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COMPARISON OF THE METHOD OF DEFROST OF PRE-COOKED FRO ZEN PRODUCTS FOR MICROBIOLOGICAL ANALYSIS

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SUMMARY

To face the problem of preparing a sample of frozen food to microbiological analysis two alternatives have been choosed: either a quick defrost at a relatively high temperature for a short period or a slow one at a fairly lower temperature for a frankly longer time. The frozen sample was considered as the pattern. It was determined the enumeration of the following micro organisms: aerobic mesophilic, psychrotrophic, yeasts and moulds.

The analysis concerned a total amount of 20 samples for med by wraps of small pastry or pies which had previous ly been baked, then frozen and then re-baked before consuming.

Results indicate that the method of slow defrost usual ly presents higher enumeration of microorganisms than the quick defrost. This is most pronounced with the mesophilic and psychrotrophic microorganisms.

INTRODUCTION

The first problem we face in the microbiological analysis of frozen food is the taking and preparation of samples

We pretend to get a sample for analysis which besides being representative of the whole food is also obtained aseptically. These two requirements often need the whole defrosting of the nourishing product or the use of special equipment for the aseptical sampling of the frozen food.

The method using frozen products is in principle the less exposed to changes in the microflora values. Sin ce it is possible to work in the required aseptical conditions, the whole sample or some of its parts do not need to be submitted to temperatures which may cau se microbial multiplication.

On the other hand when it is necessary to defrost the product in order to get a correct sampling, there are two technics which may be used: the slow defrost at about $+5^{\circ}$ C for the time necessary to complete defrost (in some cases for 18 hours) and the quick defrost at about 45° C for fairly shorter periods (some minutes).

Both defrosting methods have been recommended, reco gnizing in any of them advantages and disadvantages, considering the possible multiplication or des truction in the product existing microflora. And this may happen because there are always some parts of the sample which reach less advisable temperatures be fore the whole sample is completely defrosted. When using the quick defrost we may even obtain a pasteuri zation of the sample surface if the external temperature exceeds 45°C, due to the extreme sensibility to heat acquired by the frozen food flora. In the two de frosting methods the sample surface temperature may permit the microbial multiplication before the sample inside defrost.

Nevertheless this risk may be controlled or even avoided, considering the short period during which that temperature is maintained in quick defrost and becau-

se the highest temperature reached in slow defr^{0^S} quires a very long latency period.

However, as we had not experimented all these the tical knowledges yet, we still had some doubts a the best option when analysing frozen food. Some the structure and size of the food require a tain type of the sample preparation, but in the of pre-prapared frozen food, such as croquettes, try, etc., of small size, it is possible to use a the referred methods. What has been suggested is a parative study.

MATERIAL AND METHODS

We have analysed 20 samples of Wraps of Small ${}^{p \, g \, i}$ and Pies.

Each sample was formed by a wrap containing sever unities of the same product.

Calculations in microbiological analysis are mo^{s^i} posed to changes by the method of the sample prefition:

Counting of Microorganisms at 30°C (mesophilic) Counting of Microorganisms at 6,5°C (Psychrotrop) Counting of Moulds Counting of Yeasts

Samples preparation

Each wrap, containing several unities of the same ^f duct, was divided into 3 equal parts distributed ^b sterilized containers.

One of them was immediately submitted to microbiolical analysis (frost product), another was put in a ol chamber at about 5°C, for 18 hours (slow defrost) and the other was put in a water-bath at about 45 for 30 minutes (quick defrost).

And then they were immediately submitted to micro^{bl} logical analysis.

For the samples preparation we have used tryptones as solvent which was left in contact with the sam for about 30 minutes after they were homogenist with Ultra-Turrax.

Technics used

For the Mesophilic Microorganisms counting we used ^{\$} Standard 2293.

For the Psychrotrophic Microorganisms we adapted ¹⁵ DIS Standard 6730.

For the Moulds and Yeasts counting we followed technic described in the Portuguese Standard 32771 according to which the Cooke Rose Bengal Agar media is used added by chlorotetracycline (35 mg/l) and y namycine (50 mg/l) surface inoculated. Counting carried out after 5 days. RESULTS

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Microbial Contents (Averages)

Microorganisms	Frozen	Quick defrost	Slow defrost
Mesophilic	2,2 x 10 ⁶	2,8 x 10 ⁶	$1,1 \times 10^7$
Psychrotrophic	2,0 x 10 ⁶	2,1 x 10 ⁶	8,6 x 10 ⁶
Moulds	2,5 x 10 ³	3,1 x 10 ³	$6,3 \times 10^3$
Yeasts	1,0 x 10 ⁴	1,0 x 10 ⁴	1,3 x 10 ⁴

DISCUSSION AND CONCLUSIONS

Considering the frozen sample as a standard, we notice that the samples submitted to slow defrost show almost always higher quantities of microorganisms than those submitted to quick defrost. This difference is more significant for the Mesophilic and Psychrotronhic Microorganisms. This may result from the fact that the defrost period is long enough to permit some bacterial growth in the parts having reached higher temperatures. As to the Moulds and Yeasts we have not noticed so significant differences in any of the methods we used.

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