EFFECT OF PASTURING OR VITAMIN E SUPPLEMENTATION ON COLOUR AND METHMYOGLOBIN FORMATION IN BEEF HELD UNDER DIFFERENT PACKAGING CONDITIONS

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Keywords: MAP, aerobic packaging, α -tocopheryl acetate, beef steaks, colour

Background

The colour of fresh meat is a key factor used by the consumer to indicate meat quality and freshness and its rate of sale is directly related to the degree of discolouration (MacDougall and Taylor, 1975). The development of Modified Atmospheric Packaging (MAP) has provided a new approach for extending the shelf life of fresh meat products (Head and Media 1997). approach for extending the shelf-life of fresh meat products (Hood and Mead, 1993). MAP for fresh red meats involves the use of high approach for extending the shell-life of fresh meat products (Hood and Mead, 1993). MAP for fresh red meats involves the use of the concentrations of oxygen to maintain or produce a deep layer of bright red oxymyoglobin and lower concentrations of carbon dioxide which have a bacteriostatic effect (Church and Parsons, 1995). Dietary α -tocopheryl acetate supplementation in cattle diets has been shown to of MAP and other forms of packaging, on colour and methmyoglobin formation in *M. gluteus medius* produced by varying the dietary conditions of crossbreed Friesian cattle. conditions of crossbreed Friesian cattle.

Materials and Methods Crossbreed (Friesian x Charolais) heifers (n=12) were divided into four groups (n=3). Group 1 was overwintered indoors and fed a basel concentrate diet containing 20 mg α -tocopheryl acetate/head/day. Group 2, also overwintered indoors, was fed a supplemented concentrate indoors and fed a basel concentrate diet containing 20 mg α -tocopheryl acetate/head/day. Group 2, also overwintered indoors, was fed a supplemented concentrate indoors and fed a basel concentrate diet containing 20 mg α -tocopheryl acetate/head/day. Group 2, also overwintered indoors, was fed a supplemented concentrate indoors and fed a basel concentrate diet containing 20 mg α -tocopheryl acetate/head/day. Group 2, also overwintered indoors, was fed a supplemented concentrate indoors and fed a basel concentrate diet containing a supplemented concentrate diet containing 20 mg α -tocopheryl acetate/head/day. Group 2, also overwintered indoors, was fed a supplemented concentrate indoors and fed a basel concentrate diet containing a supplemented concentrate diet concentrate diet containing a supplemented concentrate diet containing a supplemented concentrate diet concentrate diet containing a supplemented concentrate diet concentrate d diet containing 3000 mg α -tocopheryl acetate/head/day for 50 days prior to slaughter. Both of these groups received silage *ad libitum*. Group 3 was overwintered indoors, fed basal concentrate and silage diets and pastured for 6 weeks on late Spring (April-May) grass prior of slaughter Group 4 was pastured from April and clouchtered in late Annual (Orthered in late Annual Content of the Spring (April-May) grass prior of the standard of the Annual Spring (April-May) grass prior of the standard of the stan slaughter. Group 4 was pastured from April and slaughtered in late Autumn (October). Following slaughter, carcass sides were chilled (4°C x 24 hours). *M. gluteus medius* was removed from each carcass, vacuum packed and frozen (-20°C x 3 months).

Steaks (2.5 cm thickness) were cut from each muscle and duplicate cores (2.5 cm diameter) were taken from each steak. These were packaged aerobically, under vaccum and in 80 : 20 (O_2 : C O_2) MAP prior to refrigerated (4°C / 616 lux fluorescent lighting) display for 10 days. Vacuum packaged samples were stored in darkness and allowed to bloom, following overwrapping, for a minimum of 3 hours prior to colour analysis. Meat cores were MAP and vacuum packaged union law endowed to bloom. analysis. Meat cores were MAP and vacuum packaged using low oxygen permeable (8-12 cm³/m²/24 hours) polystyrene/EVOH/polyethyleth and (45 cm³/m²/24 hours) polyamide/polyethylene films, respectively. Aerobically packaged samples were overwrapped with oxygeth permeable (6000-8000 cm³/m²/24 hours) polyvinyl-chloride film.

Hunter 'a' values for meat cores were recorded using a Minolta Colorimeter. Metmyoglobin content was determined by the method of Krzywicki (1979), using a Perkin-Elmer (Lambda 2) spectrophotometer. α -Tocopherol concentrations in muscle tissues was determined using the extraction procedures of Bieri et al. (1975) with the modifications of Butting and Dialache (1924). using the extraction procedures of Bieri et al. (1975) with the modifications of Buttriss and Diplock (1984) and quantified by the HPLC method of Sheehy et al. (1993).

Results

There was a significant (p < 0.001) difference in α -tocopherol levels between the α -tocopheryl acetate supplemented group (Group 1, 7.13) general, metmyoglobin formation decreased with respect to dietary groups in the order: Spring pastured group 3 > Autumn pastured group> basal group 2 > supplemented group 1. Metmyoglobin formation was delayed under MAP (Figure 2a) but promoted by vacuum packaging (Figure 2c), Metmyoglobin formation in perobically (Figure 2b) and the matrix of the second sec (Figure 2c), Metmyoglobin formation in aerobically (Figure 2b) packaged meat lay between those levels formed under other packaging

Conclusions

Dietary supplementation of cattle with α -tocopheryl acetate resulted in significant increases in α -tocopherol concentrations in meat tissue of the supplementation also increased colour stability of a law to the supplementation also increased colour Supplementation also increased colour stability and slowed down the development of metmyoglobin to a greater extent than observed in other dietary groups. The colour stability of supplemented methyoglobin to a greater extent than observed in other for the stability of supplemented methyoglobin to a greater extent than observed in other for the stability of supplemented methyoglobin to a greater extent than observed in other for the stability of supplemented methyoglobin to a greater extent than observed in other for the stability of supplemented methyoglobin to a greater extent than observed in other for the stability of supplemented methyoglobin to a greater extent than observed in other for the stability of supplemented methyoglobin to a greater extent than observed in other for the stability of supplemented methyoglobin to a greater extent than observed in other for the stability of supplemented methyoglobin to a greater extent than observed in other for the stability of supplemented methyoglobin to a greater extent than observed in other for the stability of supplemented methyoglobin to a greater extent than observed in other for the stability of supplemented methyoglobin to a greater extent than observed in other for the stability of supplemented methy supplem dietary groups. The colour stability of supplemented meat down the development of metmyoglobin to a greater extent than observed in the meat from all dietary groups compared to other production are the development of MAP. In general, MAP promoted better colour for meat from all dietary groups compared to other packaging methods.

References

Arnold, R.N., Arp, S.C., Scheller, K.K., Williams, S.N. and Schaefer, D.M. (1993). J. Anim. Sci., 71, 105-118. Bieri, J.G. Tolliver, T.J. and Catignani, G.L. (1979). Am. J. Clin. Nutr., 32, 2143-2149. Buttriss, J.L. and Diplock, A.T. (1984). Methods in Enzymol., 105, 131-138. Church, I.J. and Parsons, A.L. (1995) J. Food Sci., 67,143-152. Hood, D.E. and Mead, G.C. (1993). Principles and Applications of Modified Atmosphere Packaging of Food. Blackie Academic and Professional, London, pg 269-298. Krzywicki, K. (1979). Meat Sci., 3, 1-10. MacDougall, D.B. and Taylor, A.A. (1975) J. Food Tech., 10, 339-347. Sheehy, P.J.A., Morrissey, P.A. and Flynn, A. (1993). Brit. Poultry Sci., 34, 367-381.

Acknowledgements

This research has been part-funded by grant aid under the Food Sub-Programme of the Operational Programme for Industrial Development which is administered by the Department of Agriculture, Food and Forestry and supported National and EU funds



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