5:32

Effect of a Proteolytic Enzyme Preparation on the Morphological Structure of the Fille Mass of Cooked-Smoked Non-Perishable Sausage

ki

adi A

tr

le:

to

Sto

D

K. VHULKOVA, M. ZHIKOV, P. ALEXIEV and S. DANCHEV

Higher Institute of Food & Flavour Industries, 26 Maritsa Blvd., 4002 Plovdiv, Bulgaria

SUMMARY: This is a study of the microstructural changes in cooked-smoked non-peris^{ball} sausages treated with a protease with the aim to intensify the ageing process involved ¹ the technology of this type meat products. The meat for the test sausages has been treat with 0.01% and 0.02% Mezenterin 11-11 enzyme preparation. The studies have been carried out using a transmission electron microscope. In the non-treated sausages there have been observed optically thick parts in the 2-line zone. In the sausages prepared with 0.01% ²⁰ 0.02% enzyme the protease has hydrolysed the myofibrils and as a result the sarcomeres have lost their individual contours and have formed a net of fine granulated myofibrilly proteins. There are certain changes in the collagen fibers as well, and there are section where the cross furrows have disappeared. The changes established have improved the ²⁰⁰ ge texture in the cases where the meat has been protease-treated.

INTRODUCTION: Texture is an important organoleptic characteristic for all meat product whereby the consumers appreciate their quality. The technological processing of meat for cooked-smoked non-perishable sausages largely depends on the progress of the long autour tic changes during the postmortal ageing period (Bandman et al. 1988). It is well-known that the tenderizing of meat post mortem is assisted by the proteolytic processes which dependent on the degree of activity of the lysosomic enzymes (cathepsins) and the calcium use proper enzyme preparations in order to intensify the proteolytic processes. Many sum point out that the changes in the muscles treated with proteolytic enzymes (tripsin, full papain, etc.) are analogical to the changes occurring during meat ageing (Vassilyev et al. 1985).

The objective of the present work is to identify the microstructural changes in the fill ing mass of cooked-smoked non-perishable sausages treated with a bacterial proteolytic enzyme preparation.

MATERIALS and METHODS: The microbial protease Mezenterin 11-11 was obtained from ^{pacilo} mesentericus, with a proteolytic activity of 210 PU/g enzyme, 55-60°C temperature optime 6.5 pH optimum, and was used in our study. The enzyme prepararion was added in cooked. ^{polo} non-perishable sausages which had the following composition: 30 kg veal, 40 kg semifet ^{of} 30 kg non-fat pork, 2.2 kg salt, 0.007 kg sodium nitrite, 0.1 kg sugar, 0.3 kg pepper, kg garlic. The veal was cooled to 0°-4°C and minced on a 4 mm plate. After adding ^{salt} for another 3-4 min. Then the pork cuts were added and the grinding continued until ^{4.5} The sausages were shaped and then treated according to the technology established for And of meat products. Control samples were prepared in exactly the same way without enzyme addition. Samples of the test and control sausages were sliced on a ultramicrotom LKB 8800. After being contrasted according to Reynolds (1963), the meat sections were studied on a transmission electron microscope Tesla BS-613 at 80 kV. Finally, the sausages were organoleptically evaluated by a 9-member panel using the 9-grade scale of the Higher Institute of Meat Technology Research in Moscow. The results from the evaluation were analysed according to the methods of mathematical statistics (Georgieva et al. 1987).

RESULTS and DISCUSSION: The microstructural changes which have occurred in the cookedstoked non-perishable sausages without enzyme can be seen on Fig.1.



INE

18

hable

a tr

ed been

76 200

5

1181

tion

aust

ducti

FOT

015

M

ch an

t to

122200

Picifi

5 21

a fill

,

cili

imu

SIDO

. Pol

ti

-5 m

ster

this

It is obvious from the picture that the heat treatment has denaturated the myofibrillar proteins, and, as a result, the myofibrils have lost their fibrillar nature (Fig.1). In the Z-line zone there are light bands among which are the optically thick parts of the Zlines and some very fine grains which outline the sarcomeres clearly.

Fig.1. Electron microscopic picture of the longitudinal section of cooked-smoked non-perishable sausage without enzyme. BAR = $0.07 \,\mu$ m.

In the middle of the anisotropic sections there are clear traces of the H-zones and M-lines (MiG.1). The electron microscopic picture of the sausages with added Mezenterin 11-11 enzyme (0.010). The electron microscopic picture of the sausages with added Mezenterin 11-11 enzyme (0.01% or 0.02%) is different from the one described above. It can be seen that the enzyme has hydrolysed the myofibrils to a different extent, and as a result the contours of the ^{sercomeres} have been obliterated, and a net of fine-grain myofibrillar proteins has been tormed (Figures 2-4). At some places there are light spatial bands of hydrolysed and extra-^{cted} ^{Wofibrillar} proteins. We suppose that the proteolytic enzyme preparation has assisted the dethe degradation of the major proteins localized in the Z-lines (desmin and \measuredangle -actinin) as Well p ^{Ne}ll as the gap-filaments that are built up of the proteins titin and nebulin. This is also Probable controllight and nebulin. This is also Probably the reason for the microstructural changes established, namely the degradation of the 21. We accept that these changes the z-lines and the loss of the sarcomere contours (Fig.5). We accept that these changes impart less density to the above-mentioned fine net of myofibrillar proteins. The electron ¹ess density to the above-mentioned fine net of myorrested sausages is a typical one. (Fig. 3) (Pig.3). Along the length of single collagen fibers there are sections of well-preserved Cross for Cross furrows and others where they have disappeared, and the fibers are frayed. At some places there can be seen some destruction and swelling of the collagen fibers. The changes in the in the myofibrills and those in the collagen fibers are probably responsible for the better Seuse texture found at the organoleptic analysis (Table 1).



Fig.2. Electron microscopic picture of cooked-smoked non-perishable sausage, treated with 0.01% enzyme. BAR=0.10 μ m.



Ta

Fig. 3. Electron microscopic picture of cool smoked non-perishable sausage, treated with 0.01% enzyme. BAR=0.05 µm.

The results from the organoleptic evaluation given in Table 1 show that Mezenterin 11-11 has improved the organoleptic properties of the cooked-smoked non-perishable sausages. The highest grade is seen for the sausages with 0.01% enzyme addition (8.82) as compared to the controls (7.20). This is expressed best in their outer appearance, texture and juiciness.

<u>CONCLUSIONS</u>: 1. The microstructural changes observed indicate that the enzyme preparative Mezenterin 11-11 accelerates protein proteolysis in meat. These processes are accompanied by a reduced density of the myofibrillar structures of the muscle fibers and improved ter ture of the finished product. 2. When added in a concentration of 0.01% in relation to the filling mass, the proteolytic enzyme improves the organoleptic properties of the cooked smoked non-perishable sausages.



Fig.4. Electron microscopic picture of cooked-smoked non-perishable sausage, treated with 0.02% enzyme. BAR=0.10 Jum.



Fig.5. Electron microscopic picture of cook smoked non-perishable sausage, treated with 0.02% enzyme. BAR=0.07 /m.

802

lable 1. Organoleptic evaluation of cooked-smoked non-perishable sausages prepared with di-

fferent additions of Mezenterin 11-11 enzyme preparation.

| Characteristic | Control | Enzyme concentration, % | |
|------------------|-----------|-------------------------|-------------|
| | | 0.01 | 0.02 |
| Texture | 6.43±0.27 | 8.86 ± 0.10 | 8.25 ± 0.28 |
| Juiciness | 6.00±0.20 | 8.91 ± 0.09 | 8.17 ± 0.27 |
| Flavour | 6.72±0.28 | 8.10 ± 0.21 | 7.98 ± 0.30 |
| Taste | 7.34+0.29 | 8.57 ± 0.31 | 8.00 ± 0.31 |
| Outer appearance | 8.80±0.20 | 8.97 ± 0.30 | 8.41 ± 0.28 |
| Colour | 8.13=0.31 | 8.79 ± 0.17 | 8.60 ± 0.23 |
| Overall grade | 7.20±0.19 | 8.82 ± 0.17 | 8.41 ± 0.26 |

:00% vith

.11 The

o the

551

ratil ied

tex the

d-

vith

REFERENCES: BANDMAN, E., ZDANIS, D. (1988): An immunological method to assess protein de-Bradation in post-mortem muscle. Meet Sci. 22:1.

 W_{AN} , S.F., BANDMAN, E. (1989): Studies of Desmin and \angle -Actinin Degradation in Bovine Senitendinosus Muscle. J.Food.Sci. 54:1426-1430.

KOOHMARAIE, M., BABIKER, A.S., SCHROEDER, A.L., MERKEL, R.A., DUTSON, T.R. (1988): Accele-Pation of Postmortem Tenderization in Ovine Carcasses Trough Activation of Ca²⁺ -Dependent Proteases. J. Food Sci. 53:1638-1641.

VASSILIEV, A.A., LUZAN, V.H. (1985): Obzornaya informatsiya myasnaya promishlenost. 6:22-27.

YOUNG, O.A., GRAAFHUIS, A.E., DAVEY, C.L. (1980): Postmortem changes in cytoskeletal proteins of muscle. Meat Sci. 5:41.

LAZARIDES, E. (1980): Intermediate filaments as mechanical integrators of cellular space. Nature. 283:249.

ELGASIM, E.A., KOOHMARATE, M., ANGLEMIER, A.F. KENNICK, W.H., ELKHALIFA, E.A. (1985): The Combined effect of the calcium activated factor and cathepsin D on skeletal muscle. Food Microstruc. 4:55.

^{10DOR}INOV, S., TANCHEV, S. (1987): Matematiko-statisticheski metodi v tehnologichnite ^{1281edvaniya}. Selskostopanska nauka. 25: 100-109. REINOLDS, E. (1963): Cell. Biol. 17:208.