

# EFFECT OF CRYOPROTECTANTS DURING FROZEN STORAGE ON GELLING PROPERTIES OF MYOFIBRILLAR PROTEIN ISOLATES .

F.J.G. SCHREURS, T.G. UIJTENBOOGAART, T.L. TRZISZKA<sup>1</sup> and H.G.M. REIMERT

Welderholt Centre for Poultry Research, Agricultural Research Department, Ministry of agriculture, NL-7361 AZ Beekbergen, The Netherlands.

<sup>1</sup>T.L. Trziszka is with the Department of Food Technology of Animal Origin, University Of Wroclaw, Poland.

## INTRODUCTION

Scientific work on isolation and use of mechanically deboned poultry meat (MDPM) (Dawson *et al.*, 1989, Hernandez *et al.*, 1986, Schreurs *et al.*, 1991, Trziszka *et al.*, 1991a/b, 1992, Uijttenboogaart *et al.*, 1992) have shown great potential for the industrial production of protein preparations like 'Chicken-surimi' or 'Chicken Myofibrillar Protein Isolate'. A large problem with these types of protein isolates is their storage because myofibrillar proteins are rather susceptible for microbiological deterioration even under cooled circumstances. Meat and meat products are commonly preserved by freezing, since under these conditions there is relatively little deterioration in product quality even after prolonged storage (Barbut and Mittal, 1990; Krivchenia and Fennema, 1988; Sikorski *et al.*, 1976). Muscle tissue structure is affected to a greater extent by freezing and frozen storage than are its chemical properties. This is mainly due to crystallization of water in muscle tissue, especially the size and location of ice crystals produced (Sebranek, 1982). Frozen storage of finely comminuted meat products requires particular attention due to the possible occurrence of fat oxidation and freezing-induced protein denaturation which markedly affects the functional properties of frozen products.

Wiarowicz *et al.* (1990, 1991) found that enhanced oxidative changes occurring in mechanically deboned poultry meat (MDPM) during frozen storage caused a marked deterioration in the functional properties of proteins. They suggested that frozen storage of MDPM should be extended beyond 2 to 3 months.

Isolation of myofibrillar proteins (MPI) from MDPM, as an alternative to the conventional utilization of by-products in further processed poultry products, gives manufacturers the opportunity to produce MPI on an industrial scale (Dawson *et al.*; 1989, Schreurs *et al.*, 1991; Trziszka *et al.*, 1992). However MPI can only be commercially applied in products provided it will not lose its functional properties during frozen storage.

The possibility to overcome these problems is frozen storage in the presence of cryoprotectants. In the fish industry, surimi is usually stored in the presence of sucrose, sorbitol and polyphosphates (Lee, 1984).

In our earlier study (Trziszka *et al.*, 1991) it was found that the addition of sucrose and sorbitol as well as monosodium glutamate (0.5%) and tetrasodium pyrophosphate (0.3%) to MPI significantly increased gelling properties during heating. The use of mono- and disaccharides increases sweetness, simultaneously decreasing the usability of the endproduct (Sych *et al.*, 1990, Sych *et al.*, 1991). For this reason studies on other protectants capable of inhibiting freeze induced protein denaturation in MPI are required. Of particular interest are the studies on the use of polydextrose during frozen storage of fish surimi (Lanier and Akahane, 1986, Park *et al.*, 1988), and the use of other high molecular weight carbohydrates (HMWC) (MacDonald & Lanier, 1991) like starch.

Recent studies on protein protection during frozen storage have focussed on 4 groups of compounds, i.e. carbohydrates, polyalcohols, amino acid hydrolyzates and hydrocolloids (Sych *et al.*, 1990).

It is of major importance that the protein isolates can be stored for extended periods of time without significant changes and that functional properties will be retained as much as possible.

The first scope of our investigations was to determine the possibility of frozen storage of MPI obtained by procedures used in earlier experiments (Trziszka *et al.*, 1991). The studies included the effects of various cryoprotectants and drastic changes of the conditions i.e. exposure to several cycles of freezing and thawing over a storage period of several weeks.

The second scope of this study was to obtain insight in the changes taking place in myofibrillar protein isolates with and without cryoprotectants during extended storage time at -21 °C.

## MATERIALS AND METHODS

The experimental material consisted of MPI obtained by one-stage extraction of mechanically deboned chicken meat (MDCM) in 0.075 M NaHCO<sub>3</sub> using a separator (Westfalia Separator AG, type SB-7-06-076, Oelde, Germany) according to a procedure described by Trziszka *et al.* (1991).

The MPI was divided into four groups:

- |          |   |     |
|----------|---|-----|
| Group 1. | Control, MPI without any cryoprotectants:                           | C   |
| Group 2. | MPI supplemented with 4% (v/w) of Karion F and 4% (w/w) of sucrose: | KAS |
| Group 3. | MPI supplemented with 8% (w/w) polydextrose:                        | PDX |
| Group 4. | MPI supplemented with 4% (v/w) Karion F and 4% (w/w) of starch:     | KST |

Karion F\* is the trade name of a food grade sorbitol syrup (70 % w/w) from Merck, Darmstadt, Germany; sucrose (Analar grade) was obtained from BDH Chemicals, Poole, UK; polydextrose\* was from Pfizer; the potato starch used was obtained from a local grocery store. Mixing of the cryoprotectants in the MPI occurred by using a Hobart N50 mixer during 5 minutes at speed 2. After mixing the material was portioned and vacuum packed into individual plastic bags, each containing ca 300 g.

The first experiment (short term storage and repeated freezing and thawing) was carried out according to scheme 1. A sample was taken from fresh MPI as well as from the 1, 2 or 3 times frozen and thawed MPI, stored respectively for 2, 3 and 4 weeks. Another sample was taken at -21 °C for 4 weeks prior to thawing.

In the second experiment (long term frozen storage) portions of each group of cryoprotectant were submitted to frozen storage at approximately -21 °C for 0, 1, 3, 6 and 9 months respectively.

### Gel production and characteristics.

Twenty g of MDCM, or the appropriate MPI, containing 2.0% of NaCl were heated at 90°C for 15 minutes in a polypropylene tube ( $\phi$  21 mm length 73 mm), cooled with tap water and refrigerated at 4°C until the next day (Trziszka *et al.*, 1992).

Cooking losses of the gels were calculated from the difference in weight before and after heat treatment.

Gel strength (hardness), expressed in newtons, and springiness, expressed in mm, were determined using an Overload Dynamics food texture measuring device as described by Lyon *et al.*, (1980).

Gel elasticity (folding test) was determined by twice folding 3 mm thick gel slices (15 mm diameter) and evaluating cracking of the gel using a 5-point scale established by Nippon Suisan Kaisha Ltd. (Lee, 1984).

All data were statistically analyzed by analysis of variance using the GENSTAT V program (Copyright, 1984, Lawes Agricultural Experiment Station, Rothamsted Agricultural Station, UK).

## RESULTS AND DISCUSSION.

### Experiment 1

The data obtained in this study are shown in the Table 1. It shows the major parameters determining the functional properties of MPI on the basis of gels after heat treatment. Weight losses significantly differed depending on the cryoprotectant used. Each MPI with cryoprotectant added showed an increased loss due to prolonged storage and multiple freeze thaw treatments. The values obtained with KAS (except 0 and 2 weeks) and PDX proved to be significant.

The losses were the highest in the groups in which the MPI was frozen and thawed repeatedly.

Regarding the individual cryoprotectants, the weight losses using KAS and PDX were high compared with those noted with KST. KAS and PDX, cryoprotectants widely used in surimi production, did not prove to be suitable in cases where MPI was exposed to repeated freezing and thawing. KAS can be used as a cryoprotectant in MPI provided that freezing and thawing is not repeated. PDX proved to be unsuitable as a cryoprotectant for MPI due to high cooking losses.

The low weight loss observed with KST (sorbitol and starch) present is very likely due to water binding of starch. Since a wide variety of meat products is supplemented with starch as a binding agent, its addition to MPI can be beneficial for two reasons:

On one hand it functions as a gel-binder (Oakenfull, 1987) and on the other hand as a cryoprotective agent (MacDonald & Lanier, 1991). Based on these findings it seems favorable that the addition of starch is advantageous.

Gel strength, expressed in newtons is a major factor determining the texture of gels. The data in Table 1 show a few significant differences in gel resistance (hardness) for the groups with added cryoprotectant as compared to the control. Addition of KAS and PDX showed a significant increase on gel hardness after storage for 2 and 3 weeks. After that period the hardness values decreased to non-significant levels as compared to the controls.

Each freezing and thawing cycle of the MPI from the control groups decreased the gel strength significantly, whereas in the cryoprotectant supplemented MPI the original gel strength values were maintained at a level twice as high as the respective control samples regardless of the drastic freezing and thawing operations.

Springiness, defined as the distance a sample recovers between first and second bite (Lyon *et al.*, 1980), is also a parameter which cannot be neglected in the evaluation of the texture of products. As can be seen from Table 1, freezing and thawing of MPI without any cryoprotectant significantly decreased the value of this parameter, thus indicating that elasticity of the gel had been reduced.

The cryoprotectants used in this experiment significantly increased springiness of gels produced from frozen stored MPI. In this respect the best cryoprotectant proved to be KST while the least effective was PDX.

The folding test, another indicator of gel springiness and commonly used in the textural analysis of surimi (Hastings, 1989, Lee, 1984) confirmed the data obtained above which means a significant loss of this property by the freezing and thawing treatment in the control groups.

The use of cryoprotectants only resulted in a significant lower springiness in case of addition of KAS and stored for 4 weeks during repeated freezing and thawing. The greatest springiness was noted for gels from MPI with added KST.

Summing up, it can be noted that MPI which is to be stored frozen should be supplemented with cryoprotectants to protect the functional properties of the proteins probably by inhibition of freeze-induced protein denaturation.

The best cryoprotectant of the three used in our experiments proved to be 4 % Karion F (sorbitol) in combination with 4 % starch although a combination of 4 % Karion F and 4 % sucrose, widely used in surimi technology, can also be used for MPI. Sorbitol and sucrose, however, will impact a sweet taste to the MPI which can be a problem (Sych *et al.*, 1990).

### Experiment 2

The data obtained in this study are shown in tables 2 to 6. The yield values after heating, as shown in table 2 clearly show a large (statistically significant) difference between KST and other treatments. In the product with polydextrose added large cooking losses are found. These results are comparable to those found in experiment 1 and opposed to the results found with fish surimi (Park *et al.*, 1988) where polydextrose was found to be a good cryoprotectant. Karion and sucrose seem to be better cryoprotectants.

Table 3 shows the data for gel strength measurements. Obviously the utilization of the different cryoprotectants has a large effect on the gel strength compared to the starting material. Controversial effects were observed in groups with and without cryoprotectants. During storage of the materials, gel strength increased in the cryoprotected groups as opposed to the non protected groups.

Table 4 shows the springiness values obtained. These data clearly show the increasing effect of cryoprotectants on springiness, especially the increase due to the addition of Karion and starch.

Table 5 shows the shear force values. The supplementation of MPI with Karion/starch and polydextrose results in a significant increase in gel hardness.

Table 6 shows the folding test data as a measure of elastic properties of the gels. As mentioned before this test shows similar results as do the instrumental elasticity measurements. During storage a significant deterioration of elastic properties was observed in the control group but in the cryoprotected groups the folding test values remained relatively stable. The effects observed in gels with starch as a cryoprotectant obviously can partly be attributed to the gel forming properties of the starch itself.

Based on these findings, it is clear that long term frozen storage has a detrimental effect on functional properties of MPI. Utilization of cryoprotectants clearly has a positive effect on quality of these isolates. The mechanical properties of the gels, made from materials stored

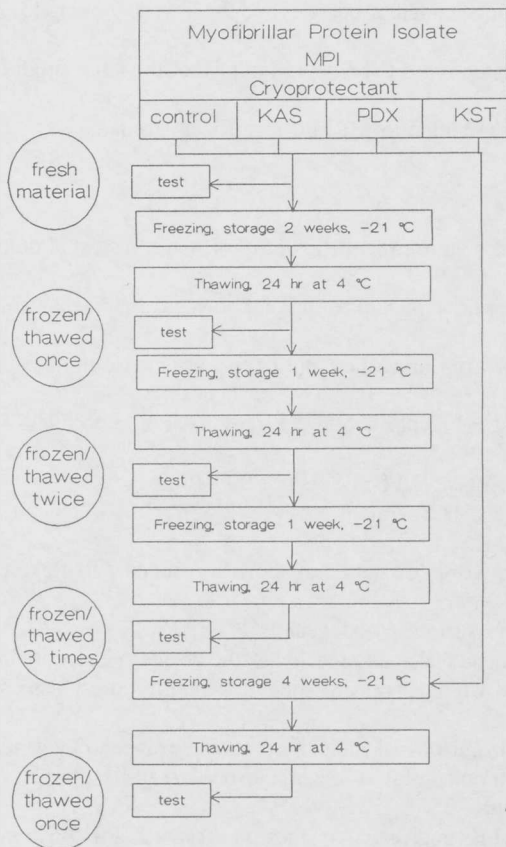
er cryoprotection, were found to be significantly better than the control gels. The best mechanical properties and the lowest cooking losses were found when Karion/starch was used as a cryoprotectant. The use of polydextrose or sorbitol/sucrose for cryoprotection in MPI is possible but again it has to be kept in mind that the latter products result in excessive sweetness (Sych *et al.*, 1990, 1991). Further research is required to find the optimal condition for use of cryoprotectants to be added to MPI during frozen storage.

#### REFERENCES

- out, S., and Mittal, G.S. 1990. Influence of the freezing rate on the rheological and gelation properties of dark poultry meat. *Poultry Sci.* 69: 827-832.
- son, L.E., Sheldon, B.W. and Ball Jr., H.R. 1989. Pilot-plant washing procedure to remove fat and color components from mechanically deboned chicken meat. *Poultry Sci.* (68) 749
- ings, R.J. 1989. Comparison of the properties of gels divided from cod surimi and from unwashed and once-washed cod mince. *Int. Food Sci. Tech.* 24:93-102
- andez, A., Baker, R.C. and Hotchkiss, J.H. 1986. Extraction of pigments from mechanically deboned turkey meat. *J. Food Sci.* 51: 655.
- chenia, M. and Fennema, O. 1988. Effect of cryoprotectants on frozen whitefish fillets. *J. Food Sci.* 53: 999-1003.
- er, T.C., and Akahane, T. 1986. Method of retarding denaturation of meat products. U.S. Patent No. 4,572,838.
- er, C.M. 1984. Surimi process technology. *Food Technol.* 38: 69.
- on, C.E., Lyon, B.G., Davis, C.E., and Townsend, W.E. 1980. Texture profile analysis of patties made from mixed and flake-cut mechanically deboned poultry meat. *Poultry Sci.* 59:69-76.
- Donald, G.A. and Lanier, T. 1991. Carbohydrates as cryoprotectants for meats and surimi. *Food Techn.* 45: 3, 150.
- awiarowicz, A., Kijowski, J. and Pikul, J. 1990. Właściwości funkcjonalne mięsa drobiu odzyskanego mechanicznie oceniane bezpośrednio po odzyskaniu; W czasie zamrażalniczego przechowywania. [Functional properties of mechanically deboned poultry meat assessed immediately after deboning and after frozen storage] *Gospodarka Miesna*, 11:25-27.
- awiarowicz, A., Kijowski, J. and Pikul, J. 1991. Sposoby przedłużania trwałości mięsa drobiu odzyskanego mechanicznie przechowywanego w stanie zamrożonym. [Methods of extension of shelf-life of mechanically deboned poultry meat stored frozen] *Gospodarka Miesna* 4:21-23.
- enfull, D. 1987. Gelling agents. *CRC Critical Reviews in Food Science and Nutrition* 25: 1, 1.
- er, J.W., Lanier, T.C. and Green, D.P. 1988. Cryoprotective effects of sugar, polyols, and/or phosphates on Alaska Polok Surimi. *J. Food Sci.* 53: 1,1.
- reurs, F.J.G., Trziszka, T. and Uijttenboogaart, T.G. 1991. Isolating myofibrillar proteins from deboned poultry meat. *World Poultry Sci.* (3):21.
- reurs, F.J.G., Trziszka, T., Uijttenboogaart, T.G. and Reimert H.G.M. 1992. Long term frozen storage of myofibrillar protein isolates obtained from poultry meat. *J. Food Sci.* Paper submitted.
- ranek, J.G., 1982. Use of cryogenics for muscle foods. *Food Technol.* 36(4):121-127.
- owski, Z., Olley, J. and Kostuch, S. 1976. Protein changes in frozen fish. *Crit. Rev. Food Sci. Nutr.* 8:97-129
- ch, J., Lacroix, C., Adambounou, L.T. and Castaigne, F. 1990. Cryoprotective effect of some material on cod-surimi proteins during frozen-storage. *J. Food Sci.* 55: 5, 1222.
- ch, J., Lacroix, C., Adambounou, L.T. and Castaigne, F. 1991. The effect of low- or non-sweet additives on the stability of protein functional properties of frozen cod surimi. *Int. J. Food Sci. Technol.* 26: 185.
- szka, T., Popiel, A.K. and Kulpa, B. 1991a. Washing procedure to remove fat and colour components from mechanically deboned turkey meat. Proceedings of the 10th European Symposium on the Quality of Poultry Meat, Doorwerth, May 12-17, 1991, 167.
- szka, T., Uijttenboogaart, T.G. and Schreurs, F.J.G. 1991b. Use of  $\text{Na}_2\text{CO}_3/\text{NaHCO}_3$  buffer for the extraction of myofibrillar proteins from MDPM and the influence of freezing on the functional properties of the isolates. Proceedings of the 10th European Symposium on the Quality of Poultry Meat, Doorwerth, May 12-17, 1991.
- szka, T.L., Uijttenboogaart, T.G. and Schreurs, F.J.G. 1992a. Characteristics of myofibrillar protein isolate obtained by different methods from mechanically deboned chicken meat. *J. Food Sci.* In press.
- boogaart, T.G., Trziszka, T.L. and Schreurs, F.J.G. 1992b. The use of cryoprotectants in myofibrillar protein isolates obtained from poultry meat, destined for frozen storage. *J. Food Sci.* In press.

**Table 4** Results of experiment 2: Springiness of gels, as the distance between first and second bite (mm).

Cryoprotectant	Control	KAS	PDX	KST	Mean
Storage time (months)					
0	11.7	12.0	12.1	14.2	12.5 <sup>a</sup>
1	11.2	11.8	12.3	13.2	12.1 <sup>a</sup>
3	11.2	12.5	12.3	13.1	12.3 <sup>a</sup>
6	10.6	11.6	11.7	12.8	11.7 <sup>b</sup>
9	10.8	12.2	11.7	13.2	12.0 <sup>ab</sup>
Mean	11.2 <sup>a</sup>	12.0 <sup>b</sup>	12.0 <sup>b</sup>	13.3 <sup>c</sup>	



Scheme 1: protocol of first experiment

Table 1 Results of experiment 1

Parameter and statistics	Storage period P (weeks)	# of freeze /thaw treatments	Cryoprotectants C			
			control	KAS*	PDX*	KST*
Cooking loss (g) LSD: P = 5.09 C = 13.26 P/C = 11.25	0		5.75	5.37	8.25	0.92
	2	+	10.42	7.50	14.17	2.00
	3	++	12.75	18.17	26.21	3.75
	4	+++	17.50	28.67	34.25	5.83
	4	+	12.62	8.13	13.83	2.33
Hardness [N] LSD: P = 17.78 C = 13.34 P/C = 24.14	0		36.39	30.55	28.66	66.25
	2	+	27.11	48.92	53.08	55.83
	3	++	24.33	52.67	45.89	49.46
	4	+++	19.87	41.53	34.46	53.87
	4	+	18.73	38.81	36.96	56.33
Springiness [mm] LSD: P = 1.97 C = 1.23 P/C = 2.57	0		14.28	12.50	13.67	14.52
	2	+	10.50	13.25	12.77	13.65
	3	++	10.98	13.38	11.80	13.50
	4	+++	10.21	11.15	10.00	12.38
	4	+	10.63	12.63	12.64	13.40
Folding test LSD: P = 0.51 C = 0.80 P/C = 0.84	0		4.04	4.42	3.58	4.67
	2	+	3.17	4.58	4.54	4.58
	3	++	2.67	3.96	3.21	4.21
	4	+++	2.96	3.67	3.42	4.17
	4	+	2.95	4.04	4.04	4.12

The differences are statistically significant if the difference between the means are higher than LSD for each parameter.

Table 2 Results of experiment 2: Cooking loss after heating of MPI (%).

Cryoprotectant	Control	KAS	PDX	KST	Mean
Storage time (months)					
0	4.90	3.83	5.60	0.75	3.77 <sup>ab</sup>
1	6.77	5.19	7.25	1.50	5.18 <sup>c</sup>
3	6.10	4.69	6.08	0.88	4.44 <sup>bc</sup>
6	5.27	2.67	5.06	1.25	3.55 <sup>a</sup>
9	6.85	3.92	7.69	1.04	4.88 <sup>c</sup>
Mean	5.98 <sup>a</sup>	4.06 <sup>ab</sup>	6.34 <sup>a</sup>	1.09 <sup>b</sup>	

Table 5 Results of experiment 2: Shear-Force of gels (N).

Cryoprotectant	Control	KAS	PDX	KST	Mean
Storage time (months)					
0	2.11	1.89	2.18	2.91	2.27 <sup>a</sup>
1	2.60	2.61	3.57	4.15	3.29 <sup>b</sup>
3	2.26	2.57	2.95	3.46	2.81 <sup>c</sup>
6	2.46	2.32	2.61	3.24	2.66 <sup>c</sup>
9	2.33	2.33	2.76	3.14	2.64 <sup>c</sup>
Mean	2.35 <sup>a</sup>	2.35 <sup>a</sup>	2.81 <sup>b</sup>	3.38 <sup>c</sup>	

Table 3 Results of experiment 2: Gel strength (max force in N).

Cryoprotectant	Control	KAS	PDX	KST	Mean
Storage time (months)					
0	16.73	16.89	19.23	31.18	21.01 <sup>a</sup>
1	15.25	19.37	24.03	33.31	22.99 <sup>bc</sup>
3	15.81	23.27	24.84	33.41	24.33 <sup>bc</sup>
6	15.21	19.42	22.78	29.24	21.66 <sup>ab</sup>
9	14.94	22.32	25.95	35.01	24.55 <sup>c</sup>
Mean	15.59 <sup>a</sup>	20.25 <sup>ab</sup>	23.37 <sup>b</sup>	32.43 <sup>c</sup>	

Table 6 Results of experiment 2: Folding test of gels.

Cryoprotectant	Control	KAS	PDX	KST	Mean
Storage time (months)					
0	3.79	4.06	3.73	4.31	3.97 <sup>a</sup>
1	4.06	4.40	4.56	4.85	4.47 <sup>b</sup>
3	3.85	4.33	4.17	4.58	4.23 <sup>ab</sup>
6	3.35	4.27	4.33	4.67	4.16 <sup>c</sup>
9	3.17	4.29	4.35	4.58	4.10 <sup>c</sup>
Mean	3.65 <sup>a</sup>	4.27 <sup>ab</sup>	4.23 <sup>ab</sup>	4.06 <sup>b</sup>	