

## BACTERIOLOGICAL QUALITY ASSURANCE OF PORCINE TONGUES.

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## SUMMARY

## INTRODUCTION

Porcine tongues are mainly used for human consumption and generally incorporated in heat processed meat products. They are often visibly contaminated with blood, saliva, hair and dirt. Their keeping quality is limited. Bacteriological Quality Assurance (BQA) along the meat production-line has to rely on longitudinally integrated Good Manufacturing Practices (GMP's). So far, BQA concerning porcine tongues has appeared to be insufficient. The purpose of the described investigation is (a) to obtain information on the bacteriological quality of porcine tongues in the different stages of the slaughtering process; (b) to elaborate GMP's for the production of porcine tongues.

## MATERIALS AND METHODS

Line studies were carried out in sixteen slaughterhouses. Two investigators qualified the contamination grade of porcine tongues relying on fixed criteria, using a check list. At two plants a bacteriological line control was carried out twice. At five stages of the slaughtering process i.e. after (a) bleeding (b) dehairing (c) evisceration (d) collection (e) centrifugation, samples of tongue mucosa were taken. The following colony counts per cm<sup>2</sup> were assessed: mesophilic aerobic colony count, *Enterobacteriaceae*, Gram negative bacteria and *Brochothrix thermosphacta*. Histobacterioscopic examination was carried out, in order to assess the numbers and location of

microorganisms in tongues. Experiments on an advanced centrifuge process under standardised conditions were involved in the investigation. The effects of time: (20s vs 40s), load: (10kg vs 15kg) and water expenditure: (20 l per min. vs 40 l per min.) on the bacteriological quality and cleaning efficiency were assessed. Aerobic colony counts and *Enterobacteriaceae* per cm<sup>2</sup> tongue mucosa were determined and histobacterioscopic examination of the mucosa was carried out.

## RESULTS

After stunning and bleeding the mucosa of the tongues was contaminated with dirt, blood and mucus. Contamination of tongues with stomach contents and damage of tongues took place during dehairing and polishing. The grade of the visual perceptible contamination depends on: (a) filling grade of the stomach (b) type of machinery.

Specific cleaning equipment in the slaughterline for tongues has hardly been developed. Cleanliness of the tongues was insufficient at 11 of the 16 plants. The highest bacteriological contamination of porcine tongues was found after bleeding. Mesophilic aerobic colony count, *Enterobacteriaceae*, Gram negative bacteria and *Brochothrix thermosphacta* were approx. 6.1, 2.9, 3.4 and 4.1 log<sub>10</sub> N per cm<sup>2</sup> respectively. Production stages reducing the contamination were scalding and the centrifuge process. The results of the bacteriological examinations were substantiated by the histobacterioscopic findings. By application of the advanced centrifuge process a 100 fold reduction of the transient flora viz. *Enterobacteriaceae* and in optimal cleanliness of the mucosa of porcine tongues can be attained. Considering economic as well as hygienic aspects a time, load, water expenditure ratio of 20 s/ 15 kg/ 20 l per min. is preferable. GMP concerning BQA of porcine tongues includes (a) slaughtering of pigs with an empty stomach only. (b) avoiding contamination during evisceration (c) application of a cleaning process, e.g. the centrifuge process.

## INTRODUCTION

Porcine tongues are mainly used for human consumption and generally incorporated into heat-processed meat products. After slaughter they are often visibly contaminated with blood, saliva, hair, and dirt. Their keeping quality is low. When they are used for incorporation into meat products, tongues must be clean and must have an acceptable bacteriological quality.

Bacteriological Quality Assurance (BQA) of various meats along the meat production-line has to rely on longitudinally integrated Good Manufacturing Practices (GMP's). So far, BQA concerning porcine tongues has proved to be insufficient. The purpose of the investigation described below is (a) to obtain information on the bacteriological quality of porcine tongues in the different stages of the slaughtering process; (b) to elaborate GMP's for the production of porcine tongues. The described experiments here are part of the GMP-development research programme for variety meats of the Departement.

## MATERIAL AND METHODS

## Line studies

Line studies were carried out in sixteen pig slaughterhouses. Two investigators assessed the contamination grade of porcine tongues, relying on fixed criteria and using a checklist. The effect of cleaning processes was expressed in the following code: ++ = good, + = sufficient, - = insufficient, -- = bad.

## Bacteriological line control

At two of the sixteen plants (A, B) a bacteriological line control was carried out twice. At five stages of the slaughtering process, i.e. after (a) bleeding (b) dehairing (c) evisceration (d) collection (e) centrifugation 10 tongues were collected, and samples of tongue mucosa were taken. With a cork borer (φ15 mm) one tissue sample per tongue was punched out. At each line control, tongues collected for sampling originated from one herd.

The following colony counts per cm<sup>2</sup> were assessed: mesophilic aerobic colony count: Tryptone Glucose Extract Agar (TGEA, Difco 002.01 3 d at 30°C). *Enterobacteriaceae*: Violet Red Bile Glucose Agar (VRBG, Oxoid CM 485, 1 d at 37°C) (3). Gram-negative bacteria: Olson's medium (3 d at 20°C) (5).

*Brochothrix thermosphacta*: Gardner's medium (3 d at 20°C) (1).

In order to get an impression of the distribution of the numbers of micro-organisms on tongues, histobacterioscopic examination was carried out. At each of the 5 stages of the slaughtering process, 5 samples of the tongue mucosa were taken with a scalpel. Per sample two paraffine sections, 5 mm thick, were cut perpendicularly to the surface, stained with hematoxylin and eosin and Löffler's methylene blue. A classification scheme for the presence of bacteria was used. The following counts (magnification 400x) were used: + = >100 bacteria per field of view, +/- = 1-100 bacteria per field of view, - = 1-10 bacteria per field (number of bacteria calculated as the mean of 4 counted fields).

The bacteriological contamination of stomach contents and scalding water was investigated. At each line control two samples of 100 ml of water were taken from the scalding tank. Stomach contents were examined from 5 pigs with an almost empty stomach and from 5 pigs with a full stomach. In both examinations mesophilic aerobic colony counts and *Enterobacteriaceae* were assessed.

## Centrifuge process

Experiments on a centrifuge process working on the same principle as found in the line studies, under standardized conditions were included in the

investigation. A centrifuge consisting of a fixed drum with a rotating bottom (RSeio 750 RPM,  $\phi$ 600 mm, content 60 l) originally designed for the cleaning of tripes was used. During rotation tongues are intensively rinsed with water. The effects of centrifugation time (20 s vs 40 s), load (10 kg vs 15 kg), and water expenditure (20 l per min. vs 40 l per min.) were assessed. Mesophilic aerobic colony counts and *Enterobacteriaceae* (per cm<sup>2</sup> tongue mucosa) were determined before and after centrifugation. A histobacterioscopic examination of tongue mucosa was carried out. The visually perceptible contamination grade of the tongues was also evaluated.

#### Statistical analysis of data

Colony counts were expressed in colony forming units (cfu) per cm<sup>2</sup> of tissue and then transformed into log values. To determine the significance of the differences between counts these were submitted to an analysis of variance. Samples with less than 7 colonies on the first decimal dilution plate and therefore inappropriate for colony counting (2) were assigned counts corresponding to the statistical limits of detection.

### RESULTS

#### Line studies

After stunning and bleeding, the muscosa of the tongues was contaminated with dirt, blood and mucus. During scalding at 62°C the surface layer of the mucosa of the tip of the tongues became detached. Contamination of the tongues with stomach contents and dirt from the machinery took place during dehairing and polishing. Damage of the tongues by the machinery also took place at these stages. The grade of the visually perceptible contamination depended on (a) fulness of the stomach (b) type of machinery (c) cleanliness of the machinery. During evisceration tongues were contaminated by contact with the apron of the worker. In most slaughter houses specific cleaning equipment for tongues in the slaughter line had hardly been developed. The cleaning effect of showering, washing, and centrifugation are summarized in Table 1. Cleanliness of the tongues at the end of the slaughter line was far from optimal at 11 of the 16 plants.

#### Bacteriological line control

The results of the bacteriological line control are presented in Fig. 1 and Tables 2, 3, and 4. Fig. 1 and Table 2 show that the highest bacteriological contamination of porcine tongues was found after bleeding. Mesophilic aerobic colony count *Enterobacteriaceae*, Gram-negative bacteria, and *Brochothrix thermosphacta* were approx. 6.1, 2.9, 3.4 and 4.1 log<sub>10</sub> N per cm<sup>2</sup> respectively. A significant decrease of colony counts was found after dehairing. After evisceration an increase in the mesophilic aerobic colony counts of tongues was found. After collection at slaughterhouse A a decrease and at slaughterhouse B an increase in colony counts were observed. The effect of the centrifuge process at slaughterhouse A was less effective than that at slaughterhouse B (especially at day 1). At day 2 at slaughterhouse A and days 1 and 2 at slaughterhouse B a significant decrease in the mesophilic aerobic colony counts and *Enterobacteriaceae* after centrifugation was observed. Following the processing stages, a decrease in *Brochothrix thermosphacta* was found. Colony counts for Gram-negative bacteria of the tongues varied, no tendency to decrease was observed. In scalding water *Enterobacteriaceae* were found to be below the detection level (<1.8). The mean value for mesophilic aerobic colony count was  $2.4 \pm 0.4$ . Histobacterioscopic examination shows that in all stages of the slaughtering process >100 bacteria (score +) per field of view of a section of the mucosa were found.

An analysis of variance of the bacteriological data (Table 3) shows that there was a significant two-way interaction between stage of the slaughtering process and sampling day for *Enterobacteriaceae* and for Gram-negative bacteria. Apart from the two-way interactions there was mostly a significant main effect caused by sampling day for *Enterobacteriaceae*, Gram-negative bacteria, and *Brochothrix thermosphacta*. Stage of the slaughtering process was the only variable which caused a highly significant one-way variance for every bacteriological parameter.

In Table 4 are presented pH values and colony counts for stomach contents of 10 pigs. Six samples contained high colony counts for *Enterobacteriaceae* (>5.8). Below a pH of 3.5 (of the stomach contents) *Enterobacteriaceae* were found to be below the detection level (<2.8).

The results of the centrifuge experiments under standardized conditions are presented in Tables 5, 6, and 7.

Table 5 shows that the centrifuge process caused a significant decrease in the mesophilic aerobic colony counts and *Enterobacteriaceae* of the tongues.

No significant differences were found between colony counts caused by different centrifugation times, loads, and water expenditures. Only the long centrifugation time (40 s) resulted in lower numbers for mesophilic aerobic colony counts (Table 6).

Table 7 shows significant two-way interactions of variance for the mesophilic aerobic colony counts but not for *Enterobacteriaceae*. All the combinations used resulted in the same visually assessed cleaning efficiency, e.g. ++ = Good.

Histobacterioscopic examination showed that dirt and adherent bacteria adhering to the surface were removed. The colonized bacteria on the mucosa were still not removed. The presence of bacteria was classified in all the cases as +. (>100 bacteria per field).

### DISCUSSION AND CONCLUSIONS

During transport of pigs to and storage at the slaughter house tongues are contaminated by the intake of dirt, faeces, and water. Pigs are slaughtered in a hanging position, as a result of which contents of the upper digestive tract, mucus, and dirt flow onto the tongues. This explains the high contamination grade of tongues after bleeding. During scalding in a tank at 62°C, thermolabile Gram-negative bacteria like *Enterobacteriaceae* are killed. (See colony counts for scalding water.) Decontamination of the tongues takes place during scalding because the mouth is open. The strong sphincter of the stomach does not allow any contents to flow towards the throat. During dehairing and polishing a strong pressure is exerted on the carcass, e.g. on the stomach, especially when it is full. The sphincter may open a little and contents of the stomach may flow towards the throat and contaminate the tongue, the machinery, and the carcass. Stomach contents may contain high numbers of bacteria and even *Enterobacteriaceae*. The increase in bacteriological contamination after evisceration is caused by the polishing machinery, stomach contents, and contact with the aprons of the workers during evisceration. The decrease in bacterial colony counts after collection at the slaughterhouse B is probably caused by the use of water during collection. At

slaughterhouse B the visual cleanliness of the tongues after centrifugation was better. This explains the differences in results between the assessed colony counts for tongues sampled at slaughterhouse A and slaughterhouse B. From the results of the standardized centrifuge process it is clear that dirt and stomach contents can be removed effectively. Independent of the combination of centrifugation time, load, and water expenditure there is a significant decrease in

transient bacteria, but not in the initial flora. These findings are substantiated by the histobacterioscopic findings. From the literature it is well known that colonized bacteria are strongly attached to the tissue (4).

Nevertheless, experiments with veal tongues showed that it is possible to remove colonized bacteria from the mucosa by the use of 2% v/v lactic-acid dilution during centrifugation (6). Considering economic as well as hygienic aspects, a time, load, water expenditure ratio of 20 s/15 kg/20 l per min. is preferable.

Finally, it can be concluded that GMP concerning BQA of porcine tongues must include:

- (a) slaughtering of pigs with an empty stomach only.
- (b) cleanliness of the machinery (easy to clean and disinfect).
- (c) avoiding contamination during evisceration.
- (d) application of a cleaning process to the tongues, e.g. a centrifuge process.

#### LITERATURE

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#### ACKNOWLEDGEMENTS

This investigation was supported by the Veterinary Public Health Inspectorate, Rijswijk, The Netherlands. The authors wish to thank Mr. W.J. Kruger for experimental assistance.

B0081/HYG7

Fig 1. Mesophilic aerobic colony count (in  $\text{Log}_{10} \text{N/cm}^2$  and standard deviation) of porcine tongues at different stages of the slaughtering process.

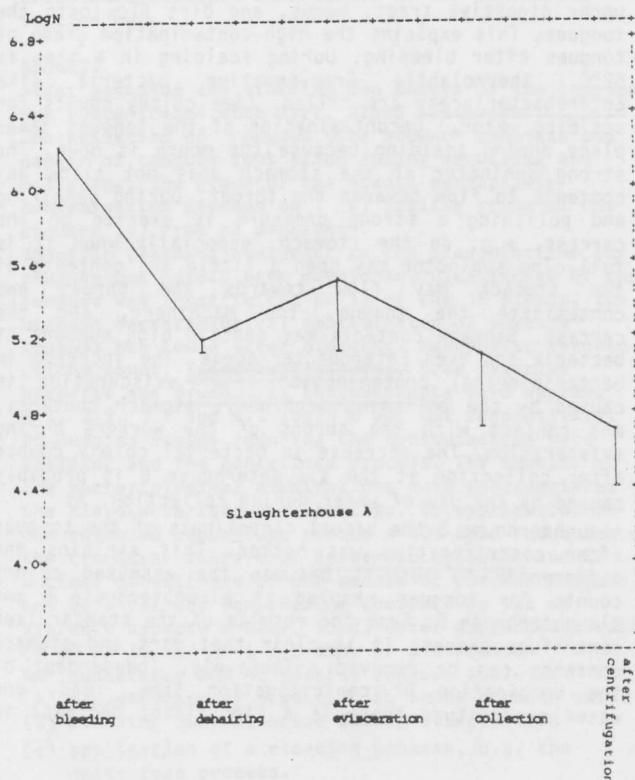


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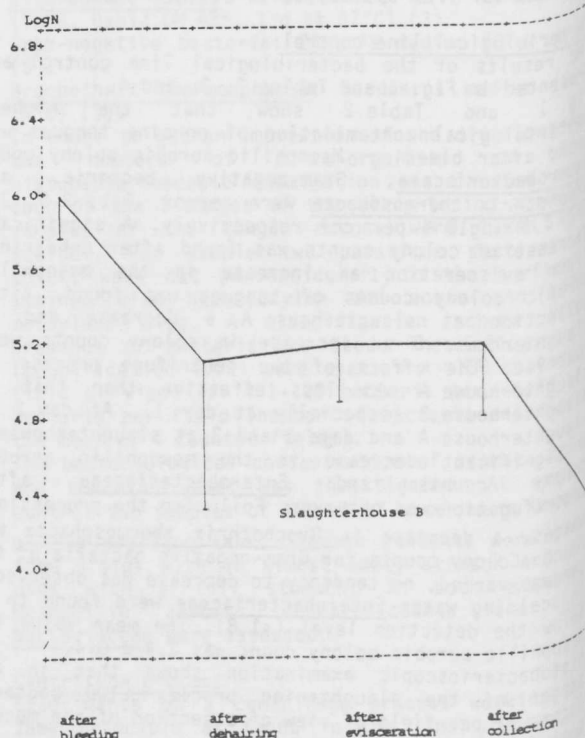




TABLE 1. Visually perceptible cleaning effects on the contamination grade of porcine tongues at 16 slaughterhouses.

|    | Processing |         |                | Total result |
|----|------------|---------|----------------|--------------|
|    | Showering  | Washing | Centrifugation |              |
| 1  |            |         |                |              |
| 2  | --         | -       |                | -            |
| 3  |            |         | +              | +            |
| 4  |            | -       | -              | -            |
| 5  |            |         | +              | +            |
| 6  |            |         |                | --           |
| 7  |            | -       |                | -            |
| 8  |            |         | ++             | ++           |
| 9  |            |         | -              | -            |
| 10 | --         |         |                | --           |
| 11 |            |         | +              | +            |
| 12 |            | --      |                | --           |
| 13 | -          |         | -              | -            |
| 14 | -          |         |                | -            |
| 15 |            | --      |                | --           |
| 16 | +          |         |                | +            |

++ = good  
+ = sufficient  
- = insufficient  
-- = bad

TABLE 4. Colony counts (in  $\text{Log}_{10} \text{Ng}^{-1}$ ) of stomach contents of 10 pigs in the slaughterline.

| Pig* | pH  | Mesophilic aerobic colony count | Enterobacteriaceae |
|------|-----|---------------------------------|--------------------|
| 1    | 5.6 | 7.7                             | 7.3                |
| 2    | 5.4 | 8.0                             | 7.5                |
| 3    | 3.9 | 7.4                             | 7.3                |
| 4    | 3.3 | 5.9                             | <2.8               |
| 5    | 2.6 | 7.0                             | <2.8               |
| 6    | 6.9 | 7.1                             | 5.8                |
| 7    | 6.7 | 7.6                             | 6.9                |
| 8    | 5.5 | 7.4                             | 6.9                |
| 9    | 3.5 | 4.9                             | <2.8               |
| 10   | 2.4 | 4.8                             | <2.8               |

\*Pig 1 - 5 : full stomach  
6 - 10 : almost empty stomach

TABLE 5. Effect of the centrifuge process on bacterial counts (in  $\text{Log}_{10} \text{N}$  per  $\text{cm}^2$  and standard deviation) of porcine tongues.

|                     | N  | Mesophilic aerobic colony count | Enterobacteriaceae |
|---------------------|----|---------------------------------|--------------------|
| unprocessed tongues | 20 | $5.5 \pm 0.4$                   | $3.8 \pm 0.5$      |
| processed tongues   | 80 | $4.5 \pm 0.5$                   | $2.0 \pm 0.5$      |
| significance        |    | $p < 0.0001$                    | $p < 0.0001$       |

The statistical procedure used was an analysis of variance.

TABLE 2. Bacterial counts (in  $\text{Log}_{10} \text{N/cm}^2$  and standard deviation) of porcine tongues at different stages of the slaughtering process.

| slaughter house | Day | STAGE                | N  | Mesophilic aerobic colony count | Enterobacteriaceae | Gramneg. bacteria | Brochothrix thermosphacta |
|-----------------|-----|----------------------|----|---------------------------------|--------------------|-------------------|---------------------------|
|                 |     |                      |    |                                 |                    |                   |                           |
| A               | 1   | After bleeding       | 10 | $6.1 \pm 0.4$                   | $2.9 \pm 0.3$      | $4.0 \pm 0.6$     | $4.2 \pm 0.2$             |
|                 |     | After evisceration   | 10 | $5.5^b \pm 0.4$                 | $2.3^d \pm 0.4$    | $3.1^d \pm 0.4$   | $3.1^d \pm 0.8$           |
|                 |     | After collection     | 10 | $5.0^a \pm 0.4$                 | $2.3 \pm 0.5$      | $3.0 \pm 0.3$     | $2.9 \pm 0.3$             |
|                 |     | After centrifugation | 10 | $4.7 \pm 0.3$                   | $2.3 \pm 0.3$      | $3.9^c \pm 0.7$   | $2.9 \pm 0.5$             |
| A               | 2   | After bleeding       | 10 | $6.4 \pm 0.3$                   | $3.0 \pm 0.7$      | $2.9 \pm 0.2$     | $4.1 \pm 0.5$             |
|                 |     | After dehairing      | 4  | $5.1^d \pm 0.2$                 | $2.0^a \pm 0.3$    | $2.9 \pm 0.4$     | $2.9^c \pm 0.5$           |
|                 |     | After collection     | 10 | $5.2 \pm 0.4$                   | $2.6^a \pm 0.5$    | $3.0 \pm 0.3$     | $2.7 \pm 0.2$             |
|                 |     | After centrifugation | 10 | $4.7^b \pm 0.3$                 | $<1.6^d$           | $3.2 \pm 0.3$     | $<2.6$                    |
| B               | 1   | After bleeding       | 10 | $6.0 \pm 0.2$                   | $3.0 \pm 0.3$      | $3.2 \pm 0.3$     | $4.3 \pm 0.5$             |
|                 |     | After dehairing      | 10 | $4.8^d \pm 0.3$                 | $2.4^c \pm 0.4$    | $<2.6^d$          | $<2.6^d$                  |
|                 |     | After collection     | 10 | $5.2 \pm 0.4$                   | $3.2^d \pm 1.0$    | $3.8^d \pm 0.7$   | $3.2^d \pm 0.4$           |
|                 |     | After centrifugation | 10 | $4.4^d \pm 0.3$                 | $<1.6^d$           | $3.8 \pm 0.3$     | $2.7^d$                   |
| B               | 2   | After bleeding       | 10 | $6.0 \pm 0.2$                   | $2.7 \pm 0.3$      | $3.6 \pm 0.3$     | $3.9 \pm 0.6$             |
|                 |     | After evisceration   | 10 | $5.2^d \pm 0.3$                 | $2.5 \pm 0.6$      | $3.2^a \pm 0.5$   | $2.8^d \pm 0.3$           |
|                 |     | After collection     | 10 | $5.3 \pm 0.4$                   | $2.7 \pm 0.7$      | $3.6 \pm 0.5$     | $2.8 \pm 0.4$             |
|                 |     | After centrifugation | 10 | $4.0^d \pm 0.5$                 | $<1.6^d$           | $2.9^c \pm 0.4$   | $<2.6$                    |

Colony counts at each stage are only compared with colony counts at the preceding stage. Statistics were carried out with analysis of variance.

a =  $p < 0.05$       b =  $p < 0.01$   
c =  $p < 0.005$       d =  $p < 0.001$

TABLE 3. Significance in variance analysis of: Mesophilic aerobic colony counts, Enterobacteriaceae, Gram negative bacteria and Brochothrix thermosphacta of porcine tongues at two slaughterhouses, five stages of the slaughtering-process and two sampling days.

| Slaughterh.<br>two-way var.<br>stage / day | DF | Mesophilic aerobic<br>colony count |         | Enterobacteriaceae |         | Gramnegative bacteria |         | Brochothrix therm. |         |
|--------------------------------------------|----|------------------------------------|---------|--------------------|---------|-----------------------|---------|--------------------|---------|
|                                            |    | 1                                  | 2       | 1                  | 2       | 1                     | 2       | 1                  | 2       |
|                                            | 2  | ns                                 | ns      | p<0.001            | ns      | p<0.005               | p<0.001 | ns                 | ns      |
| one-way var.<br>stage<br>day               | 4  | p<0.001                            | p<0.001 | p<0.001            | p<0.001 | p<0.005               | p<0.001 | p<0.001            | p<0.001 |
|                                            | 1  | ns                                 | ns      | ns                 | p<0.05  | p<0.001               | p<0.05  | ns                 | p<0.01  |

DF = Degrees of Freedom  
ns = not significant

TABLE 6. Effect of (a) water expenditure (b) centrifugation time (c) load in the centrifuge process on bacterial counts (log N per cm and standard deviation) of porcine tongues.

|         | N  | Mesophilic aerobic<br>colony count | Enterobacteriaceae |
|---------|----|------------------------------------|--------------------|
| a 20 l  | 40 | 4.4 ± 0.4                          | 1.9 ± 0.4          |
| 40 l    | 40 | 4.6 ± 0.6                          | 2.2 ± 0.6          |
| sign.   |    | N.S.                               | N.S.               |
| b 20s   | 40 | 4.6 ± 0.6                          | 2.1 ± 0.5          |
| 40s     | 40 | 4.4 ± 0.5                          | 2.0 ± 0.5          |
| sign.   |    | p<0.05                             | N.S.               |
| c 10 kg | 40 | 4.5 ± 0.6                          | 2.1 ± 0.6          |
| 15 kg   | 40 | 4.5 ± 0.5                          | 2.0 ± 0.5          |
| sign.   |    | N.S.                               | N.S.               |

N.S. = not significant

The statistical procedure used was an analysis of variance.

TABLE 7. Two- and three-way interaction in an analysis of variance on water expenditure, centrifugation time, and load on bacterial counts of porcine tongues in the centrifuge process.

| Interactions of the variables                  | Mesophilic aerobic<br>colony count | Enterobact |
|------------------------------------------------|------------------------------------|------------|
| Waterexpenditure-centrifuga-<br>tion time-load | N.S.                               | N.S.       |
| Waterexpenditure-load                          | p<0.05                             | N.S.       |
| centrifugation time-load                       | p<0.01                             | N.S.       |
| Waterexpenditure-centrifuga-<br>tion time      | p<0.01                             | N.S.       |

N.S. = Not Significant