CONTROL OF <u>Listeria monocytogenes</u> IN MEAT PRODUCTS BY A STRAIN OF <u>Lactobacillus plantarum</u> M.CAMPANINI, I.PEDRAZZONI, S.BARBUTI AND P.BALDINI ^{Experimental} Station for the Food Preserving Industry - Via Tanara 31/A - Parma - Italy

SUMMARY - It was shown that <u>Lactobacillus plantarum</u> MCS isolated from salami produced inhibitor compounds towards <u>Listeria monocytogenes</u>. In a pH-adjusted culture supernatant, the growth of <u>Listeria</u>, monitored by ^{impedometric} method at 30°C, was delayed at pH 6.0, while no growth occurred at pH 5.5.

A mutant MCS1 strain showed reduced inhibitor activity. After 6-7 days growth of inoculated <u>Listeria</u> was observed in minced raw pork with added curing agents stored at 18°C and in salami. Therefore the addition of <u>L. plantarum</u> strains prevented <u>Listeria</u> growth during the first few days of meat storage and salami maturation. <u>Listeria</u> counts tended to decrease after 6 and 14 days in all minced meat and salami samples, respectively. At 31 days small differences were observed in the survival of listerias in salami inoculated with <u>L. plantarum</u> MCS or MCS1 (bacteriocin-negative strain). The inactivation of <u>Listeria</u> was much less effective in meat products than in culture systems, but inoculated <u>Lactobacillus</u> strains prevent L. monocytogenes growth.

INTRODUCTION - Listeria monocytogenes has been shown to occur frequently in raw meat and has also been isolated from cured and fermented products (SCHMIDT et al, 1988, KARCKES and TEUFEL, 1988, FARBER et al, 1988, BARBUTI et al, 1989, FARBER et al, 1989, JOHNSON et al, 1990). A potential means of preserving fermented meats from this pathogen is the use of bacteriocin-producing lactic-acid bacteria as starter cultures.

^{Inh}ibition of <u>L. monocytogenes</u> by <u>Pediococcus</u> spp.and some <u>Lactobacillus</u> strains was observed not only in culture ^{media}, but also in meat and meat products (HOOVER et al, 1989, SCHILLINGER and LUCKE, 1989, BERRY et al, 1990, ^{SCHILLINGER} and LUCKE, 1990, SCHILLINGER et al, 1991, BERRY et al, 1991). Strains of <u>Lactobacillus</u> spp. ^{iso}lated from dry Italian salami were found to have anti-listeria activity in culture medium (CAMPANINI et al, 1991).

^{The} aim of this study was to assess the bactericidal activity of a <u>L. plantarum</u> strain against <u>L.monocytogenes</u> in a culture ^{Ine}dium, in minced meat and in salami during maturation.

MATERIALS AND METHODS

Microrganisms

L. plantarum MCS isolated from Italian salami and previously selected for anti-listeria activity, was grown in APT ^{medium} at 30°C for 24 h. To produce bacteriocin-negative variant (MCS1 strain) the MCS strain was cultured several ^{times} in MRS broth with acriflavine (10 micrograms/ml) at 30°C for 24 h.

<u>L. monocytogenes</u> strains 38 and 150 isolated from salami were grown in Brain heart infusion broth at 30°C for 20 h. Antagonistic activity in culture media

Impedometric method: The growth of <u>L. monocytogenes</u> 38 was evaluated by capacitance measurements using a Bactometer Instrument (Bactomatic). The growth of listerias $(10^3 - 10^4 \text{ cells/ml})$ was monitored at 30°C in APT broth as ^{control} and in filtered (0.2 micron) pH adjusted (6.0 and 5.5) supernatants from 24 h-cultures of <u>Lactobacillus</u> MCS and MCS1 strains. To eliminate the effect of lactic-acid, the APT broth controls were acidified to pH 4.4 with lactic acid, then ^{adjusted} to 5.5 and 6.0 with NaOH solution. The growth of <u>Listeria</u> was also monitored in supernatant of MCS strain at pH 6.0 with added protease (1 mg/ml).

The anti-listeria activity was verified at different incubation times in the same culture media by plate count on ^{Tr}ypticase soy agar and Palcam.

Agar diffusion assay: In the agar spot test the <u>Lactobacillus</u> strains were dotted onto APT agar and after 24 h-^{anaerobic} incubation at 30°C, the agar plates were then coated with 8 ml of soft APT agar (0.7%) inoculated with 0.2 ml ^{of} an overnight culture of <u>L. monocytogenes</u> strains. After 24 h-incubation at 30°C the antagonistic activity was evident ^{from} the appearance of a clear inhibition area around the spot colony.

Inoculation experiments

- Minced meat: it was prepared using raw pork mixed with NaCl (3 %), sodium nitrite (150 mg/kg) and sucrose (0.3%). Portions of 30 g each were inoculated with MCS strain (10⁶ - 10⁷ cells/g) and/or <u>L. monocytogenes</u> 38 (10³-10⁴ cells/g) and placed into sterile plastic bags; after vacuum heat sealing the bags were stored for up to 14 days at 18°C.

- Salami: fresh salami mix containing 2.5 % NaCl salt, 250 mg/kg potassium nitrate and 0.3 % sucrose was inoculated with 24-h cultures of <u>Lactobacillus</u> MCS and MCS1 strains (about 10⁷ cells/g) and/or two strains of <u>L.monocytogenes</u> (10³. 10⁴ cells/g). After filling into cellulose casings of 50 mm diameter, salami were sprayed with a suspension of <u>Penicillium</u> <u>nalgiovense</u>. The salami were allowed to mature at 18°C for 2 days, at 17-15°C for 3 weeks and at 14°C for 1 week. During maturing pH and aw were measured.

Microbiological determinations

Samples of minced meat and portions of salami (about 50 g each) were mixed with a sterile solution (0.85 % NaCl and 0.1% peptone) and homogenized in a stomacher. <u>L. monocytogenes</u> numbers were determined on Palcam agar (2 days ^{at} 37°C) and confirmed by motility and emolysis tests. The lactobacilli were enumerated on Rogosa SRL agar (3 days ^{at} 30°C).

RESULTS AND DISCUSSION - The impedometric method showed that the growth of <u>L. monocytogenes</u> 38 was inhibited in a culture supernatant of <u>L. plantarum</u> MCS (Fig.1). At pH 6, a delay of 32 h in detection time was observed; on the contrary, at pH 5.5 no growth occurred in 96 h (flat impedometric curve). The anti-listeria activity of the cultur^e supernatant was lost when this was treated with pronase, proving that a protein or a proteinaceaus compound is probably responsable for the antagonistic effect.

An agar spot test was used to screen, after acriflavine treatment, mutant strains of <u>L. plantarum</u> MCS by their reduced inhibitor activity. Only one strain (MCS1) among the selected ones, showed little residual antilisteria activity by impedometric method (Fig.2). The delay in detection time in pH 6-adjusted supernatant dropped from 32 to about 5 h.



Fig. 1 - Growth of <u>L. monocytogenes</u> 38 at 30°C in APT control (→) and in filtered supernatants of a culture of <u>Lactobacillus</u> MCS (----).
△ adjusted to pH 6.0
□ adjusted to pH 5.5

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Fig. 2 -

Growth of <u>L. monocytogenes</u> 38 at 30°C in media adjusted to pH 6.0.

- \triangle APT broth control
- □ filtered supernatant of a culture of <u>Lactobacillus</u> MCS1
- O filtered supertantant of a culture of <u>Lactobacillus</u> MCS.



The antagonistic effect of MCS strain verified by plate counts of surviving listerias is shown in Fig.3. <u>L. monocytogenes</u> grew faster in MCS1 supernatant at pH 6.0 and more slowly at pH 5.5; slow delayed growth was observed in MCS supernatant at pH 6.0, while at pH 5.5 <u>Listeria</u> was inactivated. At this pH value a bactericidal effect previously observed for a similar strain (CAMPANINI et al, 1991) was confirmed, proving a synergic antagonistic action.

Growth and survival of <u>L. monocytogenes</u> 38 at 30°C in media adjusted to pH 6.0 (----) and 5.5 (----)

APT broth control

filtered supernatants of a culture of Lactobacillus MCS1

▲ filtered supernatants of a culture of <u>Lactobacillus</u> MCS



Fig. 4 -

Fig. 3

Behaviour of <u>L. monocytogenes</u> in minced raw pork at 18°C. No added lactobacilli, pH 5.8 (\blacktriangle) and 5.6 (\bigcirc) In presence of <u>L. plantarum</u> MCS, pH 5.8 (\bigtriangleup) and 5.6 (\bigcirc). The results obtained in minced meat (pH 5.8 and 5.6) are shown in Fig.4. In samples inoculated with MCS strain <u>L. monocytogenes</u> counts declined by 0.8 and 1.3 logarithmic cycles in 15 days at 18° C in meat at initial pH 5.6 and 5.8, respectively. On the contrary, in samples inoculated only with <u>Listeria</u>, the counts declined by a 0.3 logarithmic cycle in meat at pH 5.6, while <u>L. monocytogenes</u> grew in meat at pH 5.8. Recently SCHILLINGER et al (1991) observed a rapid growth of listerias at 15° C in comminuted cured raw pork at high pH (6.3), while no growth occurred at pH 5.7, independently of inoculum level.

During maturing, in salami inoculated only with <u>Listeria</u> after 7 days the lattobacilli naturally present increased from about 103 to more than 5*107, but the pH value remained almost constant at 5.7-5.8 and a growth of listerias (an increase of about ten fold) was observed (Fig.5). However, in the presence of <u>L. plantarum</u> MCS and MCS1 strains, pH decreased to 5.3 after 7 days and the viable number of <u>L. monocytogenes</u> remained more or less constant. After 14 days, pH values were unchanged but <u>Listeria</u> counts tended to decrease in all samples; at 31 days in salami inoculated only with <u>Listeria</u> (pH 5.8) the survival counts came back to the initial level, while in the samples inoculated with lactobacilli (pH 5.5), a reduction of about 0.7 and 0.6 log cycles was observed with MCS and MCS1, respectively. The small differences in the

survival of listerias, in salami inoculated with two different lattobacilli strains, suggest that the effectiveness of bacteriocin may be reduced by a limited diffusion of the protein in meat, adsorption by meat particles or inactivation by proteolytic enzymes produced during maturation. Similar observations in different meat and dairy products have been reported by other authors (PUCCI et al, 1988, SCHILLINGER et al, 1991).



CONCLUSIONS

Inactivation of <u>L. monocytogenes</u> in cured meat and in salami inoculated with <u>L. plantarum</u> MCS strain is much less effective in these products than in culture systems; no significant differences in <u>Listeria</u> counts were observed in matured salami inoculated with MCS or MCSI bacteriocin-negative strain. However, inoculated <u>Lactobacillus</u> strains prevent <u>L. monocytogenes</u> growth, which confirms the importance of using suitable starter culture for salami maturation.

Fig. 5 - Behaviour of <u>L. monocytogenes</u> (-----) and lattobacilli (-----) in salami during maturation.

- no added lattobacilli
- in presence of L. plantarum MCS1
- in presence of L. plantarum MCS

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