

ULTRASONICATION OF LAMB SKELETAL MUSCLE FIBRES ENHANCES POSTMORTEM PROTEOLYSIS.

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SUMMARY. The ability of ultrasound treatments to disrupt membranes and to release enzymes from cellular compartments is well known. The aim of our work was to investigate if such a treatment would be able to activate endogenous proteolysis in muscle fibres. We demonstrated that ultrasonication was indeed effective in releasing lysosomal enzymes from liver cells, as shown by the presence of acid phosphatase activity in cytoplasm extracts. Cell membranes were damaged only to a limited extent, as revealed by turbidity measurement of the medium. Proteolysis brought about by endogenous proteinases, after two days of fibre storage at 4°C, was assessed by means of SDS-PAGE in controls as well as in fibres treated with ultrasounds varying in intensity and time of exposition. The incipient appearance of a series of faint bands in the 30000 Da region was the only evident change of control gels. Results revealed that ultrasonic treatment of fibres enhanced proteolytic degradation, depending on time and output of sonication, as shown by the increased intensity of 30000 Da bands. A distinct change featuring sonicated fibres was the degradation of a 87000 Da protein and the appearance of a 83000 Da band. It is discussed whether calpains or cathepsins are to be responsible for observed changes. Cell damage appeared to be not very severe, although it was dependent upon ultrasonication conditions. As a consequence, ultrasonication of muscle fibres may be regarded as a good method for increasing postmortem proteolysis, and thus meat tenderisation, despite technical problems derived from ultrasound which must be previously solved.

INTRODUCTION. It is well known that meat becomes more tender during aging, so that postmortem storage of meat is the method currently used for achieving suitable tenderisation. Proteolysis of myofibrillar proteins, mainly of high molecular weight proteins, is thought to be the major contributor to the tenderising process during postmortem storage (Asghar & Bhatti, 1987). Of the endogenous proteases of skeletal muscle, calcium-dependent proteases (calpains) are likely the main responsible for the increase in tenderness during postmortem storage (Koochmaraie et al., 1988a). The cysteine proteinases cathepsins B, H and L and the aspartate proteinase cathepsin D, have received also attention concerning their potential role in postmortem aging (Moeller et al., 1976). However, Zeece & Katoh (1989) suggested that the effectiveness of cathepsins is greatly reduced under low temperature conditions and suggested that these proteases do not play a principal role in the tenderisation process occurring in muscle postmortem.

A number of methods have been proposed to accelerate the tenderising of meat: electrical stimulation (Davey et al., 1976), calcium ionization (Mac Farlane, 1985), infusion of Ca²⁺ (Koochmaraie et al., 1988b) or addition of endogenous and exogenous proteases (Moeller & Mc Dowell, 1987). High temperatures and low pH postmortem exerted a tenderising effect through activation of cathepsins (Moeller et al., 1976). On the contrary, Jaime et al. (1992) reported an increased postmortem proteolysis by effect of low temperature and low pH on muscle calpains. Several methods have been tried out to destabilize the lysosomes and increase lysosomal protease activity in muscle: acid or alkaline treatments, triton X-100, hypertonic solutions of ClNa, pyrogens, osmotic shock and UV radiation (Park & Alliger, 1967). Stagni & Bernard (1968) obtained the same effect by subjecting the muscle to the action of ultrasound. Alliger (1975) reported on this new application of ultrasound, focusing on the release of enzymes and proteins from cells and subcellular particles. On this basis, we have carried out a study conducted to examine the effects of ultrasound on destabilization of lysosomes and other vesicles as well as on the sarcoplasmic reticulum and mitochondria, allowing cathepsins to be released from their compartments as well as increasing free calcium concentration. The main purpose was to increase postmortem myofibrillar protein degradation and thus accelerate meat tenderisation.

MATERIALS & METHODS. **Ultrasonic treatment of liver.** Lamb liver was obtained a day after slaughter and cut in cubes of about 5 g that were immersed in MFI buffer (20 ml) (100 mM KCl, 20 mM KPO₄H₂, 0.1 mM EDTA, 1 mM MgCl₂ and NaN₃, pH 7) and subjected then to ultrasound action for different times of application at 62 w (Branson sonifier, model 250/450). Control consisted of a sample immersed in buffer but not treated with ultrasounds. **Extraction of cell plasma fraction.** After ultrasound treatment liver samples (10-15 ml of MFI buffer) were homogenized with a potter and cell plasma fraction was obtained following the method used for myofibrillar protein extraction (Olson et al., 1976). Finally, supernatant was centrifuged at 30,000 g 60 min. **Acid phosphatase assay.** Acid phosphatase activity of cellular plasma fraction (20 mg of protein, approximately) was measured according to the method described by Trouet (1974) using glycero-phosphate as substrate. **Muscle fibres preparation.** Muscle fibres were obtained from lamb Longissimus muscle 30 min after slaughter. Small groups of fibres (2 cm x 0.5 mm, approximately) were separated from a piece of muscle. The muscle bundles were included in MFI buffer. **Ultrasonic treatment of fibres.** Muscle fibres immersed in MFI buffer were treated with ultrasound at 57 or 62 w for different times (10 s to 3 min) at atmospheric pressure. Muscle fibres were then aged at 4°C for two days. Controls not treated with ultrasounds were aged under the same conditions. After sonication, muscle fibres were centrifuged at 4,000 g 10 min and turbidity of supernatants was measured at 540 nm. **Myofibrillar protein isolation.** Myofibrillar proteins were isolated by the MFI method (Olson et al., 1976) at 3rd day postmortem, boiled in SDS samples buffer (Porzio & Pearson, 1977) and stored at -20°C. **Protein concentration.**

Protein concentration was determined by the Nessler method (Johnson, 1941). Sodium-dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE). SDS-PAGE was performed according to Greaser et al. (1983). Approximately 0.35 mg of protein were loaded on each gel. Samples were runned at 120 V and 250 V through stacking and resolving gel, respectively.

RESULTS & DISCUSSION. As shown in Figure 1, ultrasonication of lamb liver slices was able to destabilize lysosomes in this tissue and thus release their enzymes to cytoplasm. Acid phosphatase activity increased in a first phase (until 7 min of treatment) and decreased thereafter, probably due to ultrasound-caused inactivation. Alliger (1975) already reported that ultrasounds are able of inactivating enzymes after some time of treatment. Absorbance at 540 nm of the supernatant, used as an index of turbidity, increased with time of treatment, being three times higher at 10 min than at 0.5 min. Our results demonstrated that plasmatic membrane damage depended upon time of sonication, since increasing the time of treatment resulted in an important increase of absorbance. This fact suggest that short times should be applied to muscle samples in order to destabilize lysosomes and endoplasmic reticulum while maintaining as intact as possible the plasmatic membrane.

The plasma membrane of skeletal muscle was slightly affected by effect of ultrasound when a low intensity of treatment was used (Figure 2). A constant absorbance at 540 nm was obtained after one minute of application of 56 w. A more severe alteration resulted at 62 w since higher values of turbidity were obtained with small increases of treatment time (six times higher after 30 s of treatment at 62 w than at 56 w). This level of sarcolemma damage was however very little when compared to that caused to muscle samples treated with 56 w after rigor onset; supernatant absorbance was in this case even five times greater than in samples treated at prerigor with the same power. This fact should be referred to the loss of sarcolemma integrity associated to postmortem degradation (Stanley, 1991).

SDS-PAGE of ultrasound treated myofibrils (Figure 3) demonstrated that sonication brought about an activation of proteolysis since several peptides appeared at the region of 30,000 Da (30,500, 29,000, 27,500, 26,000 and 25,000 Da) as soon as after only two days of aging. Proteolytic effects were even magnified with time of treatment. However, only small differences were observed between the two ultrasonication intensities so far used. Surprisingly, no protein of a molecular weight in the range 30-45,000 Da was observed to be degraded, although appearing peptides could proceed from any protein of a high molecular weight not well defined in the gel. Moeller et al. (1976) and Troy & Tarrant (1987) reported an increase of myofibrillar fragmentation in muscle samples electrically stimulated and aged at 4°C. These authors suggested that cathepsins released after treatment could be responsible for proteolytic effects; the appearance of several peptides in the region of 30,000 Da was the most constantly found effect. Results obtained in this work have demonstrated that ultrasonication treatment brought about an increase of meat aging proteolysis. These effects could be the result of either calpains activation by an increase of calcium released from sarcoplasmic reticulum or cathepsins released from lysosomes. Although we do not know thus far what proteases are most likely responsible for the changes observed, cathepsins would probably show a limited proteolytic action due to low temperature and short time of aging (Zeece & Katoh, 1989). It can be also observed in Figure 3 that the intensity of a band corresponding to a 87,000 Da protein decreased at the same time that a new band of 83,000 Da appeared in samples treated with the longest times of ultrasounds. Some researchers have reported the appearance of peptides in this region by action of cathepsins on myosin and α -actinin (Matsukura et al., 1981). Troponin T, desmin and α -actinin have been reported to be degraded by cathepsins D and L (Matsukura et al., 1981), although their degradation was very slow at 4°C and pH 5.5. Zeece & Katoh (1989) also reported that several proteins were sensitive to proteolysis.

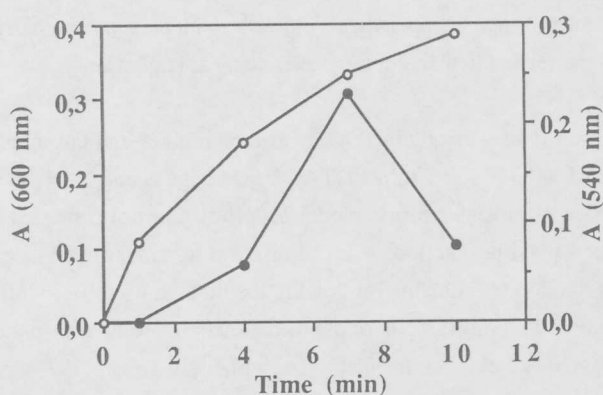


FIGURE 1. Acid phosphatase activity of lamb liver samples treated with ultrasounds for different times (○), expressed as absorbance at 660 nm, and turbidity of immersion buffer of liver samples during sonication (●), expressed as absorbance at 540 nm.

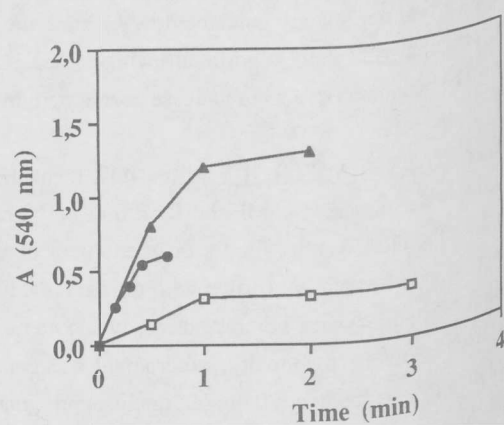


FIGURE 2. Turbidity of immersion buffer used for the sonication of lamb muscle fibres, expressed as absorbance at 540 nm. Treatments: Prerigor, 57 w (□); Prerigor, 62 w (●); Rigor, 56 w (▲).

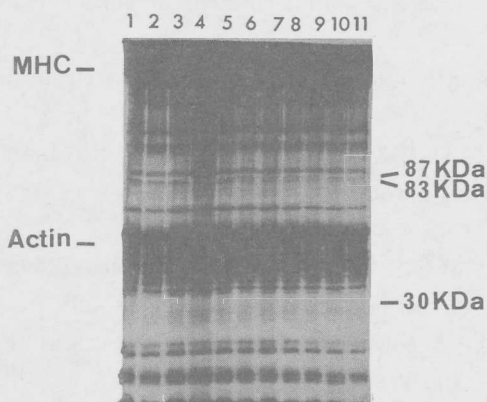


FIGURE 3. 15% SDS-polyacrylamide gels of myofibrils obtained from lamb Longissimus dorsi samples treated with ultrasounds and stored at 4°C during 2 days. 1, 2, 3) controls; 4, 5, 6, 7) samples treated with ultrasounds at 57 w; 4: 30 s, 5: 60 s, 6: 2 min, 7: 4 min; 8, 9, 10, 11) samples treated with ultrasounds at 62 w; 8: 10 s, 9: 18 s, 10: 25 s, 11: 40 s.

proteins action: troponin T, myosin, α -actinin, α -tropomyosin, actin, titin, nebulin and M and C proteins, but the proteolytic effects were nonexistent at 4°C. Some of these proteins have been reported, too, to be sensitive to the action of calpains: troponin T and I, myosin, M and C proteins, titin and nebulin (Etherington, 1984).

Ultrasounds had been shown before to be capable of releasing several enzymes from lysosomes of bovine skeletal muscle to the cytoplasm, such as β -glucuronidase and cathepsins (Stagni & Bernard, 1968). Now, we have demonstrated the same effect of releasing proteolytic enzymes in lamb liver and presumably a similar effect on lamb skeletal muscle since proteolysis brought about at an early phase of aging was greatly increased. Among the methods proposed to increase meat tenderisation, electrical stimulation is currently being the most used. Several authors have suggested that the proteolytic effects associated to electrical stimulation are due to calpains released from lysosomes (Moeller et al., 1976; Troy & Tarrant, 1987), and even to calpains activation by increase of sarcoplasmic calcium levels (Koohmaraie et al., 1988a). Troy & Tarrant (1987) reported the appearance of bands of 39,000 and 32,000 Da at 49-72 h postmortem in lamb muscle electrically stimulated and aged at 4°C. According to these authors proteolytic effects were magnified if time or temperature of aging were increased. Pressurization has been reported, too, as a suitable method to increase meat tenderness by activating calpains and proteolytic effects in this case were also observed at an early stage of aging (Koohmaraie et al., 1984). Results presented in this paper suggest that the proteolytic increase observed after ultrasonication of meat is similar to that caused by electrical stimulation and pressurization at the first moment of aging. On the basis of the high correlation existing between the intensity of peptides in the 30,000 Da region and the tenderising of meat (Penny, 1980), our results suggest that ultrasound treatment could be envisaged as a novel method for producing tender meat after a short aging time. However, the results presented here are a first approach to the subject and subsequent research is obviously necessary, particularly concerning the practical application to meat cuts and carcasses.

CONCLUSIONS. SDS-PAGE of muscle fibres subjected to ultrasonication and stored thereafter two days at 4°C demonstrated that this treatment significantly enhanced postmortem proteolysis, as revealed by an increased intensity of peptide bands in the region of 30,000 Da, while fibre membranes were damaged only to a limited extent.

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