, R.H., Shaw, B.G., Harding, C.D. & Michanie, S. 1979. In *Cold Tolerant Microbes in Spoilage and the Environment*, ed. Russell, A.D. & Fuller, R. pp. 83-100. London: Academic Press.
- Gardner, G.A. 1966. *J. Appl. Bacteriol.* **29**, 455-460.
- Gill, C.O. 1976. *J. Appl. Bacteriol.* **41**, 401-410.
- Gill, C.O. & Newton, K.G. 1978. *Meat Sci.* **2**, 207-216.
- Hitchener, B.J., Egan, A.F. & Rogers, P.J. 1979. *Appl. Environ. Microbiol.* **37**, 1047-1052.
- Monod, J., Cohen-Bazire, G. & Cohn, M. 1951. *Biochim. Biophys. Acta* **7**, 585-599.
- Newton, K.G. & Gill, C.O. 1978. *J. Appl. Bacteriol.* **44**, 91-95.
- Pierson, M.D., Collins-Thompson, D.L. & Ordal, Z.J. 1970. *Food Technol.* **24**, 1171-1175.
- Roth, L.A. & Clark, D.S. 1975. *Can. J. Microbiol.* **21**, 629-632.
- Shay, B.J. & Egan, A.F. 1981. In *Psychrotrophic Microorganisms in Spoilage and Pathogenicity*, ed. Roberts, T.A., Hobbs, G., Christian, J.H.B. & Skovgaard, N. pp. 241-251. London: Academic Press.