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SUMMARY: The experiment was conducted to study the metabolism of 2,3-DPG and pyruvate kinase (PK) and mitochondrial ATPase activities in PSS pig. LYD three crossbred pigs were screened into normal and PSS pigs with Halothane test, which were used as test animals. The muscle samples obtained from the stressed animals for determining DPG and PK activity before slaughtered, and Longissimus dorsi muscle was used for measuring mitochondrial ATPase activity 30 min postmortem.

The concentration of DPG in the blood in the PSS pig was lower than that of the normal pig (6.70 ± 0.25 and 7.16 ± 0.19 , respectively). The concentration of DPG in the blood of the PSS pig was lower than that of the normal pig. This result was in agreement with the structure of DPG changed or dissociated in the blood of the PSS pig. PK activity in the PSS pig was higher than in the normal animals. However, no difference in mitochondrial ATPase was found in the stressed and the normal pigs. The result was also found there was difference in DPG concentration between male and female pigs in the groups, but no difference in PK activity was detected in different sex.

INTRODUCTION: Since DPG is very important factor for maintaining normal physiological function of oxygen transportation. It can bind with hemoglobin and lower the affinity of hemoglobin and oxygen under normal condition. As blood flow through the muscle HbO_2 release O_2 to muscle for aerobic metabolism. However, when the chemical structure and concentration of DPG change, it may retard HbO_2 release O_2 to muscle and blood (Chiba and Sasaki, 1978). The phenomenon is similar to the findings of PSS pigs when they are suffered stress. Meanwhile, glycolysis takes place very fast in PSS pigs. Whether this is caused by Pyruvate Kinase action, since it is a very important enzyme for glycolysis. So its activity should be studied. And the ATP depletes very fast in PSS porcine muscle thus ATPase in mitochondria is also measured (Yang et al., 1991). In the past years, research workers all stressed on work of catecholamine, corticosteroid and halothane test and CPK assay (Cassens et al., 1975). Since no differences were found in the concentrations of catecholamine, corticosteroid between normal and PSS pigs. Thus, the purpose of this study is to use ^{31}P -NMR and spectrophotometry and biochemical measurements to study the 2, 3-diphosphoglycerate, pyruvate kinase and mitochondrial ATPase activities of PSS pigs, as compared to the normal pigs.

MATERIALS AND METHODS:

Longissimus dorsi muscle and blood specimen were obtained from the normal and PSS pigs which were slaughtered at local slaughterhouse. The muscle specimen added with anticoagulant-heparine and EDTA for measuring DPG and PK. The muscle samples were placed in liquid nitrogen for PK activity measurement. DPG content was determined according to the method described by de Verdier and Ericson (1981), and NMR-spectra was measured with the procedures of Fujii and Miwa (1984). NMR-spectra were measured by NMR-spectrophotometer (7.05 Tesla, Varian Instrument Ltd, USA). Specimen of venous blood was collected in heparinized tubes and mixed with D_2O and keep the tubes at room temperature for NMR-spectra measurement. Mitochondria isolation and mitochondrial ATPase activity were carried out according to the procedures described by Cain and Skilleter (1987).

RESULTS AND DISCUSSION:

The pH of blood from PSS pigs was lower than that of the normal pigs. Blood proteins has buffering ability so it pH does not drop below 5.8 like the blood of the PSS pigs. The pH of the normal pig was 7.16 and the PSS pig was 6.70 in the blood (Table 1).

The NMR-spectra for NMR-spectra analysis was collected from the PSS and normal pigs which were suffered stress with electrical shock. From the NMR spectra of blood, it could be observed the structure of 2,3-DPG was changed in the blood of PSS pigs after stress, but not observed in the blood of the normal pigs (Fig. 1). DPG which was shown in Fig. 1 indicated the structure of PSS pigs stressed with electrical shock seemed dissociated or might be converted into glycerol-3-phosphate.

The 2,3-diphosphoglycerate (DPG) content in the blood of the PSS pigs was averaged at 3.885 mmole per liter of blood (male 3.40, female 4.17 mmole/l) which was lower than the blood of the normal pigs with 5.925 mmole/l (male 5.71, female 6.15 mmole/l) (Table 1). The result indicated that the DPG content in female pig was higher than that of the male pigs both the normal and PSS pigs. This result was in accordance with the structure or NMR-spectra of DPG changed or dissociated in the blood of the PSS pigs shown in the Fig. 1.

The glycolytic enzymes were regulated mainly by three enzymes---hexokinase, phosphofructokinase, and pyruvate kinase which were responsible for the regulation of glycolysis. Therefore, one of these three major enzymes, pyruvate kinase activity in blood of PSS pigs was measured in this experiment. Pyruvate kinase activity in the blood of the PSS pigs was higher than that of the normal pigs. They

were 41.9 unit/ml red cell for male and 45.1 unit/ml red cell for female; and the normal pigs were 20.4 unit/ml red cell for male and 19.4 unit/ml red cell for female (Table 1).

Pyruvate kinase activity can catalyze the conversion of PEP to pyruvate with regeneration of ATP. Since ATP is also important for the maintenance of cell integrity, it is not surprising that highly sophisticated mechanisms for regulating levels of adenine nucleotides have evolved. As was stated above, the importance of ATP to energy metabolism of red cell, whether ATPase activity was related to physiological properties of muscle from PSS pig and normal pigs. Therefore, mitochondrial ATPase activity in muscle for the normal and PSS pigs was measured. The mitochondrial ATPase activity in normal pork was higher than that of PSE pork. They were 0.1575 $\mu\text{mole pi/min/mg protein}$ and 0.1509 $\mu\text{mole pi/min/mg protein}$ for the normal pork and PSE pork, respectively. No difference was detected between the normal and PSE pork. In conclusion, pH, DPG content of blood of PSS pig were lower than the normal pig, but PK activity of the blood in PSS pig was higher than that of the blood in the normal pig, however, no difference in mitochondrial ATPase in muscle was detected between the normal and PSS pigs.

REFERENCES:

Cassens, R. G., Marple, D. N., and Eikelenboom, G., 1975. Animal physiology and meat quality. *Adv. Food Res.*, 21, 71-155.

Chiba, H., and Sasaki, R., 1978. Functions of 2,3-bisphosphoglycerate and its metabolism. *Current topics in cellular regulation*, 14, 75-95.

Cain, K. and Skilleter, D. N., 1987. Preparation and use of mitochondria in toxicological research, in "Biochemical Toxicology-A Practical Approach" edited by by Snell, K., and Mullock, B., IRL PRESS, Washington, D. C., USA.

Fujii, H., and Miwa, S., 1983. "Pyruvate kinase assay in serum and erythrocytes" in Bergmeyer's Methods of Enzymatic Analysis. 3rd ed., Vol. III., pp. 496, Verlag Chemie, Weinheim, pp. 496.

de Verdier, C. H., and Ericson, A., 1984. "D-Glycerate 2,3-Bisphosphate" in Bergmeyer's Methods of Enzymatic Analysis. 3rd ed., pp. 547-552, Verlag Chemie, Weinheim.

Yang, W. D., Chen, M. T., and Liu, D. C., 1991. Study on biochemical properties of blood of PSS pigs by ^{31}P -NMR. *J. Chinese Animal Sci.* 20, 217-226.

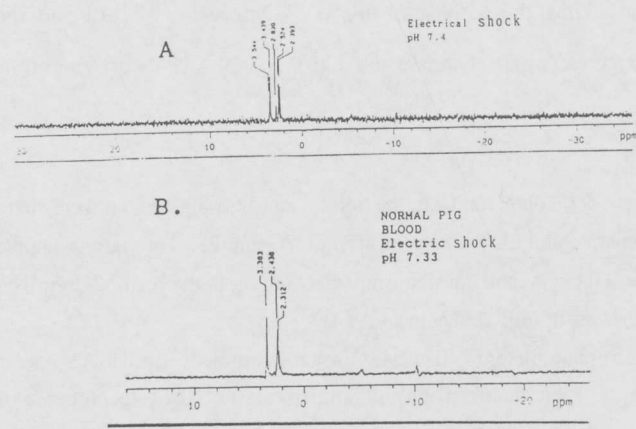


Fig 1 ^{31}P -NMR spectra of blood from PSS and the normal pig (A)PSS (B)normal.

Table 1. Some selected Biochemical characteristics of blood and mitochondrial ATPase in muscle of the normal and PSS pig.

	PH	ATPase $\mu\text{mole pi/min/mg prot.}$	DPG mmole/l blood	PK unit/ml RBC
Normal	7.16 ± 0.19^a	0.1575 ± 0.0080	δ 5.71 ± 0.20^{ab}	20.4 ± 1.3^a
			ϕ 6.15 ± 0.09^a	19.8 ± 1.8^a
PSS	6.70 ± 0.25	0.1510 ± 0.0105	δ 3.40 ± 0.14^b	41.9 ± 3.3
			ϕ 4.07 ± 0.19	45.1 ± 3.8

Value given are the mean \pm the standard error

^athere are significantly different between two groups ($p < 0.01$)
^bthere are significantly different between male and female ($p < 0.01$)