INVESTIGATIONS INTO WHY BEEF OF INTERMEDIATE ULTIMATE pH IS OFTEN LESS TENDER

R W Purchas & X Yan

Department of Animal Science, Massey University, Palmerston North, NEW ZEALAND Keywords: Beef tenderness, ultimate pH, myofibrillar breakdown, meat ageing

INTRODUCTION

The tenderness of beef longissimus muscle as measured by shear force values has been shown in several studies to decrease as ultimate (pH_{ult}) increases from about 5.5 to 6.0, and then to improve with further increases in pH_{ult} to about 7.0 (Bouton et al. 1957; Purchas with Jeremiah et al. 1991). Proposed explanations for the higher shear force values at intermediate relative to low pH_{ult} include a greater myofilm shortening (Purchas 1990; Olsson et al. 1995), and a lower level of proteolytic activity (Yu & Lee 1986). The former suggestion is support by evidence of shorter sarcomeres and by the absence of any effect when muscles are stretched; and the latter by microscopic evidence of myofibrillar disruption and by slower rates of ageing in some studies (Watanabe et al. 1996), but not in others (Wulf et al. 1996). The am the current study was to assess the relationship between myofibrillar breakdown and shear force in samples of beef longissimus muscle low and intermediate levels of pH_{ult} .

MATERIALS AND METHODS

Samples of longissimus muscle in the 8-12 rib region were taken within 90min *post mortem* from the 156 cattle described by Purchase (1997). These included bulls and steers of several breeds and crosses. From the 156 samples, 24 were selected which included ¹² bulls and steers and by year. The aim in selecting samples on the basis of both pH_{ult} as well as WB peak force was to have the means for two groups close to the points for pH_{ult} values of 5.5 and 6.0 on the overall regression curve of WB peak force versus pH_{ult} .

Following ageing at 0-2°C for either 1 or 20 days, muscle samples were frozen at -20°C until processed. Measurements of pHu (nonhomogenate), WB peak force (after cooking at 70°C for 90min), and sarcomere length (by laser diffraction) were made as described by Pure & Aungsupakorn (1993). Myofibrillar fragmentation indexes (MFI) were measured using a slight modification of the method of Johnson et (1990) that involved passing an homogenate through a 231 μ m screen. Analysis of myofibrillar proteins by SDS-PAGE was conduct according to the method of Claeys et al. (1995), with BSA included as an internal standard at a concentration of 1 μ g BSA/20 μ g myofibrillar protein. Myofibrillar extracts (4 μ g protein/ μ l) of 5 μ l and 2.5 μ l were run for each sample on 7.8% gels with a 4.3% stacker gel. Slabs we stained after electrophoresis with Coomarsie blue and bands were quantified by densitometry. Statistical analysis was by paired t-test is assessing ageing effects, and by oneway analysis of variance for comparing low versus intermediate pH groups.

RESULTS AND DISCUSSION

Figure 1 shows the position of the 24 samples used in this study relative to the polynomial regression line showing the relationship betwee WB peak shear force and pH_{ult} for the whole population of 156 samples from which the 24 were selected. The overall pattern is similar to be of just below 6.0.

Characteristics of the low and intermediate pH groups (Table 1) were as intended with respect to pH_{ult} and WB peak force, but the scatter for the intermediate group was wider because there were fewer samples to select from in that region. The decrease in peak force from 1 to 20 days was greater for the Inter-pH group.

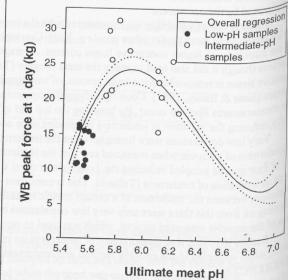


Table 1: Characteristics of the low and intermediate (Inter) pH_{ult} groups. Low-pH Inter pH Ga

Low-pn	Inter-DH	S"	
12			
5.55		0.15***	
	5.50	0.15	
13.01	23 43	3.76***	
10.05		2.85***	
2.96		3.10**	
	7.10	5.10	
81.7	823	2.0 ns	
84.0		3.2 *	
-2.3		3.1*	
1.52		0.17 ns	
	12 5.55 13.01 10.05 2.96 81.7 84.0 -2.3	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	

Pooled within-group standard deviation; ns = P > 0.05; * = P < 0.05; ** = P < 0.01*** = P < 0.001.

^b Myofibrillar fragmentation index (higher values indicate greater fragmentation).

Figure 1: The overall cubic-fit regression line between p^{egk} force and pH_{ult} for all 156 samples with 95% confidence intervals (R²=61%), together with the 24 samples of this study

MFI values did not differ between the two groups at 1d, but were greater for the Inter-pH group after 20d ageing and the increase from 1 to 2 days was greater for that group. Sarcomere lengths were not significantly shorter for the Inter-pH group as they were for Purchas (1990).

Differences between the Low-pH and Inter-pH groups in the concentration of nine myofibrillar proteins as assessed by SDS-PGE are shown Table 2 at 1-day of ageing and as the change from 1 to 20 days of ageing. At 1 day the Inter-pH group had significantly lower AA, ACT of

KD30, while the change from 1 to 20 days for the Inter-pH group was less for MHC and greater for ACT. The fact that KD30 was lower for the less lender Inter-pH samples is consistent with the fact that it is usually at lower levels in tougher unaged meat. The size of the effects in Table 2, however, are small compared with the differences in the average shear forces (Table 1). Correlations (n=24) also provided limited support for differences in shear force being attributable to variation in myofibrillar breakdown. Thus,

I(1d WB peak f).(1d MFI) = -0.04

 $r_{(20d WB pcak f).(20d MFI)} = 0.07$

 $\Gamma(1d WB peak f).(1d KD30) = -0.44*$

 $f_{(20d WB peak f)}(20d KD30) = -0.26$

The tendancy for higher peak forces to be associated with lower KD_{30} KD_{30} values at day 1 was supported by a significant negative contral to day 20 ^correlation between the change in peak force from day 1 to day 20 and the and the change in MFI over that period $(r = -0.39^*)$.

> Figure 2: Concentration (on a log scale) of the 9 myofibrillar proteins defined in Table 2, for samples at 1 and 20 days ageing.

See footnote to Table 1

The ageing effect on the 9 myofibrillar proteins (Figure 2) is generally consistent with other reports (reviewed by Koohmaraie 1992) in showing that TROT concentration declined and that of KD30KD₃₀ proteins increased with ageing.

Myofibrillar protein levels given in Table 2 and Figure 2 are the averages of values of values of the second of th Values obtained from SDS-PAGE bands that received a 2.5µl and a 5µl load. A measure of the linearity of the quantification method of these two loads is Provided by results in Figure 3, where it is shown that the quantity of BSA estimate the answer of t estimated by densitometry increased at a slightly decreasing rate as the amount added added to the myofibrillar extract increased from 0 to $1.8\mu g/ml$, and that the relation relationship was similar when a 2.5 or 5.0 μ l load was applied. Over the two BSA BSA levels used in this study (0.5 or 1.0µg) the relationship was close to linear. Estimate $E_{stimates}$ of the concentrations of myofibrillar proteins based on either the 2.5 or solution of the concentrations of myofibrillar proteins based on either the 2.5 o_{Γ} 5.0µl loads corresponded satisfactorily except that MHC levels were higher for the notation of this protein for the 2.5µl sample, presumably because the high concentration of this protein \log_{10} to 10^{-10} led to it being on the non-linear part of the curve.

In conclusion, it appears that, in this set of samples of beef longissimus muscle, there we have a state that the greater toughness of the there was very limited evidence to suggest that the greater toughness of the internet of myofibrillar protein intermediate-pH samples was due to a lower level of myofibrillar protein breakdown over a 20-day ageing period.

REFERENCES

Bouton, P.E., Howard, A. & Lawrie, R.A. (1957). CSIRO Div. Food Pres., Tech D. Tech. Paper No. 6, pp 1-23. Claeys, E., Uytterhaegen, L., Buts, B. & Demeyer, D. (1995). Meat Science, **39**, 177-193.

Jeremiah, L.E., Tong, A.K.W. & Gibson, L.L. (1991). Meat Science, **30**, 97-114. Johnson, D.D. & Hargrove, D.D.

Johnson, M.H., Calkins, C.R., Huffman, R.D., Johnson, D.D. & Hargrove, D.D. (1990). J.Anim.Sci., 68, 2371-2379.

Koohmaraie, M. (1992). Biochimie, 74, 239-245. ^{Ourma}raie, M. (1992). Biochimie, 74, 239-245. Olsson, U., Wahlgren, N.M. & Tornberg, E. (1995). Proc 41st Internat.Congr.Meat Sci. Technol. 41, 614-615. Purchas, R.W. (1990). Meat Science, 27, 129-140.

Purchas, R.W. (1990). Meat Science, 21, 129-140. Purchas, R.W. & Aungsupakorn, R. (1993). Meat Science, 34, 163-178.

Purchas, R.W. & Aungsupakorn, R. (1993). Meat Science, 34, 105-176. Watanak, R.W., Hartley, D.G. & Yan, X. (1997). New Zealand J.Agric.Res. (Submitted for publication). Watanabe, A., Daly, C.C. & Devine, C.E. (1996). Meat Science, 42, 67-78. Wull Devine, A., Daly, C.C. & Devine, C.E. (1996). Meat Science, 42, 67-78.

Wulf, D.M., Tatum, J.D., Green, R.D., Morgan, J.B., Golden, B.L. & Smith, G.C. (1996). J.Anim.Sci., 74, 2394-2405. Yu, L.P. & Lee, Y.B. (1986). J.Food Sci., 51, 774-780.

Table 2: The concentration of 9 myofibrillar proteins following I day of ageing and the change in concentration from day 1 to day 20 of ageing.

Myofibrillar protein(s) (µg/mg myofibrillar protein)	Abbrev	After 1 day ageing			Change: 1 to 20 days		
		Low	Inter	S ^a	Low	Inter	S ^a
Myosin heavy chain	MHC	156.1	145.8	31.1ns	21.5	-9.3	35.2*
Alpha actinin	AA	24.0	19.0	5.4*	-2.6	-3.7	6.6ns
Actin	ACT	132.0	112.7	17.8*	5.0	-20.0	26.7*
Troponin-T	TROT	21.0	16.9	7.9ns	6.4	-3.9	6.6ns
26-37 kDa proteins	KD30	20.0	14.9	5.7*	-6.7	-5.9	9.1ns
Myosin light chain 1	MLC1	16.6	14.1	4.1ns	3.0	-3.2	6.1ns
Troponin-I & -C	TRIC	11.6	9.5	3.8ns	0.7	-1.7	4.7ns
Myosin light chain 2	MLC2	23.6	22.9	4.0ns	2.9	4.9	6.7ns
Proteins less than 18kDa	LT18	6.1	3.3	6.3ns	-1.6	-5.9	6.9ns

