Effect of Dietary Fat Quality and Alpha-Tocopheryl Acetate Supplementation on the Susceptibility of Porcine Tissue to Lipid Peroxidation

T.K. MURPHY<sup>a</sup>, P.B. LYNCH<sup>c</sup>, D.J. BUCKLEY<sup>b</sup>, F.J. MONAHAN<sup>b</sup> and P.A. MORRISSEY<sup>a</sup>

<sup>a</sup>Department of Nutrition, University College Cork, Republic of Ireland; <sup>b</sup>National Food Biotechnology Centre, University College Cork, Republic of Ireland; <sup>c</sup>Teagasc, Pig Husbandry Unit, Moorepark, Fermoy, Co. Cork, Republic of Ireland

#### SUMMARY

Dietary fat quality had a marked influence on the fatty acid composition of porcine muscle and liver. The ratio of C18:2/C18:1 was significantly higher in the muscle and liver from pigs fed the fresh soya oil diet compared to pigs fed the oxidized soya oil diet. Tissues from pigs fed the oxidized soya oil diet were also found to be more susceptable to lipid peroxidation than tissues from pigs fed fresh soya oil. Alpha-Tocopheryl acetate supplementation significantly reduced the susceptability to lipid peroxidation of liver and muscle from pigs fed either fresh or oxidized soya oil diets. It can be concluded that dietary fat quality may cause changes in the fatty acid composition leading to changes in the susceptability of tissue lipids to oxidative attack. However, dietary alpha-tocopherol acetate may offer an effective means of minimizing the adverse effects of dietary fat quality.

#### INTRODUCTION

Dietary fat quality influences the fatty acid composition of porcine muscle and adipose tissue (Marchello <u>et al.</u>, 1983; Rhee <u>et al.</u> 1988). Changes in fatty acid composition may in turn be accompanied by changes in the susceptability of tissue lipids to Oxidative attack (Rhee <u>et al.</u>, 1988). Alpha-tocopheryl acetate supplementation of pig diets has been shown to result in increased alpha-tocopherol concentration in porcine tissues with a resultant increase in the stability of muscle (Monahan <u>et al.</u>, 1989).

The objectives of this study were to investigate the effects of dietary fat quality and alpha-tocopheryl acetate supplementation on: (a) the fatty acid profiles of porcine muscle and liver (b) the susceptability of porcine muscle and liver to lipid oxidation and (c) the alpha-tocopherol levels in the various tissues.

# MATERIALS and METHODS

Thirty-two Landrace X Large White pigs, weighing approximately 45 Kg liveweight were randomly divided into 4 groups of eight. Each group was fed a diet containing 3% fresh soya oil or 3% oxidized soya oil with either a basal level (10mg/Kg diet) or a supplemented level of alpha-tocopheryl acetate (200mg/Kg diet). The oxidized soya oil had a peroxide value of approximately 100meq O<sub>2</sub> /Kg oil. The pigs were given water and feed ad libitum and were slaughtered when they weighted in excess of 85Kg. At evisceration, liver samples were taken. The carcasses were then chilled overnight to 5°C and a sample of the Longissimus dorsi muscle was taken.

All samples were vacuum packed and stored at -20°C until required. Fatty acid methyl esters of the neutral and polar lipid fractions isolated from muscle and liver were prepared by the method of Slover and Lanza (1979) and Maxwell and Marmer (1983), respectively. The methyl esters of the fatty acids were analysed on a glass column, 2m X 3mm i.d. containing 10% Silar-10C on 100/120 mesh Gas-Chrom Q. The analysis was performed on a Shimadzu GC-14A gas chromatograph equipped with a flame ionization detector. A Shimadzu C-R6A Chromatopac intergrator was used for the calculation of peak areas. Alpha-tocopherol levels in liver and muscle were determined using the HPLC method of Buttriss and Diplock (1984). The stability of muscle tissue homogenates to iron-induced lipid peroxidation was determined by a modification of the method of Kornbrust and Mavis (1980). 2-Thiobarbituric acid reactive substances (TBARS) was determined by the method of Buege and Aust (1978) and reported as nmoles malonaldehyde per mg of protein as outlined by Monahan et al. (1989).

#### **RESULTS and DISCUSSION**

Muscle tissue fatty acids: The neutral and polar lipid fractions isolated from the L. dorsi muscle of pigs receiving the 3% oxidized soya oil diet had a higher proportion of C18:1 and a lower proportion of C18:2 than the lipid fractions from pigs fed the 3% fresh oil diet. The differences were significant in all cases (p<0.05) except for levels of C18:1 in muscle samples from pigs fed the oxidized soya oil diet with basal levels of alpha-tocopheryl acetate.

Both lipid fractions in liver from pigs fed the 3% oxidized soya oil diet had significantly higher levels of C18:1 (p<0.05) and significantly lower levels of C18:2 (p<0.05) than pigs fed the 3% fresh oil diet. In liver and muscle the ratio of C18:2/C18:1 was significantly lower (p<0.05) in both the neutral and polar lipid fractions. Alpha-tocopheryl acetate supplementation reduced the effect of oxidized soya oil on fatty acid profiles. However, the effect was not significant. In the case of the neutral and polar fractions of liver and the polar fraction of muscle, the ration of unsaturated/saturated fatty acids was significantly higher (p<0.05) in pigs fed the fresh oil diet.

<u>Tissue alpha-tocopherol levels:</u> The alpha-tocopherol levels were significantly lower (p < 0.05) in liver and muscle from pigs fed the oxidized soya oil diet compared to the fresh soya oil diet except for muscle from pigs fed the basal alpha-tocopherol diets (Table 1). These results are in agreement with those reported from experiments with chicks by Lin <u>et al.</u> (1989) and Sheehy <u>et al.</u> (1990). It is likely that hydroperoxide degradation products from the oxidized soya oil in the diet induced rapid oxidation of the membrane-bound lipids in the tissues. Thus, the lower alpha-tocopherol levels in tissues from pigs fed the the oxidized soya oil is due to its action as a free radical scavenger. Century and Horwitt (1959) observed a vitamin E deficiency disease, encephalomalacia, in broilers fed high levels of an oxidized diet.

<u>Lipid oxidation in muscle and liver</u>: Dietary alpha-tocopheryl acetate supplementation significantly increased (p < 0.05) the stability of liver and muscle tissues to lipid peroxidation after 160 min incubation, from pigs fed both the fresh and oxidized soya oil diets (Fig. 1). These results are in agreement with findings by Monahan <u>et al.</u> (1990).

Muscle and liver tissues from pigs fed the oxidized soya oil diets were more susceptable to lipid oxidation than tissues from pigs fed the fresh oil diets of comparable alpha-tocopheryl acetate supplementation except for muscle samples in pigs fed the high supplementation diet. The increased susceptability to lipid peroxidation in the oxidized soya oil groups may be due to the increased number of allyl groups present (Arkawa and Sagai, 1986). In addition the residual levels of alpha-tocopherol in liver and muscle tissues from pigs fed the oxidized soya oil diet may contribute to the increased susceptability to lipid peroxidation.

### CONCLUSIONS

Feeding an oxidized soya oil diet to pigs had the effect of lowering the residual alpha-tocopherol levels in muscle and liver for both basal and supplemented diets. An increased susceptability to lipid oxidation was observed. Oxidized fat had a significant effect on the fatty acid profile resulting in decreased unsaturated to saturated and C18:2 to C18:1 ratios.

Table 1. Mean  $\alpha$ -tocopherol content and standard error of the mean of liver and muscle of pigs fed fresh and oxidized soya oil diets with basal and supplemented levels of  $\alpha$ -tocopheryl acetate.

Dietary treatment	α-tocopherol <sup>1</sup>			
	Liver (mg/g)	Muscle (mg/g)		
Fresh soya oil:				
basal	11.52±0.76a	3.95±0.28a		
supplement	98.57±6.15b	13.57±0.71b		
Oxidized soya oil:				
basal	9.30±0.65°	3.33±0.41a		
supplemented	82.03±3.19d	9.67±0.49d		

<sup>1</sup> Mean values and standard error of the mean of six analyses a b c d Means in the same column bearing different superscripts differ significantly (p<0.05)

Fig. 1: Effect of dietary fat quality and alpha-tocopheryl acetate supplementation on iron iduced lipid peroxidation in pig muscle and tissue

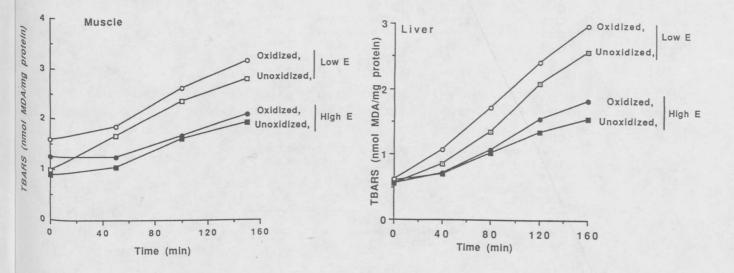


Table 2. Fatty acid profiles of the neutral lipid fraction of liver from pigs fed fresh soya oil and oxidized soya oil diets with control and supplemented levels of  $\alpha$ -tocopheryl acetate.

Fatty acid	Fresh Sova oil 1.2		Oxidized Sova oil	
	Control	Supplemented		
	%	%	%	%
214:0	0.526	0.576	1.532	1.392
216:0	13.352	14.012	15.342	
216:1	1.832	1.742	0.921	1.142
218:0	23.932	23.35ª	25.632	24.712
218:1	12.99b	13.53b	15.792	
C18:2	21.042	20.852	18.23b	18.95b
C18:3	1.272	1.073	0.732	1.012
220:4	13.912	14.362	12.962	13.572
C20:5	1.792	1.628	0.946	1.096
C22:0	2.132	2.072	3.112	2.932
C22:5	3.822	3.912	2.526	3.11ab
222:6	2.942	2.882	1.63b	2.23ab
Total saturates	39.93b	40.00b	45.61a	43.41a
Total unsaturates Ratio:	59.59a	59.962	53.726	55.81b
Unsaturate/Saturate	1.492	1.502	1.13b	1.29b
C18:2/C18:1	1.622	1.542	1.15b	1.29b

<sup>1</sup> Percent of total peak area of fatty acids listed

Table 3. Fatty acid profiles of the polar lipid fraction of liver from pigs fed fresh soya oil and oxidized soya oil diets with control and supplemented levels of  $\alpha$ -tocopheryl acetate

	Fresh Sova oil 1.2		Oxidized Sova oil	
Fatty acid	Control %	Supplemented %	Control %	Supplemented %
C14:0	0.716	1.04b	1.51a	1.412
C16:0	14.532	15.062	17.692	16.932
C16:1	0.93b	1.572	0.53b	0.926
C18:0	29.431	27.932	30.632	29.382
C18:1	12.23b	11.68b	13.992	14.062
C18:2	22.932	22.312	19.43b	20.47b
C18:3	1.322	0.932	0.622	0.892
C20:4	13.322	14.522	11.47b	11.52b
C20:5	0.932	0.632	0.432	0.562
C22:0	1.18c	2.12bc	2.5921	3.132
C22:6	2.102	1.3426	0.716	0.73b
Total saturates	45.85b	46.09b	52.422	50.852
Total unsaturates Ratio:	53.762	52.99a	47.18b	48.59b
Unsaturate/Saturate	1.172	1.152	0.90b	0.96b
C18:2/C18:1	1.872	1.912	1.39b	1.46b

<sup>1</sup> Percent of total peak area of fatty acids listed

Table 4. Fatty acid profiles of the neutral lipid fraction of muscle from pigs fed fresh soya oil and oxidized soya oil diets with control and supplemented levels of  $\alpha$ -tocopheryl acetate.

Fatty acid	Fresh Sova oil <sup>1,2</sup>		Oxidized Sova oil	
	Control %	Supplemented %	Control %	Supplemented %
C14:0	0.931	1.032	0.712	0.802
C16:0	23.742	24.322	26.412	26.012
C16:1	3.022	3.62ª	3.932	3.872
C18:0	13.932	13.69ª	13.922	13.932
C18:1	38.58b	40.32b	44.612	44.473
C18:2	13.572	13.47a	8.02b	8.43b
C18:3	2.712	2.86ª	2.12a	2.262
C20:0	0.412	0.35ª	0.392	0.282
C20:4	0.042	0.02a	0.092	0.062
C20:5	0.032	0.042	0.082	0.052
C22:6	0.382	0.412	0.162	0.232
Total saturates	39.012	39.392	41.432	41.022
Total unsaturates Ratio:	58.33ª	60.742	58.562	59.372
Unsaturate/Saturate	1.492	1.542	1.412	1.452
C18:2/C18:1	0.352	0.332	0.18b	0.196

<sup>1</sup> Percent of total peak area of fatty acids listed

<sup>&</sup>lt;sup>2</sup> Means in the same row followed by different superscripts are significantly different (p < 0.05).

<sup>&</sup>lt;sup>2</sup> Means in the same row followed by different superscripts are significantly different (p < 0.05).

 $<sup>^2\,\</sup>text{Means}$  in the same row followed by different superscripts are significantly different (p < 0.05).

Table 5. Fatty acid profiles of the polar lipid fraction of muscle from pigs fed fresh soya oil and oxidized soya oil diets with control and supplemented levels of  $\alpha$ -tocopheryl acetate

	Fresh Soya oil1.2		Oxidized Sova oil	
Fatty acid	Control	Supplemented		Supplemented
	%	%	%	%
C14:0	1.15b	0.916	1.94a	1.3126
C16:0	17.51b	18.32ab	20.482	19.922
C16:1	1.462	1.05ª	1.532	1.782
C18:0	14.13b	15.59ab	17.432	18.10a
C18:1	15.68b	14.38b	18.62ª	19.32a
C18:2	36.232	35.472	30.486	29.48b
C18:3	3.232	2.97ab	2.37ab	2.026
C20:4	7.582	7.99a	5.26b	6.47ab
C20:5	1.23a	1.43a	0.536	0.62b
C22:0	0.01a	Пa	0.03a	0.042
C22:6	1.082	1.222	0.59a	0.442
Total saturates	32.80b	34.82b	39.882	39.372
Total unsaturates	66.49a	64.512	59.35b	60.13b
Ratio:				
Insaturate/Saturate	2.034	1.85ª	1.496	1.53b
C18:2/C18:1	2.31a	2.47a	1.64b	1.53b

1 Percent of total peak area of fatty acids listed

 $^2$  Means in the same row followed by different superscripts are significantly different (p < 0.05).

## REFERENCES

Arkawa, K. and Sagai, M. (1976). Species differences in lipid peroxide levels in lung tissue and investigation of their determining factors. Lipids <u>21</u>: 769

Buege, J.A. and Aust, S.D. (1978). Microsomal lipid peroxidation. Methods in Enzymology. 52: 302

Buttriss, J.L. and Diplock, A.T. (1984). High performance liquid chromatography methods for vitamin E in tissues. Methods in Enzymology. 105: 131

Century, B. and Horwitt, M.K. (1959). Effect of fatty acids on chick encephalomalacia. Proceedings of the Society of Experimental and Biologicla Medicine. 102: 375

Kornbrust, D.J. and Mavis, R.D. (1980). Relative susceptibility of microsomes from lung, heart, liver, kidney, brain and testes to lipid peroxidation: correlation with vitamin E content. Lipids <u>15</u>: 315

Lin, C.F., Asgher, A, Gary, J.I., Buckley, D.J., Booren, A.N., Crackel, R.L. and Flegal, C.J. (1989). Effects of dietary oils and alpha-tocopherol supplementation on lipid peroxidation and stability of broiler meat. Journal of Food science 54: 1457

Marchello, M.J., Cook, N.K., Slanger, W.D., Johnson, V.K., Fischer, A.G. and Dinusson, W.E. (1983). Fatty acid composition of lean and fat tissue of swine fed dietary levels of sunflower seed. J. Food Sci. 48: 1331

Maxwell, R.J. and Marmer, W.N. (1983). Systematic protocol for the accumulation of fatty acid data from multiple tissue samples. Lipids 18: 453

Monahan, F.J., Hanrahan, T.J. and Lynch, P.B. (1989). The effect of dietary Vitamin E on the stability of raw and cooked pork. Meat Sci. 27: 99

Rhee, K.S., Ziprin, Y.A., Ordonez, G. and Bohac, C.E. (1988). Fatty acid profiles of the total lipids and lipid oxidation in pork muscle as effected by Canola oil in the animal diet and muscle location. Meat Sci. 23: 210

Sheehy, P.J.A., Morrissey, P.A. and Flynn, A. (1990). The effects of heated sunflower oil on concentrations of alpha tocopherol and thiobarbituric acid-reacting substances in chick tissues. Proceedings of the Nutrition Society 49: 112

Slover, H.T. and Lanza, E. (1979). Quantitative analysis of food fatty acids by capillary gas chromatography. JAOCS <u>56</u>: 933