

HISTOLOGIC STUDY OF SOME ORGANS IN ORGANISM OF LAMBS FED CLENBUTEROL

LILIA DITCHEVA

Institute of Animal Science, 2232 Kostinbrod, Bulgaria

SUMMARY

In three groups of lambs fed both low-, moderate- and highconcentrated, the effect of clenbuterol on the histostructures of some organs has been studied. Lambs received at 10 g/kg feed clenbuterol daily for 42 days. In each group, of four animals samples for histologic analyses of duodenum, jejunum, abomasum, liver, lung and kidneys have been taken. Results of the study show a certain effect of feeding type on the histostructures of duodenum, jejunum and abomasum. Supplementation of 10 mg clenbuterol/kg of feed daily induces no essential changes. In all three experimental groups stagnation and dystrophic effects were established in the liver, as well as a series of changes in their kidneys, concerning mainly Malpighic corpuscles. In lungs, inflammatory processes observed in experimental parenchim with a predominant hemorrhagic character occur mainly in animals of groups does not give sufficient grounds for categorical inferences.

INTRODUCTION

Studies of the effect of B-adrenergic agonist drugs are mainly related to their role changing both protein and fat contents in the organism of animals (BAKER et al, 1984; COLEMAN et al, 1988; HU et al, 1988; CLAEYS et al, 1989). An increasing of total weight and feed efficiency was also established in sheep receiving cholesterol (KIM et al, 1987). A certain interest also represent the date about the effect of these substances on both the growth and carcass characteristics of animals (MERKLEY, BISHWOOD, 1989). On the other hand side effects have been observed induced by clenbuterol using Beta agonists, also influence some organs of both digestive and secretory systems participating most actively in metabolism (WARBURTON et al, 1988). In the present study we ourselves the task of investigating the state of the instostructures in both small intestines, abomasum, liver, kidneys and lungs of lambs reseiving clenbuterol.

MATERIAL AND METHODS

The trial was conducted on 60 male lambs, three-breed cross, weaned at 45 days of age, average live weight of 15,4 kg. For 90 days, animals divided into 6 groups of 10 each received respectively: 1 and 4 groups- low concentrated feeding- 4,0 MJ energy/kg and 200 g protein/kg mixture; 2 and 5 groups- moderate- concentrated feeding- 5,1 MJ energy/kg, 200 g protein/kg of mixture; 3 and 6 groups- high- concentrated feeding- 6,0 MJ energy/kg, 200 g protein/kg mixture.

When reaching 26,5 kg of live weight, 10 mg of clenbuterol per kg of feed has been conditioned, mixed with starch at a ratio of 1:1 mg/g for 4,5 and 6 experimental groups. Control 1,2 and 3 groups receive above-mentioned energy and protein without clenbuterol until accomplishing the experiment. After 42 days, of 4 animals of experimental and control groups samples were taken for studying the histosyrtuctures in both small intestines, abomasum, liver, kidneys and lungs. Samples of these organs have also been taken a week after stopping clenbuterol in experimental groups. Materials for histologic study were fixed in formalin and coloured with heamatoxilin-eosin.

RESULTS AND DISCUSSION

Data obtained in the study for developing the histosyrtuctures in both duodenum, jejunum and abomasum may be examined in two aspects: on one hand, determining the effect of different feeding type and on the other hand studying the clenbuterol effect under low, moderate and high-concentrated feeding on these structures. In table 1, development of histostructures is given for duodenum in control 1,2 and 3 groups, and experimental 4,5 and 6 groups. From the measurements of histosyrtuctures in duodenum, the significance arises about feeding types used in the experiment under relevant combinations of energy for developing this part of intestine. Clenbuterol supplementation does not change that between control and experimental groups does not give grounds for considering Clenbuterol supplementation has influenced significantly this intestinal area.

In the jejunum of the small intestines total thickness is practically equal under different feeding types for control groups (table 2). For separate histosyrtuctures higher differences should be pointed out under high concentrated feeding compared to low one. Much more marked are the differences in the histosyrtuctures between experimental groups. Under highconcentrated feeding, Clenbuterol supplementation induced probably additionally a poorer development of most structured in that intestinal area.

In the abomasum of control groups a greater height of the glands has been established in animals under high-concentrated feeding. In the rest of structures differences have been established between the groups (table 3). In experimental groups, a trend toward poorer development of muscular layers and total depth for control animals under high-concentrated feeding, is here more clearly expressed and statistically significant. Studying the results of measurements under different feeding types, clenbuterol supplementation was established to exert no significant effect on development of histostructures in the abomasum.

The liver of animals examined in control groups was without visual changes. Only a lamb fed high concentrate a pelleting of cytoplasm of the hepatocytes and hyperemia was observed. Clenbuterol supplementation in animals under high-concentrated feeding leads to strengthening these processes, hepatocytes in liver of animals are of unclear cellular borders, pelleted cytoplasm, lysis or rexis of nuclei. Arranging of beams is very changed under the pressure of enlarged sinuses. In lambs of experimental groups under low-and moderate concentrated feeding, the condition of liver is similar to those under concentrated feeding. Stagnation phenomena are slightly poorly expressed but here often pelleting of cytoplasm and degradation of hepatocytes are observed. A week after stopping the clenbuterol, in some of these animals a trend arises toward normalization but in most cases the changes described above remain.

In the kidneys of animals of control groups, Malpighic corpuscles and functional canals of the nephrons are both form and structure normal for them. For experimental groups receiving clenbuterol, occurred changes are concentrated mainly in Malpighic corpuscles. Most of them are of irregular form, there is a narrowing of the area Baulman-Shumlyanski capsule, reaching to sticking of visceral leaf to parietal one. Other Malpighic corpuscles an exudat was observed where often exist degenerated elements of the blood. In single cases, glomerules are observed where a degeneration of capillaries occurs and in Baulman-Shumlyanski space many forms of elements, mainly leucocytes are observed. Variety in changing degree of extracapillary and intracapillary level of glomerules on the one hand as well as non-equal affecting of Malpighic corpuscles on the other hand, determine clenbuterol effect during developing. A week after stopping, these changes are being observed to some extent.

In lungs, inflammatory processes observed of pulmonary parenchyma with a predominant haemorrhage character occurred mainly in animals of experimental groups. It should be noticed, however, that a part of control animals had similar state of lungs.

CONCLUSION

Investigation of both duodenum, jejunum and abomasum shows a definite effect of feeding type on histostructures studied. Supplement of 10 mg clenbuterol per kg feed does not induce significant changes. Condition of liver, however, gives a reason to consider that as a result of clenbuterol intake for experimental animals, both stagnation and distrophic phenomena are developed. In kidneys of these animals a series of changes are observed affecting mainly Malpighic corpuscles. That determines our standpoint that these organs clenbuterol influences negatively their normal function. Observations that lungs of experimental and control animals do not give sufficient reason for inferences regarding the significance of clenbuterol concerning the changes established in these organs.

REFERENCES

- BAKER P. K., DALRYMPLE R. H., INGLE D. L., RICKER C. A., 1984. Use of a Beta adrenergic agonist to alter muscle and fat deposition in lambs. *J. Animal Sci.*, 52, 1256-1261.
- BROCKER J. A., BLUM J. K., 1990. Effects of a Beta adrenergic agonist on performance, body composition and nutrient retention in finishing pigs fed normal or amounts of protein. *Animal Production*, 51, 601-612.
- BROCKWAY J. M., MACRAE J. C., WILLIAMS P. E., 1987. Side effects of clenbuterol as a repartitioning agent. *Veterinary Record*, 120, 381-383.
- CLAEYS M. C., MULVANEY D. R., McCARTHY F. D., GORE M. T., MARPLE D. N., 1989. Skeletal muscle protein synthesis and growth hormone secretion in lambs treated with clenbuterol. *J. Animal Sci.*, 67, 2245-2254.
- COLEMAN M. E., EKEREN P. A., SMITH S. B., 1988. Lipid synthesis and adipose tissue growth in adipose tissue from sheep chronically fed a Beta adrenergic agent. *J. Animal Sci.*, 66, 372-379.
- HU C. Y., SURYAWAN A., FORSBERG N. F., DALRIMPLE R. H., 1988. Effect of cimaterol on sheep adipose tissue lipid metabolism. *J. Animal Sci.*, 66, 1393-1400.

KIM Y. S., LEE Y. B., DALRYMPLE R. H., 1987. Effect of the Repartitioning Agent, Testosterone on Growth, Carcass and Skeletal Muscle Characteristics in lambs. J. Animal Sci., 65, 1392-1399.

MERKLEY J. W., GARWOOD V. A., 1989. Growth Response to chronic Beta Agonist Treatment. Nature carcass characteristics of Quaie Selected for High and Low Body Weight. Pediatr. Crit., 62, 1500-1503.

RICKS C. A., DALRIMPLE R. H., BAKER P. R., 1984. Use of a Beta agonist to alter fat and muscle deposition in steers. J. Animal Sci., 59, 1247-1255.

WARBURTON D., PARTON L., BUCKBY S., 1988. Effects of b-2 agonist on hepatic glycogen metabolism in the fetal lamb, Pediatr res., 24, 330-332.

DEVELOPMENT OF HISTOSTRUCTURE IN DUODENUM

	Control			Experimental		
	1	2	3	4	5	6
length of intestinal villae	577 ± 60 ^a	465 ± 16 ^a	408 ± 8 ^b	526 ± 43 ^a	512 ± 13 ^a	414 ± 8 ^b
Leberkuhn's/glands	481 ± 29 ^a	385 ± 8 ^b	437 ± 29 ^{ac}	461 ± 23 ^a	418 ± 15 ^{ab}	467 ± 4 ^{ac}
Brunner's glands	455 ± 22 ^a	484 ± 56 ^a	509 ± 20 ^a	456 ± 7 ^a	496 ± 35 ^a	422 ± 23 ^a
long-shaped muscular layer	180 ± 19 ^a	193 ± 22 ^a	131 ± 9 ^a	171 ± 13 ^a	189 ± 20 ^{ab}	136 ± 25 ^{ac}
longitudinal muscular layer	70 ± 2 ^a	81 ± 7 ^a	73 ± 9 ^a	65 ± 3 ^a	88 ± 11 ^b	62 ± 5 ^{ac}
Total thickness	1763 ± 70 ^a	1612 ± 56 ^b	1558 ± 53 ^b	1680 ± 44 ^a	1665 ± 63 ^a	1500 ± 21 ^b

Means with different letters are significant,
a, b, c < 0.1; P < 0.05; P < 0.025

Table 2

DEVELOPMENT OF HISTOSTRUCTURE IN JEJUNUM

Variable	Control				Experimental	
	Groups	1	2	3	4	5
Height of intestinal papillae		412 ± 40 ^a	485 ± 64 ^{ab}	497 ± 19 ^b	448 ± 33 ^a	498 ± 28 ^{ab}
Lieberkühn's glands		439 ± 31	444 ± 10	473 ± 13	442 ± 4	459 ± 8
Ring-shaped muscular layer		146 ± 16 ^a	170 ± 26 ^{ab}	118 ± 4 ^b	177 ± 6 ^a	148 ± 17 ^{ab}
Longitudinal muscular layer		78 ± 4 ^a	83 ± 13 ^{ab}	44 ± 2 ^c	82 ± 12 ^a	82 ± 8 ^{ab}
Total thickness		1075 ± 48	1182 ± 73	1152 ± 26	1189 ± 35	1185 ± 25

Means with different leffers are significant,
 $P < 0.1$; $P < 0.05$; $P < 0.025$

Table 3

DEVELOPMENT OF HISTOSTRUCTURE IN AOMASUM

Variable	Control				Experimental	
	Groups	1	2	3	4	5
Height of intestinal papillae		429 ± 8 ^a	413 ± 30 ^{ab}	478 ± 13 ^c	451 ± 20	451 ± 9 ^{bc}
Ring-shaped muscular layer		786 ± 165	593 ± 48	523 ± 60	780 ± 48 ^a	632 ± 53 ^{bc}
Longitudinal muscular layer		209 ± 50	164 ± 18	151 ± 8	232 ± 13 ^a	211 ± 39 ^{as}
Total thickness		1424 ± 199	1171 ± 66	1151 ± 42	1462 ± 55 ^a	1293 ± 68 ^{as}

Means with different leffers are significant,
 $P < 0.1$; $P < 0.05$; $P < 0.025$