OCHRATOXIN A: TOXICOLOGICAL ASSESSMENT OF RESIDUES IN MEATS

JOHANNA FINK-GREMMELS and LOTHAR LEISTNER

Federal Centre for Meat Research, E.-C.-Baumann-Str. 20, D-8650 Kulmbach

Ochratoxin A (OA) a dihydrocoumarin derivative linked to phenylalanin by an amino bond is a secondary metabolite produced by various species of the genera Penicillium and Aspergillus. As a result OA is found frequently in a number of cereal crops and related foods (for review see HARWIG et al., 1983). Under the German agricultural and environmental conditions OA is found predominantly in wheat and barley in concentrations ranging from 0.1-200 µg/kg, the mean concentration beeing 30 µg/kg feed (THALMANN and GRUBER, 1981; GEDEK, 1985; BAUER et al., 1986). The frequency of positive samples exceeds 10% for grains and formulated feeds and varies according to seasonal and regional factors. Cereals and crops for human consumption have, as of yet, not been monitored intensively. Recently JIAO et al. (1987) who investigated various oat, rey and barley meals as well as flours demonstrated that 50% of the products contained 0.1-0.3 µg/kg and 16.6% of the samples were contaminated with OA in concentrations ranging between 0.3 and 9.8 µg/kg.

After consumption of contaminated feedstuffs OA can be found in various edible tissues of pigs and poultry. Residues have been detected in blood serum, kidneys, liver, muscle and fat. Carcasses of ruminants (with the exception of preruminant veal calves) are unlikely to be contaminated as the microbial flora of the forestomach systeme degradates OA to less toxic metabolites capacitatively. Excretion of OA with eggs has been demonstrated in laying hens under experimental conditions (BAUER et al., 1988).

In Germany the occurence of OA has been monitored in plasma and kidneys of pigs. The residual concentrations

in kidneys varied between 0.5 and 10.0 pa/kg, the main interest of the part of pg/kg, the majority of the samples range between 1.5 and paller ging between 1-2 µg/kg tissue (BANK) al., 1984: SCHELLE al., 1984; SCHEUER et al., 1984). of the test 50% of the tested serum samples of pigs were contaminated with OA in concentrations between 0.1-67.3 µg/kg (BAUFR et al. 1200) (BAUER et al., 1984). Consequently sages which were produced with pig ma (frankfurter ma (frankfurter-type sausages) were contaminated as well, indicating the high stability of the toxin during processing of meats sing of meats. In addition is it notice that the procession that the processing resulted not in a decrease of Change resulted not in a decrease of OA concentrations as the residual amount residual amount detected in sausages exceeded the mean concentrations plasma same plasma samples two fold (SCHEUER, 1988) 1988).

Thus, OA has to be considered as a contaminant of foods of animal origin following the factors following the food chain as well as contaminant contaminant of food of plant origin. However, it can not be estimated, of yet, what of yet, what are the food commodified which are predamined which are predominantly responsible for human for human exposure. Human exposure can be assessed to can be assessed from two independent studies demonst studies demonstrating that OA present has to be expected. has to be expected in blood samples of more than 50% of all consumers ER et al. 1997 ER et al., 1986, SCHEUER and LEISTN 1986). In addition 1986). In addition OA was found in renal specimen renal specimen derived from hospital patients (RALIED patients (BAUER et al., 1986). Like in monogastric animals OA may created with partial of the control of the co created with mothers milk resulting in an exposure of the neonatus necessary results. et al., 1987). Therefore, it seems necessary to evaluate public health aspects of OA residues on the latest of available to the seems of the seems of available to the seems of available to the seems of the neonatus (GAPC). of OA residues on the basis of available toxicological and line

### TARGET ORGAN TOXICITY

OA is a known nephrotoxin, inducing renal tubular necrosis, hepatic and phoid necrosis phoid necrosis and a dessiminated vascular coagulation (KROGH, 1978; MEISNER and KROCK) 1986; ALBASSAM vascular coagulation like syndrome (KROGH 1970 1986; ALBASSAM et al., 1987).

Subacute and subchronic toxicity and in rats, guinea nice in rats, guinea pigs, beagle dogs pigs demonstrated pigs demonstrated that renal in low necrosis can be observed even in low dosages. Subst dosages. Subchronic administration

of OA (0.145 µg/kg b.w.) caused increatubular in Hubular enzymes suggesting tubular injury RANE et al., 1986). The dose dependent pathologic alterations can be estimated from a 90 day feeding experiment in vely 0.01 mg/(s.p. w.) resulted in minimum. Vely 0.01 mg/kg b.w.) resulted in minimal Pathologic lesions in the kidneys and the tad as the t therefore could be accepted as the threshhold level of toxicity (MUNRO et al.,

ER E

kg sour

oces

d he

5

gin

05

ties

re.

ent. ently

STNE STNE

al

ex"

REIS REIS

eces

GHI

1[

## EMBRYOTOXICITY AND REPRODUCTI-

OA is a potent teratogen in experimental dosages and causes malformations in mice, rats, hamsters and cockerels (HAYES, 1978). Additional studies demonstrated that the late implantation losses during g sunney in rats were correlated with Gupk A sion of ovarian steroidogenesis (GUPKA et al., 1981). In vitro experiments with short term cultured interstitial with short term cultured in collection cells from gerbil testes gave evidence that dence that testosterone synthesis might be affect. Testosterone synthesis might be affected as well (FENSKE and FINK-GREMMELS, 1989).

We the aim total a set of experiments with the aim to establish a teratological noeffect-level. Reversible minor malformations (signal Reversible minor of tions (signs of delayed ossification of skeleton bones and wavy rips) could be observed and wavy rips of 0.68 red at a mininum oral dose of 0.68 mg/kg b.w.. Fetal body weight was affected at a dose of 0.17 mg/kg the gestar. OA was applied to dams during the When OA was applied to dams do. However III period (THEIN et al., 1988). However, the most sensitive test para-Meter in these studies was a significant which coul increase in placental weight, which could be observed placental weight, which served be observed by the served placental weight, which could be observed by the served placental weight. be observed at a dosage of 0.087 mg/kg of OA. The analysis of the accessment of OA. Therefore, in risk assessment hg/kg h ogical no-effect-level of 0.17 ng/kg b.w. corresponding to a feed concentration of 1.25 mg/kg can be Occepted. These findings are comparable previous (BROWN with previously published results (BROWN

# IMMUNOTOXICITY

The specific mode of action of OA is inhihit: an inhibition of the phenylalanine-tRNAsynthetase, resulting in an inhibition

of the elongation step in protein synthesis. Therefore, a reduced immunoglobuline synthesis has to be considered (CREPPY et al., 1984). This hypothesis is supported by pathohistological findings demonstrating necrotic lesions in various lymphoid tissues as well as by several in vitro tests (RICHARD et al., 1975, PRIOR and SISODIA, 1979; KLINKERT et al., 1981 and LUSTER et al., 1987).

Again, with the aim to establish a noeffect-level we performed a series of lymphocyte transformation tests, as an experimental model to assess altered cellular immune response under the influence of mycotoxins. This test can be developed with peripheral mononuclear cells from animals, which have been exposed in vivo to OA as well as for in vitro exposed cells. After oral administration to rats of 0.1, 0.5 and 1.0 mg OA/kg b.w. an inhibition of the proliferation rate of mitogen stimulated lymphocytes could be observed only in the highest dosage group (JAHN and FINK-GREM-MELS, 1988). This dosage resulted in a mean cumulative serum concentration of 478 µg/l in the exposed rats. In addition, in vitro-tests with human derived peripheral blood lymphocytes indicated a mean effective concentration of 1108 µg/l cell suspension (v. GERNLER, JAHN and FINK-GREMMELS, 1989). When these data are compared to the OA concentrations which had been detected in human blood samples (0.1-0.9 µg/l) the broad safety margin between measurable effects in the lymphocyte transformation test and the natural occuring OA concentration is evident.

#### CARCINOGENICITY

Although previous studies failed to demonstrate a carcinogenic potential of OA in long term feeding experiments in rats (PURCHASE and van der WATT, 1971) and mutagenicity studies with bacterial test systems as well as genotoxicity tests in cell cultures were inconsistent (MORI et al., 1984; BENDELE et al., 1985a), recent animal experiments with mice and rats could clearly demonstrate an OA induced carcinogenesis (KANISAWA and SUZUKI, 1978; BENDE-LE et al., 1985b, BOORMAN, 1988).

#### TOXICOKINETICS

From several studies which have been conducted with different dosages and in various animal species it is evident, that absorption, distribution and elimination of OA are influenced by the route of administration and varies considerable among different animal species. Recently, HAGELBERG et al. (1989) compared the available data for OA. From this study it can be concluded that pigs and rats have related values for the plasma elimination rate (biological half-life of the toxin) with 150 and 170 hrs., respectively. However, in monkeys the OA half life is 510 hrs after oral administration of a similar dosage. The binding abilities of OA to plasma proteins were investigated as well, and in all species (including man) protein binding exceeded 99.8%. These kinetic investigations may help to interpret the data obtained from human blood sample surveys, as it remained so far unexplained why OA was found with a high incidence but in small, consistent amounts in man.

#### RISK ASSESSMENT

Mycotoxins are considered as environmental pollutants. An approach to assess the health risks to humans from the presence of OA residues in meats and other foods has to consider toxicological as well as kinetic data. In extrapolating to humans, the no-effect-level (NOEL) obtained from various animal studies together with safety factors have to be considered, the latter accounting interspecies and intraspecies differences as well as problems regarding from the suitability of the experimental models used. From subchronic experiments in rats (MUNRO et al., 1974) a NOEL of 0.01 mg/kg b.w. can be estimated for target organ toxicity. In extrapolation to humans the significance of the NOEL, a safety factor of at least 100 has to be applied, resulting in a tolerance level of 0.10 µg/ kg b.w.. This exposure is comparable to the dosage which caused in subchronic feeding experiments in rats biochemically detectable, but reversible renal tubular effects (KANE et al., 1986).

For an assessment of the significance of embryotoxic and reproductive effects it is important that so far in all tested

animal species the response to OA expo sure during the gestation period resulted in dose dependent in dose dependent major or minor mal formations. For the extrapolation of the significance of significance of these animal experiments a safety factor of 500 should be used. Thus, for major malformations a toleral level of 0.0014 level of 0.0014 mg/kg b.w. can be calculated. However, as a result of our studies and using the same safety factor an exposure of 0.17 µg/kg b.w. seems with a higher probability to indicate tolerance level. tolerance level. Thus, both treshold levels for dose-dependent OA effects are in the same range and the safe intake estimated from the mated from these data is 0.10 µg/kg b.w./day.

The assessment of data on immunotoxicity is difficult. As mentioned above there seems to be a satisfying safety margin between the concentrations in human blood samples and the concentrations which cause measurable effects in-vitro experiments with human derived lymphocytes.

However, experimental immunosuppression ve effects are ve effects are considered to be an ifical cator for carcinogenicity. The significant of tumorigenicity data in laboratory animals to man are discussed controversially. If a consially. If a compound is avoidable and its usage is in its usage is intended, as for example for hydionic for hygienic or technical purposes in food production, carcinogens are considered as genetation dered as genotoxic, having no theshold level. Therefore level. Therefore, an estimation of an acceptable della acceptable daily intake or a virtual safe dose is not accept dose is not accepted in food toxicology (FÜLGRAFE 1992) (FÜLGRAFF, 1989). Under this premise an environmental an environmental pollutant like a mycor toxin will be room. toxin will be regulated with the by to avoid any detectable residue means of the most sensitive method for food analysis.

### RESIDUES IN CEREALS VS RESIDUES IN MEATS

From the limited survey data available on the natural occurence of OA in commodities it can be estimated cereals and other plant foods are minated with a higher incidence and in higher concentrations than foods an animal origin. However, for humans the different absorption rate of OA from the

gated. Thus, as of yet, not been investigated. Therefore, it is impossible to discuss the relevance of protein bound residues in comdues in meat and meat products in com-Parison with OA exposure form plant maletials. Additional studies on the bioavai-Ote here the signifiare hecessary to elucidate the significonce of the various food commodities in the OA food chain.

0

he.

0

551-

0

# CONCLUSIONS AND RECOMMENDA-

Based on the toxicological date OA has to be considered an undesirable compound in foods for the toxicological date UA 1100 in foods for human consumption. Further studies are recommended to assess what tood sources such as cereals or meats Ote predominantly responsible for human the exposure. It is likely that for man the biogvailability for OA residues is higher in Cereals than in meats, as in the latter OA is bounded to proteins.

that port monitoring data indicate that pork contributes significantly to human OA exposure, guideline levels hay need to be established. However, as geds produced for on-farm usage are difficult to monitore, a screening method to monitore, a screening mental the control of OA residues in pig carcontrol of UA rest

From the toxicokinetic data it can be assisted the state of the state Stimated that serum analysis will give Sufficient information about OA contahindian of carcasses. Further studies ore necessary to define a plasma threshold level in pigs which does not result any detectable residues in the carcass.

## REFERENCES

Albassam, M.A., Young, S.I., Bhatnagar, Histopathological Alectron microscopi Studies on the contraction of ochrastudies on the acute toxicity of ochra-Bouer Net Pathol. 24: 427-435.

Bouler, J., Gareis, M. & Gedek, B. (1984): Jun Nachweis und Vorkommen von Ochratoxin A bei Schlachtschweinen.

Mschr. 97: 27 Berlachtschweinen.
A bei Schlachtschweinen.
A bei Schlachtschweinen.
A bei Schlachtschweinen.
B 283. Münch. Tierärztl. Wschr. 97: 279-

Bauer, J., Gareis, M. & Gedek, B. (1986): Incidence of ochratoxin A in blood serum World Consort man and animals. Proc. World Congress Foodborne Infections,

Berlin, pp. 907-910.

Bauer, J., Niemiec, J. & Scholtyssek, S. (1986):

Ochratoxin A in Legehennenfutter. 2. Mitteilung: Rückstände im Serum, Leber und Ei. Arch. Geflügelkd. 52: 71-75.

Bendele, A.M., Neal, S.B., Oberley, T.J., Thompson, C.Z., Bewsey, J.B., Hill, L.E., Carlton, W.W. & Probst, G.S. (1985a): Evaluation of ochratoxin A for mutagenicity in a battery of bacterial and mammalian cell assays. Fd. Chem. Toxic. 23: 911-918.

Bendele, A.M., Carlton, W.W., Krogh, P. & Lillehoj, E.B. (1985b): Ochratoxin A carcinogenesis in the (C57BL/65xC<sub>3</sub>H)F<sub>1</sub>mouse. J. Natl. Canc. Inst. 75: 733-742.

Boorman, G. (1988):

NTP technical report on the toxicology and carcinogenesis studies of ochratoxin A in F 344/N rats (Gavage studies). NTP TR 358. NTP Publication No. 88-2813. National Toxicology Program, U.S. Department of Health and Human Services.

Brown, M.H., Szczech, G.M. & Purmalis, B.P. (1976):

Teratogenic and toxic effects of ochratoxin A in rats. Toxicol. Appl. Pharmacol. 37: 331-338.

Creppy, E.E., Röschenthaler, T. & Dirheimer, G., (1984): Inhibition of protein-synthesis in mice by ochratoxin A and its prevention by phenylalanine. Fd. Chem. Toxicol. 22: 883-886.

Fenske, M. & Fink-Gremmels, J. (1989): Effects of fungal metabolites on testosterone secretion in vitro. Arch. Toxicol., submitted.

Fülgraff, G. (1989): Lebensmitteltoxikologie. Verlag Eugen Ulmer, Stuttgart, pp. 13-48.

Gareis, M., Märtlbauer, E., Bauer, J. & Gedek, B. (1987): Determination of ochratoxin A in human milk. Z. Lebensm. Unters. Forsch. 186: 114-117.

Gedek, B. (1985):

Toxine, insbesondere Mykotoxine, in Futtermitteln. Dtsch. tierärztl. Wschr. 92: 215-218.

Gernler, v. J.R., Jahn, M. & Fink-Gremmels, J. (1989): personal communication.

Gupka, M., Bandyopadyay, S., Paul, B. & Mazumdlar, S.K. (1981):

Ovarian steroidogenesis and development of fetuses following ochratoxin A treatment in pregnant rats. Endocrinology 77: 152-160.

Hagelberg, S., Hult, K. & Fuchs, R. (1989): Toxicokinetics of ochratoxin A in several species and its plasmabinding properties. J. Appl. Toxicol. 9: in print.

Harwig, J., Kuiper-Goodmann, T. & Scott, P.M. (1983):

Microbial food toxicants: ochratoxins. CRC Handbook of Foodborne Diseases of Biological Origin. Recheigl, M. (ed). CRC Press, pp. 193-238.

Hayes, A.W. (1978): Mycotoxin teratogenicity. Toxicon. 1: 739.

Jahn, M. & Fink-Gremmels, J. (1988): Wirkung von Ochratoxin A im Lymphozytentransformationstest. Mitteilungsblatt der Bundesanstalt für Fleischforschung, Kulmbach, Nr. 101, S. 8064-8068.

Jiao, Y., Mehlitz, I. & Weber, R. (1987): Ochratoxin A in Cearalien des Berliner Marktes. BGA-Tätigkeitsbericht 1987.

Kanisawa, M. & Suzuki, S. (1978): Induction of renal and hepatic tumors in mice by ochratoxin A. Gann 69: 599-600.

Kane, A., Creppy, E.E., Röschenthaler, R. & Dirheimer, G. (1986):

Changes in urinary and renal tubular enzymes caused by subchronic administration of ochratoxin A in rats. Toxicology 42: 233-243.

Klinkert, W., Lorkowski, G., Creppy, E.E., Dirheimer, G & Röschenthaler, R. (1981): Inhibition of macrophage migration by ochratoxin A and citrinin, and prevention by phenylalanine of the ochratoxin A-induced inhibition. Toxicol. Europ. Res. 3: 185-189.

Krogh, P. (1978):

Causal associations of mycotoxic nephropathy. Acta Pathol. Microbiol Scand. 269:

Luster, M.I., Germolec, D.R., Burleson, G., Jameson, C.W., Ackermann, M.F., Lamm, K.R. & Hayes, H.T. (1987):

Selective immunosuppression in mice of natural Lill of natural killer cell activity by ochrotoxin A Carre toxin A. Cancer Res. 47: 2259-2263,

Meisner, H. & Krogh, P. (1986): Phosphoenolpyruvate carboxykinase selective indi selective indicator of ochratoxin and ced nephronation ced nephropathy. Development. Environment. Sci. 14: 199-206.

Mori, H., Kawei, K., Ohbayashi, F., mazaki, M., Hamasaki, T. & Williams G.M. (1984).

Genotoxicity of a variety of mycotoxic in the hengican in the hepatocyte primary culture repair test using rat and mouse her tes. Cancer De tes. Cancer Res. 44: 2918-2923.

Munro, I.C., Moodie, C.A., Kuiper God man, T. Scott Die, C.A., Kuiper H.C. man, T., Scott, P.M. and Grice, H.C. (1974):

Toxicologic changes in rats fed graded dietary levels of dietary levels of ochratoxin A. Appl. Pharman Appl. Pharmacol. 28: 180-188.

Prior, M.G. & Sisodia, C.S. (1979): Some tissue culture and immunological studies with achieves the studies with achieve the studies with achieves the studies with a st studies with ochratoxin A. Abstr. 6th Int. Symp. As: Int. Symp. Anim., Plant and Microbiol. Toxins, Uppsala, Toxicon. 1: 146.

Purchase, I. & Watt, J. van der (1971)
The long-term The long-term toxicity of ochratoxin in rats. Ed. C. in rats. Fd. Cosmet. Toxicol. 9: 681.

Richard, J.L., Thurston, J.R., Deyoe, B.L. & Booth Effect of ochratoxin and aflatoxin of serum protein serum proteins, complement activity antibody prod antibody productions to Brucella abolin guinea pigs. in guinea pigs. Appl. Microbiol.

Scheuer, R., Bernhard, K. & Leistness L. (1984):

Rückstände von Ochratoxin A in Schill nenieren. Mittail nenieren. Mitteilungsblatt der Bundeshistalt für Fleisch stalt für Fleischforschung, Kulmbach, Nr. 83, S. 5781

Scheuer, R. & Leistner, L. (1986): Nachweis von California Nachweis von Ochratoxin A in Humseller proben aus der D proben aus der Bundesrepublik land. Jahresberich land. Jahresbericht Bundesanstalt Fleischforschussen Fleischforschung, Kulmbach C 21,

Untersuchungen zum Vorkommen von Ochratoxin Ochratoxin A. Mitteilungsblatt der desanstalt fix desanstalt für Fleischforschung, bach, Nr. 102 bach, Nr. 102, S. 8235-8240.

Thalmann, A. & Gruber, S. (1981): Unitersuchungen über das Vorkommen von Mykotovin eine Vorkommen von Mykotoxinen in Futtermitteln. Forschungsbericht 78HS12,17.

ice chro

toxins pato

aded xicol.

gical 6th biol

971): ×in A 31-681

oei

on of ty to borton

chwe deson chi

nanbl

on Bun

Thein, O., Trapper, D. & Fink-Gremmels, J. (1988):

Thein, O., Trapper, D. embryotoxischen Wirkung von Constalt A. Mitteilungsblatt der Bundesan-A. Mitteilungsblatt der Bundellungsblatt der Bundel Nr. 101, S. 8068-8073.