

Evolution of the concentration of trehalose secreted
(ppb) from the moment of the administration to the

METABOLIC TYPE OF MUSCLES AND MEAT QUALITY IN STRESS-SENSITIVE
AND STRESS-RESISTANT PIGS
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SUMMARY: The experiment has been conducted on 20 male Camborough pigs. For dividing the animals in both stress-sensitive and stress-resistant, Halotan-test was used. Pigs have been fattened up to 100 kg of live body. Contents of enzymes and metabolites have been determined and physico-chemical analysis of muscles Longissimus dorsi /LD/, Semimembranosus /SM/ and Rectus abdominis /RA/ performed. Results obtained concerning glycolytic activity of muscles show that it is not influenced by stress-sensitivity of animals. Glycolytic potential is associated with metabolic type being higher in glycolytic muscles. In stress-sensitive animals glycolytic potential level is increased. Quality characteristics characterizing PSE-syndrome in stress-sensitive pigs are related to muscle type being best expressed in glycolytic type of muscles.

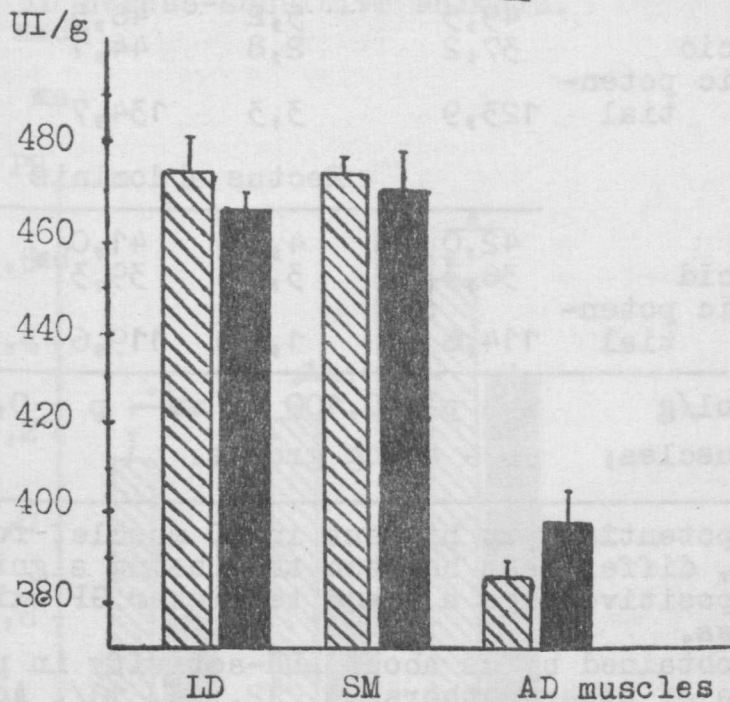
INTRODUCTION: Many pigs/in number/of commercial form derived from two-line hybrid Camborough for meat producing increased the interest to investigations on both stress-sensitivity and quality of production obtained. Spread of stress-syndrome in that population is 13 to 20 % /1, 2/. Studies on physico-chemical composition of meat are numerous and mainly in genetic aspect /5, 6, 7, 9, 10, 16, 17, 22/. Data are less about enzymatic activities of muscles which would indicate capacity of both glycolysis and mitochondrial oxidation /18, 24, 25/. The aim of study was to establish the level of metabolic processes and glycolytic ability of three muscles post mortem regarding the meat quality in both stress-resistant and stress-sensitive pigs.

MATERIAL AND METHOD: Trial was conducted on 20 male Camborough pigs. For separating the animals in stress-resistant and stress-sensitive, Halotan-test has been used. Final live weight was 100 kg. Samples of muscles Longissimus dorsi /LD/, Semimembranosus /SM/ and Rectus abdominis /RA/ have been taken 45' min.p.m. for determining enzymes and metabolites. Activity of lactat dehydrogenesis /LDH/ was determined after Ansay's method /1974/, contents of glycogen and lactic acid according to Bergmeyer's method /1974/. For determining the glycolytic potential /GP/ formula recommended by Monin et al. /1985/ was used. Samples for physico-chemical analysis from the same muscles have been taken 24 h p.m. Results obtained

were treated through one-factor variance analysis.

RESULTS AND DISCUSSION: Enzymes and metabolites.

In Fig.1 is given LDH-activity in all the three studied muscles for both groups of animals. It is highest in m.LD, being even lower in m.SM it is close to that in m.LD. Significantly lower are the values for that characteristic in RA muscle, differences between muscles being significant / $p < 0,05$ /. A tendency to higher LDH-activity is observed in stress-sensitive pigs. Glycogen content in a descending series is: m.LD, m.SM, m.RA. Differences between m.LD and m.SM are given /table 1/. Stress-sensitivity has not influenced that characteristic. Quantity of lactic acid is practically the same in m.LA and m.SM, and lower in m.RA. An effect of positive animals was established, for m.LD and m.SM it is proven at a higher significance degree / $p < 0,05$ / compared to m.RA / $p < 0,100$ /.



▨ stress-resistant, I group ■ stress-sensitive, II group

Fig.1. Values for LDH-activity in LD, SM, & RA muscles

Table 1. Metabolites and glycolitic potential in LD, SM and RA muscles.

Characteristics	Groups		II	
	\bar{X}	$\pm \sigma$	\bar{x}	$\pm \sigma$
Longissimus dorsi				
1. Glycogen	67,5 ^a	6,1	60,0 ^b	4,9
2. Lactic acid	37,5	3,2	44,8 ^b	3,0
3. Glycolitic potential	142,5	5,2	149,6 ^a	4,2
Semimembranosus				
1. Glycogen	49,5	5,2	46,5 ^b	5,5
2. Lactic acid	37,2	2,8	44,1 ^b	3,1
3. Glycolitic potential	123,9	3,3	134,7 ^a	3,5
Rectus abdominis				
1. Glycogen	42,0	4,5	41,0 ^b	3,7
2. Lactic acid	36,3	3,5	39,3 ^b	1,9
3. Glycolitic potential	114,6	1,2	119,6	1,1
1, 2 & 3 mmol/g	* - $p < 0,100$; ** - $p < 0,025$			
a - among muscles;	b - among groups.			

Glycolitic potential is highest in LD muscle, followed by m. SM and m. RA, differences between them being significant/ $p < 0,100$ /. In positive pigs a trend to higher GP exists in all three muscles.

Results obtained by us about LDH-activity in pigs are one-way to those of other authors /5, 12, 15, 16/. According to Beshar /1986/ m.LD and m.SM show a fast glycolysis. These are muscles containing relatively high part of white or anaerobic metabolic fibres. Results on metabolit content in muscles studied by us are one-way ones to those of Monin et al./1986/, according to which Halotan-sensitivity influences in a less extent on them compared to muscle type. Tendency established by us toward a lower glycogen content in Halotan-positive animals shows that glycolysis post mortem in them has taken faster. In this respect results of Tarrant et al., /1972/ on a

muscle sample taken immediately post mortem show that lactat level in white muscles is 80,4 mol/g the red ones 120,4 mol/g, i.e. extent of anaerobic glycolysis is nearly equal. After death glycolysis is more active in glycolitic muscles and pH-value is associated with formation of lactat.

Meat quality. pH_1 -values /45' post mortem/ are given in Fig.2 and are lower for glycolitic muscles /LD & SM/. In oxidative-glycolitic muscles /RA/ these vales are higher /over 6,0/ and significant / $p < 0,05$ /. Effect of stress-sensitivity has exerted different influence on that characteristic in single type of muscles. In m.LD and m.SM differences are significant at $p < 0,05$, while in m.RA $p < 0,100$. Results obtained by us for pH_1 in m.LD and m.SM are higher than these reported by Pinkas et al., /1987/ and Yablansky /1988/ for two-line hybrid pigs, but are one-way for lower values of this characteristic in stress-sensitive animals.

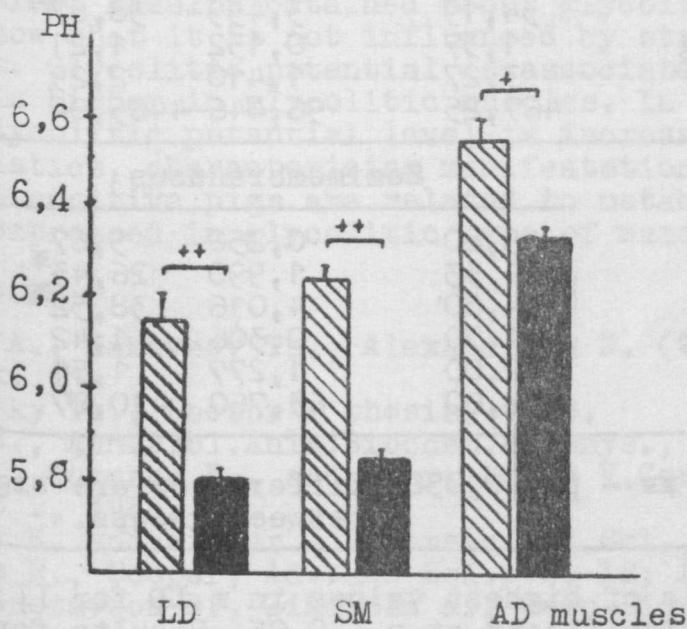


Fig.2. pH_1 - values /45 min. post mortem/

GP established by us shows a trend toward a higher level in animals of pH lower than 6,0, reducing in a course glycolytic toward an oxidative-glycolytic muscle, but relationship between these two characteristics is not yet well elucidated. Data about pH₂ measured 24 h post mortem are lower in m.LD and m.SM compared² to m.RA /table 2/. Values for this characteristic show difference in the velocity of processes in animals of both groups. According to Lister /1971/ glycolysis stopping is associated with pH-value called "boundary". It is nearly 5,2 - 5,2 in glycolytic muscles in pigs.

Table 2. Physico-chemical analysis of LD and SM muscles

Characteristics	Group I		Group II	
	\bar{x}	$\pm \sigma$	\bar{x}	$\pm \sigma$
Longissimus dorsi				
pH 24h	5,75	0,262	5,69	0,205
Colour, 525 nm	25,56	2,346	29,63 ^{***}	1,911
WBC, %	34,19	3,459	38,87 ^{**}	3,508
Myoglobin, mg/g	1,37	0,152	1,21	0,277
Fats, %	3,27	1,410	2,29	0,612
Collagen, mg %	467,25	96,446	4465,04	59,644
Semimembranusus				
pH 24h	5,80	0,250	5,67	0,196
Colour, 525 nm	21,33	1,990	26,43 ^{**}	3,000
WBC, %	34,80	4,016	38,52 ^{**}	2,185
Myoglobin, mg/g	1,70	0,306	1,42	0,296
Fats, %	2,00	1,277	1,51	0,934
Collagen, mg %	452,60	63,760	410,07	67,000

^{*} - $p < 0,100$; ^{**} - $p < 0,050$; differences are significant between groups.

Meat colour is of highest values in m.LD for II group, differences being significant at $p < 0,05$. Results for m.SM are one-way to these for m.LD, difference between groups being significant at a lower significance degree / $p < 0,100$ /. This characteristic depends in a great degree on both myoglobin content and on the proportion of its three forms /9/. Paler colour in II group animals is associated with denaturation of muscle proteins, which changes colour due to myoglobin.

The lack of significant differences in myoglobin content between groups in m.LD and m.SM confirms the results of a series of authors according to which changes in meat colour for stress-sensitive animals are associated to structural changes in muscular proteins /3, 14, 16/.

Fast post mortem glycolysis in II group animals has created conditions for increasing free water in both muscles as well, differences between groups being significative / $p < 0,100/$.

Quantities of all qualitative characteristics of meat in animals of II group characterize it as Pale, Soft, Exudative meat.

Results obtained by us have established no significant differences between groups for both fats and collagen. They are close to those of Ollivier et al., /1978/ and Monin et al., /1981/, which do not find differences in that characteristic in m.LD, between stress-sensitive pigs, being known that the first ones are of a better developed musculature.

CONCLUSION: Results obtained about glycolytic activity of muscles show that it is not influenced by stress-sensitivity of animals. Glycolytic potential is associated with metabolic type and is higher in glycolytic muscles. In stress-sensitive animals glycolytic potential level is increased. Qualitative characteristics, characterizing manifestation of PSE-syndrome in stress-sensitive pigs are related to metabolic type and are best expressed in glycolytic type of muscles.

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