AND AND INTRAMUSCULAR FAT DISTRIBUTION IN PIGS I.GARCIA AND CASAL, J.J.

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The quality and quantity of intramuscular fat (IMF) were determined in 31 muscles from by Dura The quality and quantity of intramuscular fat (IMF) were determined in the similar live-weight. We guality and quantity of intramuscular fat (IMF) were determined in the similar live-weight. We guality and quantity of intramuscular fat (IMF) were determined in the similar live-weight. ^{suroc} Jersey barrows of each of two lines, lean (L) and rat(r) to be a sub-titlerences in IMF distribution were detected between lean and fat pigs. The total amount the was We was almost identical (0.43 and 0.41 kg, respectively) in spite of big differences is total dissected fat (32.6 and ^{was} almost identical (0.43 and 0.41 kg, respectively) in spice of the fat (32.6 and ¹/₉₅, ^{dissected} fat (26.6 and 42.9 kg, respectively) or % total dissected fat (32.6 and ¹/₉₅, ^{resp} terr for Mm Vastus lateralis, Semimembranosus, Adductor, Sartorius, Obliguus internus The fatty acid composition of IMF shows a strong Tor Mm Vastus lateralis, Semimembranosus, Adductor, Sartorius, Lessons a strong and Transversus abdominis. The fatty acid composition of IMF shows a strong for the L pigs. and <u>Transversus abdominis</u>. The fatty acid composition the L pigs. Brate Atroduction

Intramuscular fat makes a positive contribution to the palatability characteristics of Select ^{1ntramuscular} fat makes a positive contribution to the palatability characterion of the ^{Selection} towards a maximum of muscularity is related to a strong reduction of the ^{of formation} of formation towards a maximum fat and has led to general decline in meat ^{Selection} towards a maximum of muscularity is related to a strong term in meat ^{Ants of fatty} tissues and intramuscular fat and has led to general decline in meat (Decline) (Decline) and the mount of intramuscular and the ⁽¹⁾ (Desmoulin, 1983). It might be possible that the amount of intrame Neity of the fatty tissues become the limiting factors in breeding extremely lean pigs. In severe the severe the severe the severe the concentration of limiting factors is between the concentration of limiting factors have been found between the concentration of limiting factors the severe the s

In Several studies low correlations have been found between the concentrations that and carcass fatness (Duniec et al., 1961; Hiner et al., 1965). This suggests that deposition and carcass fatness (Duniec et al., 1961; Hiner et al., 1965). In several studies low correlations have been found between the concentration of lipid scle and deposition in muscle is not part of the fattening process which occurs in the other fat epots of the body. FERIN

This research was undertaken to determine the differences in the quality and quantity intramuse. ^{Ins res}earch was undertaken to determine the differences in the quarter intramuscular fat from Duroc Jersey pigs with different fat carcass levels but similar weight Hatcass weights. Aterials and Methods

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^{Eight} Methods ^{Fergamine} Duroc Jersey of each of the two lines, lean (L) and fat (F), reared at Pergamino Experimental Station of INTA were used as experimental animals. They were Pergamino Experimental Station of INTA were used as experimental animats. t the to individual pens with water and a standard diet supplied ad libitum. Both lines end a standard diet supplied ad libitum. Both lines the end a standard diet supplied ad libitum. Both lines (32,9+-2.95 and the endividual pens with water and end of the trial were similar in carcass weight (80.2+-3.18 and 01.2.) and by a the trial were similar in carcass fat % (32.9+-2.95 and by a the similar the similar in carcass fat % (32.9+-2.95 and similar the similar the similar in carcass fat % (32.9+-2.95 and similar the sintersimal the sinteresime similar the similar the simi

 T_{he} right side of the carcasses were kept at -15 C and 31 muscles dissected as soon as F_{he} . Each Weight of the carcasses were here. The back muscle was weighted, minced and aliquot samples extracted with the carcasses were here. Weight of the carcasses were here. The back muscle was weighted, minced and aliquot samples extracted with the carcasses were here. The back muscle was weighted, minced and aliquot samples extracted with the carcasses were here. The back muscle was weighted, minced and aliquot samples extracted with the carcasses were here. The back muscle was weighted, minced and aliquot samples extracted with the carcasses were here. The back muscle was weighted, minced and aliquot samples extracted with the carcasses were here. The back muscle was weighted, minced and aliquot samples extracted with the carcasses were here. The back muscle was weighted, minced and aliquot samples extracted with the carcasses were here. The back muscle was weighted, minced and aliquot samples extracted were used to determine the carcasses were the carcasses were here. The back muscle was weighted, minced and aliquot samples extracted were used to determine the carcasses were used for the carca Each Muscle was weighted, minced and aliquot samples extracted with the method of Weight of chemical fat (IMF) and for fatty acid analysis. TLC and GLC were used for the lipid Separate the separate the separate terms of te lipid of chemical fat (IMF) and for fatty acid analysis. TLC and GLC more reported ^{Cia} et al ^{via} et ^{al} al., 1986). The muscles studied were: Proximal pelvic limb (GI): <u>tensor</u> (TIF), <u>Biceps femoris(BFE)</u>, <u>Glutaeus medius</u>, <u>accessorius et profundus</u> (GLU),<u>Vastus</u> (VIA) <u>Vastus intermedius</u> (VIN), <u>Rectus femoris</u> (RFE), (VIA) <u>Vastus intermedius</u> (VIN), <u>Pectineus</u> (PEC) (TLF), 1986). The muscles studied were. (TLF), <u>Biceps femoris(BFE)</u>, <u>Glutaeus medius</u>, accessorius et profundus (GLU, (RFE), (VLA), <u>Vastus medialis</u>(VME), <u>Vastus intermedius</u>(VIN), <u>Rectus femoris</u>(RFE), <u>Sources</u>(GAS), <u>Fibularis</u> Addie (VLA), <u>Biceps femoris(BFE)</u>, <u>Glutaeus median</u> (VIN), <u>Rectus lemora</u> <u>Sartorius (STE)</u>, <u>Vastus medialis</u> (VME), <u>Vastus intermedius</u> (VIN), <u>Rectus lemora</u> (PEC) <u>Sartorius (STE)</u>, <u>Gracilis</u> (GRA), <u>Semimembranosus</u> (SME), <u>Adductor</u> (ADD), <u>Pectineus</u> (PEC) <u>Sartorius</u> (STE), <u>Gracilis</u> (GRA), <u>Semimembranosus</u> (SME), <u>Adductor</u> (ADD), <u>Pectineus</u> (PEC) <u>adductor</u> (STE), <u>Gracilis</u> (GRA), <u>Semimembranosus</u> (SME), <u>Adductor</u> (ADD), <u>Pectineus</u> (PEC) <u>adductor</u> (STE), <u>Gracilis</u> (GRA), <u>Semimembranosus</u> (SME), <u>Adductor</u> (ADD), <u>Pectineus</u> (PEC) <u>adductor</u> (STE), <u>Gracilis</u> (GRA), <u>Semimembranosus</u> (SME), <u>Adductor</u> (ADD), <u>Pectineus</u> (PEC) <u>adductor</u> (STE), <u>Gracilis</u> (GRA), <u>Semimembranosus</u> (SME), <u>Adductor</u> (ADD), <u>Pectineus</u> (PEC) <u>adductor</u> (STE), <u>Gracilis</u> (GRA), <u>Semimembranosus</u> (SME), <u>Adductor</u> (ADD), <u>Pectineus</u> (PEC) Adductor (ADS), <u>Gracilis</u> (GRA), <u>Semimembranosus</u> (SME), <u>Adductor</u> (ADS), <u>Fibularis</u> (STE), <u>Gracilis</u> (GRA), <u>Semimembranosus</u> (SME), <u>Adductor</u> (ADS), <u>Fibularis</u> (SAR). Distal pelvic limb (G2): <u>Gastrocnemius et soleus</u> (GAS), <u>Fibularis</u> (SAR). Distal pelvic limb (G2): <u>Gastrocnemius et soleus</u> (GAS), <u>Fibularis</u> (SAR). Distal pelvic limb (G2): <u>Gastrocnemius et soleus</u> (GAS), <u>Fibularis</u> (SAR). Distal pelvic limb (G2): <u>Gastrocnemius et soleus</u> (GAS), <u>Fibularis</u> (SAR). Distal pelvic limb (G2): <u>Gastrocnemius et soleus</u> (GAS), <u>Fibularis</u> (SAR). Distal pelvic limb (G2): <u>Gastrocnemius et soleus</u> (GAS), <u>Fibularis</u> (SAR). Distal pelvic limb (G2): <u>Gastrocnemius et soleus</u> (GAS), <u>Fibularis</u> Supers: (STE), <u>Gracilis</u> (GRA), <u>Semimemorane</u> (GAS), <u>Supers</u> (SAR). Distal pelvic limb (G2): <u>Gastrocnemius et soleus</u> (GAS), <u>set estensor digitorum lateralis</u> (EPL) and <u>Flexor</u> (SAR). <u>Supers</u> (SAR). Distal pelvic limb (G2): <u>Gastrocnemius et soleus</u> (GAS), <u>set estensor digitorum lateralis</u> (EPL) and <u>Flexor</u> (SAR). <u>Supers</u> (SAR). Distal pelvic limb (G2): <u>Gastrocnemius et soleus</u> (GAS), <u>set estensor digitorum lateralis</u> (EPL) and <u>Flexor</u> (SAR). <u>Supers</u> (SAR). Distal pelvic limb (G2): <u>Gastrocnemius et soleus</u> (GAS), <u>set estensor digitorum lateralis</u> (EPL) and <u>Flexor</u> (SAR). <u>Supers</u> (SAR). Distal pelvic limb (G2): <u>Gastrocnemius et soleus</u> (GAS), <u>set estensor digitorum lateralis</u> (EPL) and <u>Flexor</u> (SAR). <u>Supers</u> (SAR). Distal pelvic limb (G2): <u>Gastrocnemius et soleus</u> (GAS), <u>set estensor digitorum lateralis</u> (EPL) and <u>Flexor</u> (SAR). <u>Supers</u> (SAR). <u>Supers</u> (SAR). Distal pelvic limb (G2). (MA) and <u>Superficialis et profundus & Fibularis longus</u> (FPL). Spinal (G3): <u>Psoas major</u> (OEA), Abdominal (G4): <u>Obliguus externus abdominis</u> (OEA), (NEA). Alensor digitorum longus & Extensor and Longissimus dorsi (LDO). Abdominal (G4): Obliguus externus abdominis (OEA). Transversus abdominis (TRA) and Rectus abdominis (REA). Transversus abdominis (TRA) and Rectus abdominis (TBR). ^{toximal} internus abdominis (OIA), Transversus abdominis (TRA) and <u>Rectus abdominis</u> (TBR), thoracic limb (G5): <u>Deltoides</u> (DEL), <u>Infraspinatus</u> (INF),<u>Triceps brachii</u> (TBR),

Teres major (TEM), <u>Supraspinatus</u> (SUP), <u>Biceps brachii</u> (BIB) and <u>Subcapularis</u> (SUB), ^D thoracic limb (G6): <u>Extensor carpi radialis</u>, <u>Ulnaris lateralis</u>, <u>Abductor digiti</u> Extensor digitorum communis et lateralis (ETL) and <u>Flexor carpi radialis et lateralis</u> The data were processed statistically by the NWASTATPAK Program Northwest Analytic

Inc. Portland Oregon USA.

Results and Discussion

The individual IMF content as a percent of total IMF in each anatomical muscle in and F pigs are given in Figures 1. Content of total IMF in each anatomical muscle in the second for L and F pigs are given in Figures 1, 2, 3 and 4. Significant differences in pice tribution between the L and F pigs were detected. A significant accumulation $(F_{\mu\nu})$ IMF fat occurs in Mm BFE and GLU in F pigs. IMF fat occurs in Mm BFE and GLU in F pigs and in GRA, SME and ADD in L pigs in the production (F production) (Fig. 1). pelvic limb muscle group (Fig. 1). In spite of the big SD in the abdominal muscle group (Fig. 1). fat content shows the most important changes, specially in Mm OIA and REA (p < .05) weight Differences in the other muscle groups were less significant. The total IMF fat weight each anatomical muscle group as a percenter of the show each anatomical muscle group as a percentage of total IMF in the L and F pigs is the figure 5. Statistical differences (not contact to the land F pigs is the land F between the groups are shown for groups 1, 2 and 3. The total amount of IMF fat was almost in the distribution of IMF fat was almost in the total amount of IMF fat was almost to the total amount of IMF f 0.43 and 0.41 kg in spite of big differences in carcass fat. This results explain ju Hiner et al., 1965) a and the difficulties in the estimation of the effects of vertices and the difficulties in the estimation of the effects of vertices and the the terms of the effects of the terms of terms of the terms of term correlation found between total body fat and IMF for many authors (Duniec et al. like breed, sex, diet on the IMF content fat in a given muscle taking in account fatness or any predictor indicator of carcass fatness as a back fat thickeness of IMF COR increasing incidence of meat quality problems requires new methods of meat prediction (Kempster et al 1986).

The IMF% in the muscles from the two lines are given in Figure 1, 2, 3 and 4. The higher in the F pigs compared with the were higher in the F pigs compared with the L ones except for the Mm VL^A , SME and extension (Group 1); OIA and TRA (Group 4) and extension of

The levels of linoleic acid (18:2) in the muscle lipids were always higher in the fat pigs (Fig.6). than in the fat pigs (Fig.6). The values were from 6.3 to 13.4% in the lean pigs in the set of 10.9% in the fat pigs. The % of 10.0% IMF % il muscle but also at similar IMF% the F pigs have less 18:2 in total lipids than $(0,9)^{[1]}$. No differences in the average daily gain between the two lines were detected 0.929 g for L and F pigs) and then the difference 0.929 g for L and F pigs) and then the difference may be due to the different contribution of the lipid classes to the total muscular lipid. of the lipid classes to the total muscular lipids. At similar diet the firmest observed when growth rate and fat thickness were high (Wood, 1983).

Conclusions

Differences in the distribution of intramuscular fat between lean and fat pick found. The total amount of intramuscular fat maximuscular fat between lean and fat representation of intramuscular fat was similar in both lines. The fatty of th position of intramuscular lipids showed a strong tendency for a higher percent

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a).^{phition} of intramuscular fat in Duroc Jersey and Hampshire pigs at 100 kg live-weight.

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Fig. 1. IMF% and IMF weights as a % of total IMF in the number of muscle group. IMF in the anatomical muscle group. 35 2

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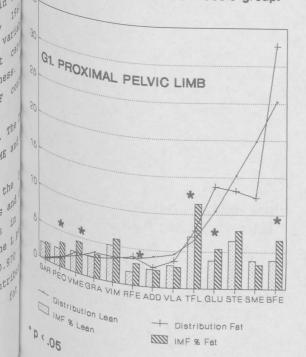
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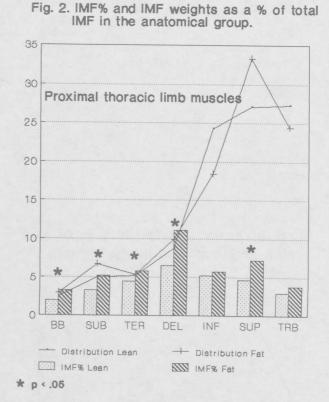
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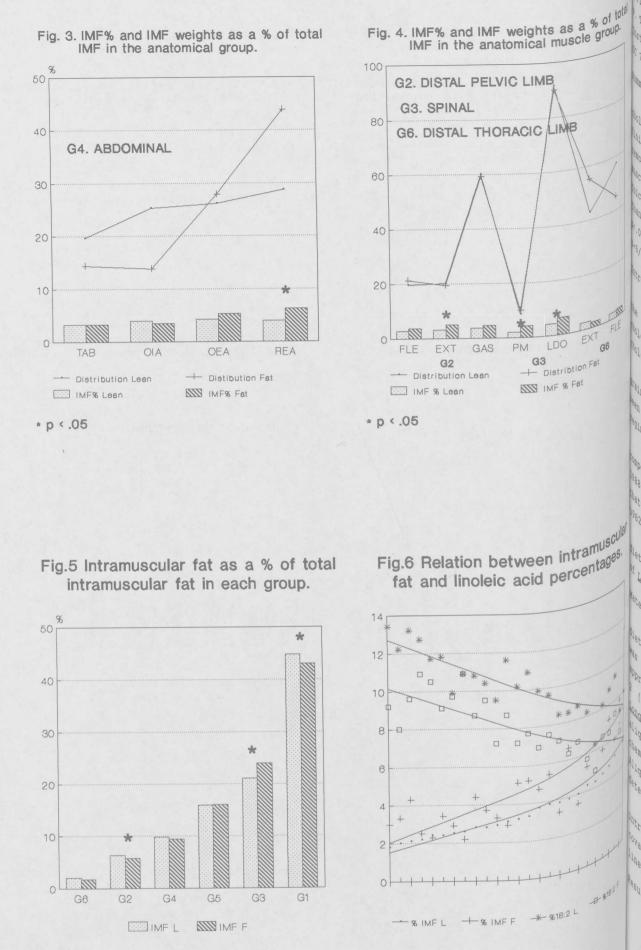
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* p< .05.

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