8:5 Determination of methylthiouracil and analogous thyreostatic drugs H.F. DE BRABANDER and R. VERBEKE

Laboratory of Chemical Analysis of Food from Animal Origin, Veterinary Faculty, University of Ghent, Belgium

### Introduction

Treatment of cattle with thyreostatic drugs is detected by the residues present in plasma, excreta, meat or organs of the animal. Optimal detection of ille-gal treatment with thyreostatics will be achieved through selection of the use of a reliable sensitive detection method. A specific detection procedure of thiourcail and analogous compounds (Fig. 1), based on fluorescence induction of the N80-derivatives (7-chloro-4-nitrobenzo-2-oxa-1,3-diazole) with cysteine.

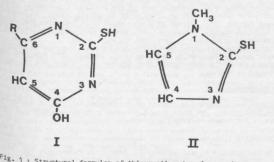


Fig. 1 : Structural formulas of thiouracil and analogous drugs I : 4(6)-R-thiouracil (R=H (TU), Me (MTU), n-propyl (PTU) or phenyl (PhTU) II : 1-methyl-2-mercaptoimidazole (tapazole (TAP))

has been described previously (1). This method was adopted by the BENELUX (2) and EEC (3) for qualitative analysis of these drugs at the 50 ppb level. (3) for qualitative analysis of these drugs at the 50 ppb level, (4) the reaction up of the extracts did not permit to exploit the sensitivity (5) the reaction. A rapid and selective extraction procedure for thyreostatic (1), is presented here allowing a quantitative determination of the thiouracils in various extracts of biological origin.

# Materials and Methods

## $P_{reparation of the mercurated resin}$

 $D_{\rm vacuum}$  1 x 2 (50-100 mesh) is washed successively with 10 bed volumes distilled water, 0.5 N NaOH, dist. water, 0.5 N acetic acid and dist. water. The wet 4. MydroxYmercurifluorescein (250 mg) during 24 hours. The mercurated resin is it treated with water until the eluate is colourless. Afterwards the resin water, treated with 100 ml 0.1 N NeOH and finally washed with 500 ml dist. The mercurated resin is stored in the dark.

 $\ensuremath{\texttt{M}_{\text{CFO-COlumn}}}$  for the clean-up of thy reostatic drugs

A diagram of the chromatographic micro-column, used for the clean-up of thyreo-static drugs, is given in Fig. 2.



The Micro-column is prepared as follows : the column is filled with water and in alars rod is removed. Approximately 0.6 ml mercurated resin is suspended the stars and added to the glass funnel. After sedimentation of the resin in capositing the glass rod on the resin bed, the column is removed. After

- Analytical procedure

2 g of tissue, 2 ml urine, plasma or skim milk are homogenised in 10 ml metha-nol using an ultra-turrax. The internal standard solution (4 (5,6)-dimethyl-2-thiouracii (DMTU), 400 µl) is added and the homogenate centrifuged at 10 000 rpm (12 000 g) during 10 minutes. The supernatant is decanted and subsequently percolated through the mercury column. The column is weshed with water and the thyreestatic drugs are displaced with 5 ml elution solution (0.5 M NaCi; 0.1 N HCi; pH = 1). The eluate is neutralized (100 µl 12 N NaOH) and adjusted to pH = 8. A methanolic MED-CI solution (0.1 ml, 25 µM/ml) is added and the reaction allowed to proceed in the dark at 40° C during one hour (3). There-after, the reaction mixture is adjusted to pH 3-4 by adding 0.2 ml 6 N HC1. The NBD-derivatives are then extracts are dried over sodiumoulfate and concen-trated under a jet of nitrogen, according to the concentration range investi-gated, to a volume of 0.2-1 ml.

Quantitative high performance thin layer chromatography (HPTLC)

The extracts are analysed by bi-dimensional HPTLC using silicage1 60 (alumi-nium sheets; first direction : methylenechloride : methanol 98:2 v/v; second direction : methylenechloride : propionic acid 98:2 v/v). After development and induction of the fluorescence with an alcaline cysteine solution (1), the relative fluorescence intensities of the thyreostatic drug derivatives are measured against the internal standard derivative (DMTU).

#### Results and discussion

- Mercuration of strong anion-exchangers

It was found that strong anion-exchangers (e.g. DOWEX 1) adsorb mercurial dyes (e.g. 2,7-dibromo-4-hydroxymercurifluorescein (DBMF)). Neither D.5 N HCl nor D.5 N NaOH are capable to strip off the dye from the resin. The binding characteristics of DBMF with DOWEX 1 resins of various cross-linkages were tested. DOWEX 1 x 2 was selected for clean-up of thyreostatic drugs; this resin bonds 25 mg DBMF/ml resin, equivalent to 6.7 mg (33  $\mu$ M) Hg /ml wet resin.

- Study of adsorption and elution characteristics of thiouracil on DBMF columns

The adsorption of thiouracil on DBMF columns was studied. After washing the column with distilled water the drug was eluted with an acid salt (0.5 M NaCl) solution. The elution yields of TU, at different pH values, in function of the elution volume, are given in Fig. 3. A salt solution, with a pH value less than 1, results in an optimal elution of thiouracil from the column.

The adsorption and elution yields of TU and other thyreostatic drugs are summarized in Table 1. The adsorption of the drugs from a methanol-water (80:20, v/v) extract (10 ml) on micro-DBMF columns (0.6 ml) is practically quantitative. Through elution with 5 ml of 0.5 M NaCl (0.1 N HCl, pH = 1) most of the thyreostatic drugs studied (TU, MTU, PTU, DMTU) are recovered in a 80 % yield. On the contrarary, lower recoveries were noted for PhTU and TAP. The interaction of the phenyl group with the polystyrene matrix of the resin may explain the strong adsorption of PhTU. The small elution yield of TAP was due to a partial oxidation of the molecule on the column.

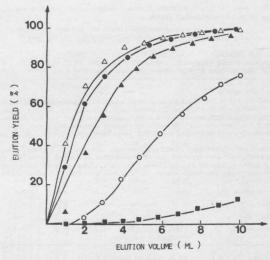


Table 1 : Adsorption and elution yields of thyreostatic drugs on DBMF columns (column 0.4 x 5 cm).

| thyreostatic drug    |     | TU  | MTU | PTU | DMTU | PhTU | TAP             |
|----------------------|-----|-----|-----|-----|------|------|-----------------|
| not adsorbed (%)     |     | 3.8 | 0.5 | 1.5 | 0.5  | 8    | 0.5             |
| elution yield : 5 ml | (%) | 82  | 80  | 78  | 79   | 17   | 60 <sup>*</sup> |
| (pH = 1) 10 ml       | (%) | 96  | 92  | 94  | 95   | 24   | 80*             |

(\*) partially oxidised

#### - Reproducibility and Recovery

The reproducibility of the determination of TU in meat extracts is given in Table 2. The coefficient of variation of the total procedure amounted to 13 %. The recovery of MTU, TU and PTU, at the 100 ppb level, using DMTU as internal standard is given in Table 3. Recoveries for meat were quantitative for the drugs studied. Recoveries in plasma and milk were quantitative for MTU but appreciably lower for PTU and TU.

Table 2 : Reproducibility of the analysis of thyreostatic drugs in meat (TU).

| step in procedure | n  | mean volume ± SD | <pre>coefficient of variation (%)</pre> |  |  |
|-------------------|----|------------------|---|--|--|
| column elution    | 26 | 81 ± 3.9         | 4.8                                     |  |  |
| derivatisation    | 26 | 76 ± 5           | 6.6                                     |  |  |
| HPTLC             | 22 | 88 ± 4.7         | 5.4                                     |  |  |
| total procedure   | 22 | 55 ± 6.9         | 12.6                                    |  |  |
|                   |    |                  |   |  |  |

n : number of determinations

Table 3 : Recovery of the thyreostatic drugs in various media using the internal standard procedure.

| biolog:<br>mater: |                  | concentration<br>added (ppb) | conce<br>PTU | ntration found ±<br>MTU | SD<br>TU  |
|-------------------|------------------|------------------------------|--------------|-------------------------|-----------|
| meat              | (3) <sup>1</sup> | 100                          | 106 ± 5.5    | 102 ± 13.7              | 97 ± 21.8 |
| olasma            | (3) <sup>1</sup> | 100                          | 84 ± 6.2     | 98 ± 5.0                | 76 ± 5.7  |
| milk              | (3) <sup>1</sup> | 100                          | 88 ± 6.9     | 105 ± 3.6               | 85 ± 8.0  |

: number of determinations

- MTU concentrations in thyroid, kidney and muscular tissues of slaughter animals

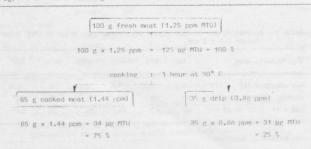
The concentration of MTU in the thyroid, the kidney and some muscles was determined in 5 animals, obtained from regulatory control. The results are summarized in Table 4. As expected, the highest concentration of MTU was found in the thyroid. The MTU concentration in the thyroid was 20-100 times the mean muscle concentration  $(0.2^{-1.9} \text{ pm})$ . The concentration of MTU in the kidney was significantly higher (p  $\leq 0.005$ ) than the concentration found in muscular tissues. A significant difference (p  $\leq 0.005$ ) in MTU content was found between the M. psoas (P), the M. diaphragma (D) and the M. trapesius (T) (non parametric test of Wilcoxon). Classification of the different tissues in descent

ding order of MTU concentration yields : thyroid > kidney > P = D = LD > N > S > G > T

Table 4 : Comparison of the MTU concentrations (ppm) in thyroid, kidney and some muscles of slaughtered animals taken from regulatory control.

| number of the animal | 1    | 2    | 3    | 4    | 5    |  |
|----------------------|------|------|------|------|------|--|
| tissue analysed      |      | 100  |      |      |      |  |
| thyroid              | 30.6 | 48   | 53.2 | 41.5 | 37.5 |  |
| kidney               | 0.30 | 0.81 | 2.1  | 2.3  | 2.4  |  |
| M. long dorsi (LD)   | 0.16 | 0.52 | 1.0  | 2.3  | 1.8  |  |
| M. psoas (P)         | 0.21 | 0.63 | 1.7  | 2.3  | 1.3  |  |
| cervical muscle (C)  | 0.22 | 0.40 | 1.2  | 1.7  | 1.5  |  |
| M. gastrocnemius (G) | 0.15 | 0.79 | 1.8  | 1.6  | 0.8  |  |
| M. trapesius (T)     | 0.17 | 0.54 | 1.0  | 1.7  | 1.2  |  |
| M. sorius (S)        | 0.18 | 0.46 | 3.0  | 1.7  | 1.2  |  |
| M. diaphragma (D)    | 0.19 | 0.59 | 1.7  | 2.0  | 1.3  |  |

Fig. 4 : Effect of cooking on the residue concentration of MTU in meat.



- Effect of cooking on the MTU concentration of meat

8:7

Most of the meat consumed is prepared by heating. The fate of MTU during her ting of meat was investigated using muscles of animals, taken from regulatory control at the slaughterhouses. A meat cut was divided in two portions : one portion was analysed directly. The other portion was heated in a plastic bag at 90° C during one hour. The cooked meat and the drip were separated. All fractions were analysed for its MTU content. In total, 4 different muscles have been analysed using the prof dure described. Typical results are given in Fig. 4. Our measurements show that MTU is not appreciably destroyed in meat after prolonged heating. Since only 25 s of the

dure described. Typical results are given in Fig. 4. Our measurements show that MTU is not appreciably destroyed in meat after prolonged heating. Since only 25 % of the total MTU content is recovered in the drip (= 35 % of the muscle weight), he MTU residues are concentrated in the cooked meat.

#### Conclusions

The use of a low cost mercurated adsorption column allows a selective and reproducible extraction of the thyreostatic drugs from samples of biological origin. This repid clean-up procedure, coupled with the specific fluorescended detection after HPTLC-chromatography of the thisuracil derivates, permits a quantitative determination of these drugs at the poblevel. In comparison we arise methods (1-3) the use of a mercurated resin increases the selective and speed of analysis : routinely more than 20 samples can be prepared for thyroid were 20-100 times higher than in the corresponding muscular tissues. Cooking experiments demonstrated that MTU residues in meat are not appreciative destroyed by heating.

### Acknowledgements

This work was supported by a grant of the Belgian Ministry of Public Heal<sup>th</sup> (Meat Inspection). We thank Mrs. A. Van Meir - Gregoire and Mrs. D'Haez<sup>a</sup> Naessens for expert technical assistance.

#### References

- 1. De Brabander, H.F. and Verbeke, R., (1975), J. Chromatogr. 108, 141.
- Verbeks, R. and De Brabander, H.F., (1975) : The Detection of Thyreostadi Drugs in Meat and Organs of Slaughter Animals, Document Benelux Economised Unie, SP/LAB/h 75 (1).
- Verbeke, R. and De Brabander, H.F., (1978): Method of analysis for deter-ting anti-thyroid substances in fresh muscle tissue. Commission of Europe Communities, Directorate General for Agriculture, Document No. 1123/VI/78-Brussele Brussels.
- 4. Verbeke, R. and De Brabander. H.F., unpublished results.