

# THE EFFECT OF FREEZING AND HEATING ON CHANGES OF SOME SARCOPLASMATIC PROTEINS OF BOVINE *LONGISSIMUS DORSI*

L. Gašperlin, B. Žlender, V. Abram, S. Ketiš

University of Ljubljana, Biotechnical Faculty, Department of Food Science and Technology, Jamnikarjeva 101, 1111 Ljubljana, SLOVENIA

## Introduction

Freezing is one of the best and the most used methods of extending the shelf-life of foods. Some consumers still now consider and believe that frozen meat is of noticeably worse quality than the chilled one. Žlender (1995) estimated that the rate and duration of freezing had noticeable effect on physical, structural, chemical, hygienic and sensory quality parameters of meat.

Cooling meat to 0°C had little or no effect on the muscle structure, but the process of freezing animal tissues led to considerable denaturation of both soluble proteins of the sarcoplasm and the contractile protein complex of actin-myosin + tropomyosin-troponin-actinin due to a disruption of cellular organelles like mitochondria, lysosomes, etc., which release the enzymes bound to these structures into the cell sap (Bendall, 1979; Hamm, 1979; Hamm et al., 1986). There are reports that freezing caused a remarkable release of cytochrome c oxidase from mitochondria (Barbagli in Serlupi-Crescenzi, 1981). The same happens to meat pigments.

The objective of this study was to investigate the changes of sarcoplasmatic proteins (soluble sarcoplasmatic proteins in general) and separated content of native and denatured myoglobin and specific activity of cytochrome c oxidase of bovine *m.longissimus dorsi* after freezing and heating.

## Material and methods

24 to 48 hours *post mortem* three *longissimus dorsi* muscles (LD) of normal quality (pH = 5.6) were cut off from the left and right halves of the young beef. On the right (fresh and unfrozen) and on the left halves (frozen to -30°C and thawed in 24 hours at +5°C) LD, divided into 10 fresh and three heated samples ( $T_i$  = 50°C, 58°C, 65°C), myoglobin quantification (Trout's Nit<sub>409</sub> method (1991) modified by Špenič (1994)), determination of the percentage of denatured myoglobin, specific activity of cytochrome c oxidase (Appelmans et al., 1954) and electrophoretic characterisation of soluble sarcoplasmatic proteins on polyacrylamide gel using a modified method of Laemmli (1970) were carried out. Proteins for electrophoresis were extracted according to the method of Toldra et al. (1992).

## Results and Discussion

Electrophoretic separation of myofibrillar proteins in polyacrylamide gels in the presence of sodium dodecyl sulphate (SDS) and mercaptoethanol is widely used for qualitative analysis of proteins (Claeys, 1995). With this method we wanted to evaluate the effect of freezing and thawing on the soluble sarcoplasmatic proteins of fresh and heated beef meat. The 15% gel concentration served to separate sarcoplasmatic proteins within a molecular weight range of 10 - 50 kDa (Figure 1). After comparison of proteins migration distance from the top of the separating gel to the center of the protein band on 15% SDS-PAGE gel we could resume that electrophoretic profiles of frozen and unfrozen samples of sarcoplasmatic proteins did not distinguish. Different intensity of separated proteins was due to the different protein concentrations in samples. We calculated the relative mobility ( $R_f$ ) of unknown proteins and estimated its molecular weight from a calibration curve constructed from the molecular weight of proteins in the Sigma standard (SDS-7 DALTON MARK VII-L) and their  $R_f$  values. We could predict that the most distant protein band was myoglobin with  $M_r$  of 17800.

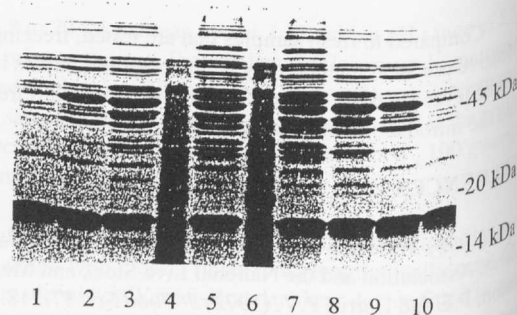


Figure 1 Electrophoretic profiles of sarcoplasmatic proteins of bovine LD separated on 15% SDS-PAGE gel

Legend: 1 = UF(65), 2 = UF(58), 3 = UF(50), 5 = UF(fresh), 7 = FT(fresh), 8 = FT(50), 9 = FT(58), 10 = FT(65), 4, 6 = Sigma standard 7L; UF - unfrozen; FT - frozen/thawed; ( $T_i$ ) - internal temperature (°C)

Table 1 Effect of freezing and heating on the content of native myoglobin in bovine LD samples (mg/g of meat)

thermal treatment	unfrozen $\bar{x}$	frozen $\bar{x}$	t-value
fresh <sup>1</sup>	10.10 <sup>4,3</sup>	6.68 <sup>4,3,2</sup>	2.96*
50°C <sup>2</sup>	8.63 <sup>4</sup>	5.81 <sup>4,3</sup>	3.38*
58°C <sup>3</sup>	6.91	4.76	3.94**
65°C <sup>4</sup>	4.65	4.43	0.23
F-value	7.34**	13.54***	

Significance level of effect is described as \*\*\*, \*\*, \* for  $p < 0.001$ ,  $p < 0.01$ ,  $p < 0.05$

We noticed quite large differences, except for the group of samples heated to  $T_i$  = 65°C, in the content of myoglobin between frozen and unfrozen samples (Table 1 and Figure 2). In spite of careful freezing and thawing sarcolemma and some cell structures were obviously damaged, and this could contribute to the loss of myoglobin in the drip from thawed and rather small frozen samples (9 x 9 x 5 cm) with relatively large surface.

Turner (1974) has found that myoglobin denatures only during prolonged heating at 60°C; after heating meat samples (one hour) with pH 5.5 at 60°C the content of denatured myoglobin was about 30%, at 70°C about 80% and at 75°C close to 100%. Our results (Figure 2) showed that the rate of denaturation was faster in unfrozen samples (relation between native myoglobin (Mb) and  $T_i$ :  $Mb = -0.11T_i + 12.7$ ) than in frozen one ( $Mb = -0.05T_i + 7.8$ ).

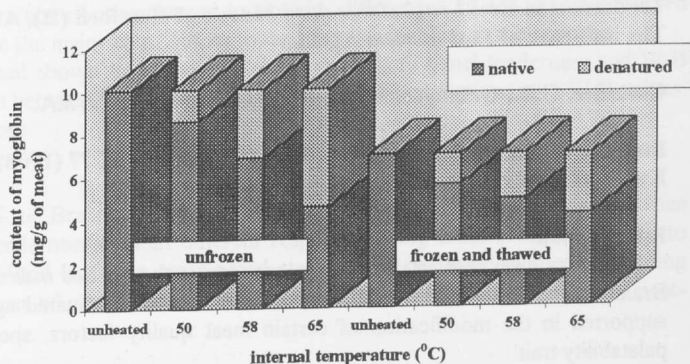


Figure 2 Content of native and denatured myoglobin determined by Nit<sub>409</sub> method in fresh and heated samples

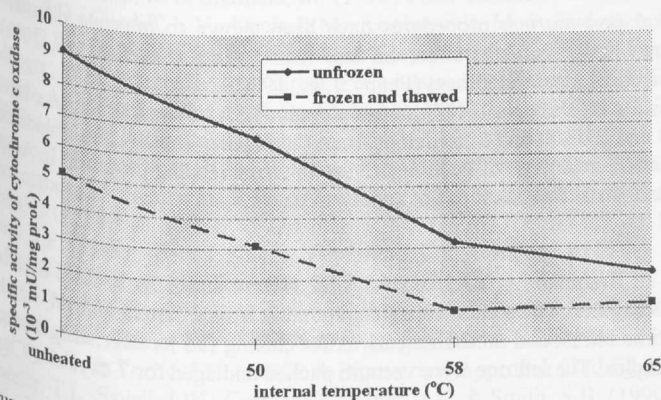


Figure 3 Specific activity of cytochrome c oxidase in fresh and heated samples before and after freezing

Table 2 Effect of freezing and heating on specific activity of cytochrome c oxidase ( $10^{-3}$  mU/mg of protein) in fresh and heated LD

thermal treatment	unfrozen	frozen	t-value
fresh <sup>1</sup>	9.1 <sup>3,4</sup>	5.1 <sup>3,4,2</sup>	1.29
50°C <sup>2</sup>	6.6	3.1 <sup>3,4</sup>	2.18
58°C <sup>3</sup>	3.2	1.0	1.71
65°C <sup>4</sup>	2.2	1.2	1.08
F-value	2.80	20.94***	

Freezing decreased the enzyme activity but differences were not significant (Table 2). Generally, the specific activity of cytochrome c oxidase decreased with increased  $T_i$ . In fresh samples enzyme activity was significantly higher ( $p \leq 0.05$ ) than in heated one.

- Conclusions**
1. Electrophoretic separation of sarcoplasmatic proteins on polyacrylamide gels in the presence of sodium dodecyl sulphate (SDS) and 2-mercaptoethanol is a suitable method for evaluating an effect of freezing and heating on the soluble sarcoplasmatic proteins of beef meat.
  2. Correct freezing and thawing did not have any influence on the electrophoretic profile of sarcoplasmatic proteins and on the specific activity of cytochrome c oxidase.
  3. As expected specific activity of cytochrome c oxidase in fresh meat was significantly higher ( $p \leq 0.05$ ) than in the heated one.
  4. The myoglobin level in fresh and heated to  $T_i = 58^\circ\text{C}$  frozen meat was significantly lower ( $p \leq 0.01$ ) than in unfrozen samples.

**Reference**

Appelmans, F./ Wattiaux, R./ Duve, C. 1955. Tissue Fractionation Studies. Biochemical Journal, 59, p. 438-445.

Barbagli, C./ Serlupi Crescenzi, G. 1981. Influence of freezing and thawing on the release of cytochrome oxidase from chicken's liver and from beef and trout muscle. Journal of Food Science, 46, 5, p. 491-493.

Bendall, J.R. 1971. Structural and biochemical changes during the cooling and freezing of meat. In: IFST-Proceedings 4(3, part I) p. 124-129.

Claeys, E./ Uytterhaegen, L./ Buts, B./ Demeyer, D. 1995. Quantification of beef myofibrillar proteins by SDS-PAGE. Meat Science 39, 2, p. 177-193.

Hamm, R./ Gottesmann, P. 1986. Biochemischer Gefrierfleisch-Nachweis beim wäβrigem, glassem Schweinefleisch. Fleischwirtschaft, 66, 4, p. 552-554.

Hamm, R. 1979. Delocalization of mitochondrial enzymes during freezing and thawing of skeletal muscle. Advances in Chemistry Series, 180, p. 191-204.

Laemmli, U. K. 1970. Nature, p. 227-680.

Špendja, R. 1994. Determination of native and denatured myoglobin in thermal treated beef. Graduation thesis, University of Ljubljana, Biotechnical Faculty, Department of Food Technology, 38 p.

Toldra, F. 1992. The enzymology of dry - curing of meat products. In: New technologies for meat and meat products. Ed. by Smulders, F.J.M. et al. Utrecht, ECCMST, Audet Tijdschriften, p. 209 - 231.

Trout, G. R. 1991. A rapid method for measuring pigment concentration in porcine and other low pigmented muscles. In: 37<sup>th</sup> international congress of meat science and technology, September 1-6, 1991, Kulmbach. Proceedings, Vol. 2. Kulmbach, Germany, p. 75-80.

Turner, L. 1974. Denaturation of proteins. In: Encyclopedia of Food Technology. Johnson, A. H. ed. / Peterson, M. S. ed. Westport, AVI Publish. Comp., p. 292.

Zander, B. 1995. Zmrzovanje mesa. V: Podaljšanje obstojnosti živil, 17. Bitenčevi živilski dnevi '95. 8-10 junij 1995 Ljubljana. Ljubljana, Biotehniška fakulteta, Oddelek za živilstvo, s. 25 - 35.