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## The effects of the calpain proteolytic system on meat tenderisation rates in different ovine skeletal muscles

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Key words: calpains, calpastatin, meat tenderness, ovine

### Introduction

Muscle fibre composition has an influence on postmortem meat tenderisation of ruminants (Olson et al., 1976, Dransfield *et al.*, 1981, Koohmaraie et al., 1988a, Ouali, 1990). The proteolytic enzymes  $\mu$ - and m-calpain, which are involved in cellular events mediated by calculated their specific inhibitor calpastatin are involved in postmortem tenderisation of meat by degrading myofibrillar proteins thus causing a weakening of myofibrillar structure (Koohmaraie *et al.*, 1988b). Differences in activity of the ovine calpain system may also be influenced muscle fibre composition (Ouali and Talmant 1990; Whipple and Koohmaraie 1992). Earlier studies suggest a relationship between calpain rates and calpain efficiency as indicated by the calpain:calpastatin ratio (Ouali and Talmant, 1990). These earlier studies report a limited moder of muscles representative of different fibre typing groups. The aim of this study was to determine the extent to which differences in calpain calpastatin activities may account for differences in ultimate tenderness and rates of meat tenderisation in a wide range of ovine muscles we differ in muscle fibre composition.

### **Materials and Methods**

Lambs at 4 months of age were fed a pelleted diet (lucerne 60% and barley 30%) and water *ad libitum* for the 3 weeks. Muscles were collicities and the second seco within 30 min of slaughter and ranged in fibre type composition. Trial 1 (n=9): muscles composed of predominantly type I (slow-twitch oxidative) fibres; *M. masseter* (M), *M. supraspinatus* (SS), *M. vastus intermedius* (VI): type IIA (fast-twitch oxidative-glycolytic) fibres; *M. masseter* (M), *M. supraspinatus* (SS), *M. vastus intermedius* (VI): type IIA (fast-twitch oxidative-glycolytic) fibres; *M. masseter* (M), *M. supraspinatus* (SS), *M. vastus intermedius* (VI): type IIA (fast-twitch oxidative-glycolytic) fibres; *M. masseter* (M), *M. supraspinatus* (SS), *M. vastus intermedius* (VI): type IIA (fast-twitch oxidative-glycolytic) fibres; *M. masseter* (M), *M. supraspinatus* (SS), *M. vastus intermedius* (VI): type IIA (fast-twitch oxidative-glycolytic) fibres; *M. masseter* (M), *M. supraspinatus* (SS), *M. vastus intermedius* (VI): type IIA (fast-twitch oxidative-glycolytic) fibres; *M. masseter* (M), *M. supraspinatus* (SS), *M. vastus intermedius* (VI): type IIA (fast-twitch oxidative-glycolytic) fibres; *M. masseter* (M), *M. supraspinatus* (SS), *M. vastus intermedius* (VI): type IIA (fast-twitch oxidative-glycolytic) fibres; *M. masseter* (M), *M. supraspinatus* (SS), *M. vastus intermedius* (VI): type IIA (fast-twitch oxidative-glycolytic) fibres; *M. masseter* (M), *M. supraspinatus* (SS), *M. vastus intermedius* (VI): type IIA (fast-twitch oxidative-glycolytic) fibres; *M. masseter* (M), *M. supraspinatus* (SS), *M. vastus intermedius* (VI): type IIA (fast-twitch oxidative-glycolytic) fibres; *M. masseter* (M), *M. supraspinatus* (SS), *M. vastus* (SS), *M.* biceps femoris (BF), M. gastrocnemius (G), M. longissimus dorsi (LD), M. psoas major (PM): type IIB (fast-twitch glycolytic) fibres: M. semitendinosus (ST), M. tensor fascia latae (TFL): type II fibres; M. cutaneous trunci (CT). Trial 2 (n=8): the same muscles were used even CT and G were replaced by the type IIA muscle *M. semimembranosus* (SM) and type IIB muscle *M. gracillus* (GR) (Briand et al., 1981; and Tamate 1988). Calpactation III and marked and Tamate 1988). and Tamate, 1988). Calpastatin, µ- and m-calpain were extracted from fresh 5 g samples and separated on a DEAE-Sephacel column using stepwise NaCl gradient (Wheeler and Koohmaraie 1991, Sainz et al., 1992). Calpain activities were determined using casein as the substra (Hammarsten, Merck, Germany). One unit of calpain activity was defined as the amount of enzyme required to increase 1.0 absorbance unit 278 nm in 60 min at 25 °C. One unit of calpastatin activity was defined as the amount of calpastatin required to inhibit one unit of m-calp Shear force analysis was carried out on muscles aged at 15°C for 1 and 3 days postmortem (trial 1) or 1, 2 and 4 days postmortem (trial 2). Samples were cooked to an internal temperature of 75°C and analysed for peak shear force by shearing perpendicular to the fibre axis using MIRINZ pneumatic tenderometer. There were no shear force measures for the CT, G and M due to muscle size. Tenderisation aging rates were determined using an exponential decay equation as described by Dransfield et al. (1981). Restricted maximum likelihood (REML). Genstat statistical package, was used to obtain means for the muscles. This also provided likelihood ratio tests (asymptotically chi-squared this effect and the relationships between tenderness and activities of calpastatin and calpains on a between and within muscle mean basis

## Results

There were significant differences among muscles in calpastatin (p<0.001),  $\mu$ -calpain (p<0.05) and m-calpain (p<0.001) activities (Table )  $\mu$ -calpain:calpastatin (p<0.001) and m-calpain:calpastatin (p<0.001) ratios were different among the muscles. There was also a significant difference among muscles for shear force measures at each time point (p<0.001) and aging rates (p<0.01, table 2). The fibre typing groups muscles does not account for all the muscle variation and this residual variation was used as the error for testing between type differences. Calpastatin (p<0.05) and m-calpain activities (p<0.05) were higher in type I muscles in comparison to type II muscles. There were no significant differences in calpastatin and m-calpain activities between type IIA and type IIB muscle. There was also no significant difference in  $\mu$ -calpa activity between the different fibre types. The aging rates were greater (p<0.05) in type IIA muscles in comparison to type IIB muscles. There was also no significant difference in  $\mu$ -calpa no significant difference in aging rate between type I muscles in comparison to type IIB muscles. There were for the fibre type IIB muscles. There was also no significant the fibre type I muscles. There was also no significant difference in  $\mu$ -calpa no significant difference in aging rate between type I muscles in comparison to type IIB muscles. There of a muscle has an influence on the calpain system and the rate of meat aging.

Calpastatin activity had a positive association (p<0.01) and  $\mu$ -calpain:calpastatin ratio had a negative association (p<0.05) with ultimate she force, between muscle means. However, these effects were mainly due to the VI. After removal of the between muscle variation, the within calpain:calpastatin ratio had a negative association (p<0.05),  $\mu$ -calpain had a negative association (p<0.05) and  $\mu$ -calpain:calpastatin ratio had a negative association (p<0.05) with ultimate she force, between (p<0.01) with ultimate she association (p<0.05),  $\mu$ -calpain had a negative association (p<0.05) and  $\mu$ -calpain (p<0.1) with ultimate shear force. These data suggest that higher calpastatin activity and lower calpain activity results in tougher between  $\mu$ -calpain:calpastatin ratio (p<0.1). These data suggest that higher calpastatin activity and lower calpain activity influences ultimate tenderness and the rate of meat aging.

#### Discussion

In the present study results demonstrated that different ovine muscles differ in their levels of calpastatin inhibitory activity and  $\mu$ - and  $m^{-c^{2\beta}}$  proteolytic activity. Calpastatin and m-calpain activities were influenced by metabolic and contractile type of a muscle, with the activities higher in slow-twitch muscles with higher oxidative activity compared to fast-twitch muscles with oxidative-glycolytic activity and  $g^{realtr}$ 

<sup>glycolytic</sup> activity. Previous reports also suggest that calpastatin activity may be influenced by metabolic and contractile muscle types (Ouali and Talpastatin activity may be influenced by metabolic and contractile muscle types (Ouali and Talpastatin activity). Talmant, 1990; Whipple and Koohmaraie, 1992). The m-calpain activity was higher in type I muscles, although the µ-calpain activity was not influenced by muscle fibre composition in the present study. Whipple and Koohmaraie (1992) report that both  $\mu$ - and m-calpain activities were <sup>greater</sup> in type I muscles compared to type II muscles. In contrast, Ouali and Talmant (1990) report the m-calpain is influenced only by metabolic type of a muscles, with the activity being higher in type IIB whiter muscles compared to type I and IIA redder muscles.

The meat aging rates were greater in type IIA fast-twitch oxidative-glycolytic muscles than type IIB fast-twitch glycolytic muscles. There was no different suggests aging rates were greater difference in aging rates between type I muscles in comparison to type II muscles. In contrast a previous report suggests aging rates were greater in fast-twitch than in slow-twitch muscles (Ouali and Talmant, 1990).

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In the present study the positive association between calpastatin activity and the negative association among the calpains and shear force of aged mean and the negative association among the calpains and shear force of aged inhibitory actions of meat, and the positive association among the calpain:calpastatin ratios and aging rates within muscles suggests the increased inhibitory actions of calpastation among the calpain:calpastatin ratios and aging rates within muscles suggests the increased inhibitory actions of calpastatin on the calpains results in decreased proteolytic activity of calpain resulting in tougher meat and differences in the rate at which meat ages. Earlier studies investigating ovine muscles report a relationship between calpastatin activity and the amount of postmortem proteolysis (Whinst Whipple and Koohmaraie, 1992) and a relationship between meat tenderisation aging rates and calpain efficiency as indicated by the calpained. calpain: calpain: calpain: calpain cal  $O_{uali}^{calpastatin}$  ratio (Ouali and Talmant, 1990). There was no relationship between calpastatin and aging rate in the entry of  $U_{uali}^{calpastatin}$  and Talmant (1990). This study has clearly shown that the fibre composition of a muscle influences the activity of the calpain system and the rate of  $U_{uali}^{calpastatin}$  and  $U_{uali}^{calpastatin}$ the rate of meat aging. The differences in calpastatin and calpain activities within muscles influence the degree of ultimate tenderness and the rate of  $a_{pinc}$ . of aging in ovine muscles.

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Muscle	calpastatin	µ-calpain	m-calpain
<sup>1</sup> Longissimus dorsi	2.37	0.906	1.353
<sup>1</sup> Biceps femoris	2.57	1.030	1.397
<sup>1</sup> Masseter	2.62	0.894	1.870
<sup>2</sup> Semimembranosus	2.68	0.946	1.324
<sup>1</sup> Semitendinosus	2.68	0.898	1.283
<sup>1</sup> Tensor fasciae latae	2.82	0.950	1.392
<sup>2</sup> Gracillus	2.84	0.936	1.470
<sup>2</sup> Cutaneous Trunci	3.04	0.815	1.199
<sup>1</sup> Psoas major	3.07	0.833	1.288
<sup>1</sup> Supraspinatus	3.21	0.952	1.396
<sup>2</sup> Gastrocnemius	3.22	1.001	1.536
<sup>1</sup> Vastus intermedius	4.92	1.004	1.730
sed			
within group 1	0.192	0.06	0.09
between groups 1 and 2	0.250	0.08	0.11
within group 2	0.299	0.09	0.14
df	157	151	170
Effect of muscle	***	*	***

subscript letters represent groupings for comparisons.

Table 1. Effects of muscle on calpastatin, µ- and m-calpain activities (U/g muscle)

#### Table 2. Effects of muscle on the rates (k)of postmortem aging

Muscle

rates (k)

<sup>1</sup> Biceps femoris	.0343
<sup>2</sup> Gracillus	.0181
<sup>1</sup> Longissimus dorsi	.0426
<sup>2</sup> Psoas major	.0325
<sup>2</sup> Semimembranosus	.0353
<sup>1</sup> Semitendinosus	.0183
<sup>1</sup> Supraspinatus	.0387
<sup>2</sup> Tensor fasciae latae	.0453
<sup>2</sup> Vastus intermedius	.0387
sed	

within group 1	0.00648
between groups 1 and 2	0.00807
within group 2	0.00896
TFL vs groups 1 and 2	0.0102
df	84

Effect of muscle

subscript letters represent groupings for comparisons Data are expressed as means.

\*, 0.05 \*\*\*, p<0.001

Data are expressed as means.

\*, p<0.01