ATIONSHIPS BETWEEN LIPID OXIDATION, ANTIOXIDANT ENZYME ACTIVITIES AND OUR STABILITY IN RAW BEEF DURING STORAGE GATELLIER, M. ANTON, FATIMA CHRAITI & M. RENERRE ^{In de} recherches sur la viande, Theix, 63122 St-Genès-Champanelle

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th of this experiment was to appreciate with two bovine muscles different from the viewpoint of colour stability (longissimus ^{the and} psoas major), the relationships between lipid oxidation and haeminic pigment oxidation; total iron and haeminic iron ^{the antions} of the meat were also determined. Additionally, to underscore the production of free radicals in these reactions during meat ^{the antioxidant} enzyme activities were carried out.

^{Mance} spectra of the meat were obtained by spectrocolorimetry and metmyoglobin content was measured on days 1, 3, 7, 10 and 13 ^{Mance} During the same time, lipid oxidation was followed by determination of TBA-RS. Total iron concentration was determined by atomic ^{Mon} spectrometry. Glutathione peroxidase, superoxide dismutase and catalase activities were measured on days 0 and 8 storage. ^{Astorage} of 13 days, it was shown that psoas major muscle was more color labile and displayed a greater extent of lipid oxidation than ^{Minus} lumborum muscle; the two phenomena were highly correlated. After a storage of 8 days, there were some significant differences ^{Minus} activities between muscles in relation with colour stability.

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^{helopment} of MetMb at the meat surface is depending on many biochemical factors but myoglobin autoxidation rate is one of the main ^{help} factor (Renerre & Labas, 1987). Although lipid and pigment oxidation are closely coupled, the relationships between the two ^{hena} are not always clearly established even if it is admitted that these oxidative process are of radical nature (Anton et al, 1991, 1992). ^g MbO2 autoxidation, hydrogen peroxide is produced endogenously in the muscle system and is conducting to oxy-radicals; in situ, ^{hele} degradation of hydrogen peroxide is catalysed by catalase and glutathion peroxidase (Bendich, 1991). Moreover, if for some authors, ^{hele} iron is effective catalyst of lipid oxidation, others stressed that major catalytic agent would be free inorganic iron. For Faustman & ^{hele} (1990), lipid oxidation is a promoter of pigment oxidation. The major objective of this study was to characterize the colour and lipid ^{hele} ^{hele} from two different muscles and to quantify antioxidant enzyme activity in relation with metmyoglobin accumulation rates.

^{trial} and methods

^{huscles} different from the viewpoint of color stability and oxidative capacity : psoas major (PM) and longissimus lumborum (LL) were ^{lh} post-mortem on Charolais cull cows (n=3) and put in liquid nitrogen. Haeminic pigment concentration was measured according to ^{key} (1956). Total iron concentration in muscles were determined by atomic absorption spectrometry (Lamand et al., 19). Colour ^{key} (1956). Total iron concentration in muscles were determined by atomic absorption spectrometry (Lamand et al., 19). Colour ^{key} (1956). Total iron concentration in muscles were determined by atomic absorption spectrometry (Lamand et al., 19). Colour ^{key} (1956). Total iron concentration in muscles were determined by atomic absorption spectrometry (Lamand et al., 19). Colour ^{key} (1956). Total iron concentration at use a second of the cite of the cite

Results and discussion

Between days 1 and 13 storage of beef meat, pigment and lipid oxidation for LL and PM muscles are presented in figures 1 and 2. During period, the MetMb accumulation increased between 13 and 29% for LL muscle and between 18 and 60% for PM muscle. These result very similar to our previous observations obtained with animals of different age, sex and breed (Renerre, 1984). Intermuscular variability effectively one of the most important factor implicated in the rate of meat discoloration in relation with fibre type profiles: Hunt & Ho (1977) have shown that PM muscle was more rich in slow-twitch oxidative fibres than LL muscle. It was established that whatever the iron was not present in greater quantities in PM muscle than in LL muscle (NS) (table 1). Haeminic iron concentrations were similar 10¹ observed previously in our laboratory and by Bousset and Dumont (1989); expressed as percent of the total iron, haeminic iron concentration were near of those of Hazel (1982). Moreover, as shown by Andersen & Andersen (1989) it was demonstrated that determination of in atomic absorption spectrometry yielded significant correlation with pigment content appreciated by Hornsey method (r=0.785). For lipid oxidation, TBA tests showed an increase between days 1 and 13 storage more prononced in PM muscle than in LL muscle (fight). The correlation between lipid and pigment oxidation was presented in figure 3 where it could be observed a strong relationship between two phenomena (first order regression): r=0.647 for LL and 0.826 for PM. The correlation coefficients between these characteristics similar to those observed on beef meat (Faustman & Cassens, 1991) or on ground veal meat (Faustman et al., 1992). Contrary to reobtained with metmyoglobin %, TBA values showed a great variability between animals probably due to the used method. Previously had observed that myoglobin autoxidation was linked to microsomal lipid peroxidation and that these interactions were of radical null Anton et al., 1991,1992). Conversely, lipid peroxides may induce MetMb formation : Mitsumoto et al. (1991) have shown that dietary vit E supplementation retarded metmyoglobin formation in the meat and highly suppressed lipid oxidation compared to the controls. With the meat, vitamin E supplementation has resulted in a lower myoglobin oxidation during meat storage (Santé et al, 1992).

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As radical products of lipid peroxidation and myoglobin oxidation may be present, it was interesting to test the antioxidant enzyme ^{actim} present in the muscles. With the used extraction method, it couldn't be possible to discrimine between Cu-Zn SOD and Mn-SOD^W catalyse the reduction of superoxide free radicals to hydrogen peroxide. For the SOD activity, no difference was found between muscles days 0 and 8 storage for mean values (table 2) as for individual values (paired t-test) (table 3). These values were near of those found^{WW} muscles of various mamalian as cows (Sohal et al., 1990) or in rat liver (De & Darad, 1991). Moreover, by using the paired t-test, found that at day 0 storage, catalase activity was less important in PM muscle than in LL muscle (P<1%); after 8 days storage, we obtain identical results but differences between muscles are less prononced (P<5%) (table 3). As catalase activity was less prononced in muscle as PM, H2O2 may be formed in greater quantities in this muscle; with microsomes preparations, it was observed that non-enlipid peroxidation was much more prononced in PM muscle than in LL muscle (Anton et al. 1992).

For glutathion-peroxidase activity, there were no difference between muscles at day 0 storage but a highly significant difference at (P<1%). As for catalase activity, glutathion peroxidase activity was less important in PM muscle, more instable from the viewpoint of the stability, than in LL muscle. Glutathion peroxidase activity was not far from those reported previously on heart (Sohal et al, 1990). Faith & Cassens (1991) found that Gluteus medius muscle, more unstable than Longissimus muscle (Renerre, 1984), had greater TBA value. MetMb and hypoxanthine concentration indicating a more prononced formation of superoxide anion.

In this first experiment, if there is a strong relationship between lipid and myoglobin oxidation, conversely, it is not easy to conclude the antioxidant enzyme activities in relation with color stability.

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Figure 1 : Increase of MetMb % between days 1 and 13 storage

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Figure 3 : Relationship between myoglobin and lipid oxidation

Muscle	Total iron (1) (μg/g)	Haeminic iron (2) (µg/g)	(2)/(1) %
LL	22.9 ± 4.3	15.7 ± 1.2	70 ± 19
PM	21.3 ± 1.0	17.1±0.6	80±5
	NS	NS	NS

Table 1: Total from and naeminic from content of LL and	1 PM	muscles
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Time	e SOD		Catalase		GPx	
	LL	PM	LL	PM	LL	PM
Day O	7.6±3.3	7.3 ± 3.6	0.9 ± 0.3	0.8±0.3	39 ± 14	28±13
Day 8	7.7 ± 2.9	7.6±2.9	0.9±0.2	0.6 ± 0.2	33±4	20±5

Table 2 : Enzymic activities of SOD, Catalase and GPx in LL and PM muscles (mean values)

Time	SOD	Catalase	GPx
Day O	NS	<1%	NS
Day 8	NS	< 5 %	<1%

Table 3 : differences between LL and PM for enzymic activities of SOD, Catalase and GPx (paired t-test)

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