COMPOSITION OF CARCASS TISSUE FROM TRANSGENIC PIGS <sup>80LOMON<sup>1</sup></sup>, V.G. PURSEL<sup>2</sup> and E.W. PAROCZAY<sup>1</sup> Dept. of Agr., ARS, Beltsville, Maryland 20705-2350 USA <sup>Res.</sup> Lab., PQDI, <sup>2</sup>Gene Eval. and Mapping Lab., LPSI

builtand lipid, fatty acid profiles and cholesterol content of whole carcass ground tissue <sup>Compared</sup> for transgenic (T) pigs expressing a bovine growth hormone gene and control (C) <sup>Pigs</sup> were slaughtered at five different weights: 14, 28, 48, 68 and 92 kg. Total <sup>35</sup> Were slaughtered at five different werghter. I., <sup>46</sup> fat followed a different pattern of deposition in T-pigs than in C-pigs. Carcass fat With increasing live weight in T-pigs compared to a continuous increase (carcass in C-pigs. A dilution effect on fatty acids was observed in T-pigs with increasing live <sup>P4gs.</sup> A dilution effect on fatty acros was outer <sup>Whereas</sup>, fatty acid concentrations increased in C-pigs. Cholesterol content was not Werens, fatty acid concentration... between T- and C-pigs at any of the slaughter weights.

INTRODUCTION. Introducing recombinant genes into laboratory animals has been introducing recombinant genes into laboratory animals has been <sup>Aunology</sup> for introducing recombinant genes into further the first successful only since 1980. Gene transfer techniques have evolved from the first success in mice <sup>transfer</sup> experiments carried out in mice in 1980 (GORDON et al, 1980). Success in mice Med similar experiments in farm animals. HAMMER et al (1985) described the first <sup>Autilar</sup> experiments in farm animals. HARTER Strength of the gene transfer <sup>Minents</sup> Carried out with farm animals during the late 1980s attempted to manipulate and related production characteristics such as feed efficiency. However, little is <sup>related</sup> production characteristics such as recursion of transgenic livestock. <sup>yarding</sup> body/carcass composition and lipid composition. <sup>v the</sup> purpose of this study was to determine what effective <sup>bovine</sup> growth hormone (rbGH) gene into pigs has on lipid and carcass composition.

MATERIALS AND METHODS Margenia and Feed. Fifty-two transgenic and control pigs were G2 generation descendants of transgenic founder 37-06. founder 31-04 and G3 and G4 generation descendants of transformed ova a Winders of these two transgenic lines were produced by microinjection of fertilized ova <sup>Malers</sup> of these two transgenic lines were produced by Micross, <sup>Maler</sup> b<sub>GH</sub> fusion gene (PURSEL et al, 1989). Subsequent generations were produced by <sup>MI-bGH</sup> fusion gene (PURSEL et al, 1989). Subsequent generation <sup>Insemination</sup> of non-transgenic crossbred females with fresh or frozen epididymal <sup>Ad insemination of non-transgenic crossbred females with free of the transgene was <sup>Ablished</sup>. Presence of the transgene was <sup>Ablished</sup>. (HAMMER et al, 1985).</sup> <sup>veoa</sup> <sup>rec</sup>overed from transgenic boars after euthanasia. Fresente <sup>heg</sup>ished by hybridization of dot blots of DNA from tail biopsies (HAMMER et al, 1985). <sup>Aled</sup> by hybridization of dot blots of DNA from tail blopsles (manned) <sup>Aleg</sup> of the transgene was established by radioimmunoassay of plasma collected at 1 week <sup>Aleg</sup> or of the transgene was established by radioimmunoassay of plasma collected at 1 week <sup>Aleg</sup> or of the transgene was established by radioimmunoassay of plasma collected at 1 week <sup>on of the</sup> transgene was established by radioimmunoassay of pro-<sup>on older</sup> using a bGH radioimmunoassay that did not measure pGH (MILLER et al, 1989). <sup>WH concentrations were >1000 ng/ml in transgenic pigs of the second and fed httransgenic pigs of the 37-06 line. Pigs were weaned at 4 to 8 weeks of age and fed</sup> Mole Carcass grinder and random samples of ground tissue were collected. Tissue samples

were analyzed for proximate-chemical composition, fatty acid composition, and cholestern content.

Isolation of Lipids. The FOLCH et al (1957) procedure was employed to obtain lipid extracts of each sample. Preparation and isolation procedures were previously described to be a solution procedure was employed to obtain the solution and isolation procedures were previously described to be a solution and the solution procedure was employed to obtain the solution and the solution procedure was employed to obtain the solution and the solution procedure was employed to obtain the solution and the solution procedure was employed to obtain the solution and the solution procedure was employed to obtain the solution and the solution procedure was employed to obtain the solution and the solution procedure was employed to obtain the solution proced SOLOMON et al (1990). Fatty acids were converted into methyl ester derivatives prior to analysis. Methylation and analysis. Methylation and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously described and preparation of samples for GLC analysis were previously desc (SOLOMON et al, 1990). Cholesterol was converted to a trimethylsilyl ether derivative. derivatizing procedure and sample preparation were previously described (SOLOMON et al 1990).

Data Analysis. Data were analyzed by the analysis of variance technique (SAS, 1985) for any second state of the significance of manine the significance of m determine the significance of variation between transgenic and control pigs at the five different weight groups for a 2 x 5 factorial arrangement which included treatment x weight group interactions. When signific group interactions. When significant (P<.05) main effects for weight groups were detected

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Carcass fat deposition followed a different pattern for T-pigs compared to C-pigs (Table . . Total carcass lipid increased (1993) 1). Total carcass lipid increased (190%) in the C-pigs from 14 to 92 kg live wt, whereas the T-pigs, total carcass lipid. the T-pigs, total carcass lipid increased (32%) only from 14 to 48 kg live weight and decreased (45%) from 48 to 92 km ( decreased (45%) from 48 to 92 kg (overall 27% decrease from 14 to 92 kg). At 14 kg slaugh weight, carcasses from T-pigs contained action weight, carcasses from T-pigs contained 38% less fat than C-pigs. At 28 kg, T-pigs had that the state of the less total carcass fat; at 48 kg, 48% less total carcass fat; at 68 kg, 78% less total carcass fat; at 68 kg, 78% less total carcass fat, and at 92 kg, T-pigs contained 85% less carcass fat than C-pigs. cholesterol content of the ground carcass tissue was not different between T- and C-pigs any of the designated weights; however with any of the designated weights; however, with increasing live weight (14 to 92  $^{kg)\prime}$ cholesterol content decreased for both C- and T-pigs (23% C-pigs; 28% T-pigs).

Carcass from T-pigs consistently contained less total saturated fatty acids (SFA) pigs at each slaughter weight. At 14 kg slaughter weight, carcasses from T-pigs and registration for the start of the44% less SFA compared to C-pigs. At 28 kg, T-pigs had 42% less SFA; at 48 kg, T-pigs from T-pigs stat 48 kg, T-pigs had 78% less SFA; at 68 kg, T-pigs had 78% less SFA; less SFA; at 68 kg, T-pigs had 78% less SFA, and at 92 kg, T-pigs contained 85% less sFA; at 48 kg, T-pigs reflects C-pigs. Carcass tissue from T-pigs reflects a much more favorable (lesser quantity) of start and a 137% increase in total SFA was observed in a second A 137% increase in total SFA was observed in C-pigs with increasing live wt (14 to 92 kg) compared to a 40% decrease in total SFA former former former and to a 40% decrease in total SFA former former former and the state of th compared to a 40% decrease in total SFA for T-pigs. These differences in SFA were not form, at form, result of changes in palmitic acid, stearic acid, and myristic acid (not in tabular form). Margaric and arachidic acids were also present. Margaric and arachidic acids were also present but in very small quantities; however, 62% of the formation o did not contain any arachidic (not in tabular form). Palmitic acid accounted for 62% of b total detectable SFA in the C-pigs (92 bo with Stearic acid accounted for 34% of the total SFA in the C-pigs (92 kg weight group) compared to 66% in T-pigs. Both myristic acid and palmitic acid because the state of the total sector acid because the state of the state of the total sector acid because the state of the st T-pigs. Both myristic acid and palmitic acid have been reported to be hyperlipidemic fatty fatty and fatty f hypercholesterolemic (KEYS et al, 1965). Human consumption of hypercholesterolemic (<sup>fatty</sup> acids has come under attack by health profession acids has come under attack by health professionals (NRC, 1988). BONAMONE and GRUNDY (1996)

"strated that a diet high in stearic acid did not elevate plasma levels of low density Motein (LDL) cholesterol, perhaps because it is poorly digested and can be easily Whated to oleic acid (KEYS et al, 1965).

to oleic acid (KEYS et al, 1965). <sup>A to</sup> oleic acid (KEYS et al, 1965). <sup>A to</sup> oleic acid (MUFA) than C-pigs <sup>A to</sup> oleic acids (MUFA) than C-pigs <sup>to Non T</sup>-pigs contained less total monouncer weight, carcasses from T-pigs slaughter weight (Table 1). At 14 kg slaughter weight, carcasses from T-pigs <sup>48</sup>% less MUFA; at 28 kg, 46% less MUFA; at 48 kg, 59% less MUFA; at 68 kg, 79% less <sup>48%</sup> less MUFA; at 28 kg, 46% less MUFA, at 10 million of the MUFA were the at 92 kg, 91% less MUFA than C-pigs. The fatty acids comprising the MUFA were <sup>Ac 92</sup> kg, 91% less MUFA than C-pigs. The factor <sup>boleic</sup>, palmitelaidic, oleic, and cis-11-eicosenoic acids. Oleic acid was the major Multiple of MUFA in the total lipid fraction (not in tabular form). The trend was for the With of MUFA in the total lipid fraction (not in tasks <sup>Was</sup> <sup>Observed</sup> for MUFA in T-pigs after 48 kg live weight. Oleic acid has been reported <sup>AS observed</sup> for MUFA in T-pigs after 48 kg ive more that is therefore not considered hypolipidemic, reducing cholesterol (GRUNDY, 1986), and it is therefore not considered that oleic an Undesirable dietary fatty acid. MATTSON and GRUNDY (1985) demonstrated that oleic the ability to reduce LDL cholesterol.

An ability to reduce LDL cholesteron. in the carcass tissue (Table 1) from T- and C- pigs. At 14 kg slaughter weight, An the carcass tissue (Table 1) from T- and C- Pige. point from T-pigs contained 38% less PUFA; at 28 kg, 24% less PUFA; at 48 kg, 22% less purch than C-pigs. The trend was for to it from T-pigs contained 38% less PUFA; at 28 kg, 24% 1000 The trend was for the <sup>108</sup> kg, 53% less PUFA, and at 92 kg, 66% less form the decrease with increasing increase with increasing live weight in C-pigs and to decrease with live weight increase weight live weight increase weight live weigh <sup>increase</sup> with increasing live weight in C-pigs and the stand the

<u>CONCLUSIONS</u> <u>Conclusions</u> Conclusions Co <sup>veheral</sup>, carcass tissue from T-pigs contained significantly ress <sup>veher</sup> <sup>Plgs</sup> at each of the designated slaughtered weights. In such a standard for fatty acid deposition in T-pigs with increasing slaughter weight. On the an <sup>for fatty</sup> acid deposition in T-pigs with increasing backs, <sup>fatty</sup> acid deposition either increased or remained the same with increasing The weight in C-pigs. Carcass tissue from T-pigs reflects much more favorable levels <sup>add Muty acids</sup> compared to C-pigs when considering dietary health recommendations and

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Treatment	Weight Group, kg	Total Lipid g/100g	Cholesterol mg/100g	Total SFA	Tota. MUFA
Controls	14	10.04	100.88	3.24	3.74
	28	12.32	94.97	3.69	4.69
	48	16.58	85.92	5.37	6.73
	68	26.78	74.15	8.22	8.74
	92	29.07	77.81	7.67	8.73
Transgenics	14	6 10	106 45	1 92	1.98
	28	7 62	99 97	2.20	2.47
	48	8.16	86.28	2.39	2.50
	68	5.97	74.55	1.60	1.57
	92	4.49	77.08	1.15	1.13
Significance	of treatments	5. P<:			
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Treatment (T)		.01	NS	.01	.01
Weight Group (W)		.01	.01	.01	.01
TXW		.01	NS	.01	.01

TABLE 1. COMPARISON OF TOTAL CARCASS LIPID, CHOLESTEROL AND FATTY ACID<sup>a</sup> CONTENT FOR TRANSGENIC<sup>b</sup> AND CONTROL PIGS

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<sup>a</sup>Grams in 100g sample. SFA = saturated fatty acids; MUFA = monounsaturat PUFA = polyunsaturated fatty acids. <sup>b</sup>Transgenic pigs = pigs expressing bovine growth hormone gene.