EXAMINATION OF TRENBOLON ACETATE RESIDUES AFTER IMPLANTATION OF REVALOR S

FATTENING STEERS

Snežana Saičić, Saša Janković, Aurelija Spirić, Milovan Đorđević

Institute of Meat Hygiene and Technology, 11000 Belgrade, P.O.Box 33-49, Yugoslavia

Key words: Steers, trenbolon acetate, determination of residues

BACKGROUND Anabolic implants used to improve growth rate and feed efficiency of cattle during fattening thus resulting economic benefits. One of the growth promoters used in raising animals for slaughter is androgen - trenbolon acetate (TBA), whose active component is trenbolon (TBOH). It is most frequently applied alone or in combination with estrogen - estradiol. In that case, according to the investigation of Gerken et al., (1995) and Bartlea et al. (1992), its effect is intensified. Many investigations are performed with the aim to develop new methods or to improve the present methods for TBOH determination (Degan G. et al., 1989; ; Daeseleire E. et al., 1991; Bagnati R. and Fanelli R., 1991; Lagana A. and Marino A., 1991; Daeseleire E. et al., 1992; Hevitt et al., 1993).

OBJECTIVES The aim of this work is to examine the residues of trenbolon acetate, after its application in combination with 17.4 estradiol in fattening steers of Simental breed, and to establish the time of their excretion from the organism.

METHODS The examinations were performed on a private cattle farm. Ten steers of Simental breed, eight months old, were assigned randomly. The first five steers were used as a control group (no implant), and the other five were treated with Revalor S. The Implantation S (24mg of 17-β estradiol and 120mf of trenbolon acetate) was surgically implanted under the skin behind the ear. Sixty day after implantation, analysis of faeces and urine did not show any excretion of trenbolon residues from the organism. The first implantation was done. The samples of faeces and urine were always taken at the same time. The steers were slaughter 120 days the first implantation of anabolic and the residues of trenbolon were examined in muscle tissue, as well. For determination of The residues, we used the HPIC method developed by Laitem et al., (1978) and the Laboratory residue analysis (Bilthoven, Holland) procedure was modified in our laboratory to achieve the necessary analytical requirements. The experiment (method) involving the HPIC column. The experimentally established detection limit was 5μg/kg; the retention time was 5 minutes. With the almost through the HPIC column. The experimentally established detection limit was 5μg/kg; the retention with the samples we obtained from the recovery percentage, the blank and spiked samples were analyzed in conjunction with the samples we obtained treated animals. The recovery we obtained was approximately 85%.

RESULTS AND DISCUSSION The established quantities of TBOH residues in faeces and urine are presented in Figures 1. and 2. The residues of anabolic and the elimination rate of residues from the body depends on the way of anabolic administration composition implant, animal type and the expired time after drug application (Karg H. et al., 1984).

From our examination, it is evident that there is a hugh variability of TBOH concentration in urine and faeces. Similar results we obtained by other investigators as well (Karg H. et al., 1984; , Henricks D.M., 1981). In addition, higher quantity of TBOH were obtained in samples of faeces in relation to urine. Maximum quantities of TBOH residues in both excretions were established 24h, namely after the application of Revalor S and they are in accordance with the findings of Karg et al. (1984). However, sudden decrease of the concentration is observed in faeces 40 days after implantation and in urine after 15 days. After slaughter of steers, TBOH residues not established in muscle tissue. It can be concluded that TBOH excretion from the organism occurred prior to steer slaughter

CONCLUSION On the basic of the obtained results it can be concluded that the excretion of TBOH in fattening steers of Simental brown happened two months after the application of Revalor S. Having in mind that there is variability in concentrations, it is necessary repeat the experiment with a higher number of animals

LITERATURE 1. Bagnati R., Fanelli R., 1991, J. Chromatogr., Jun 28, 547(1-2), 325; 2. Bartle S.J., Preston R.L., Brown R.E., Grant 1992, J. Anim. Sci., 70(5), 1326; 3. Daeseleire E., de Guesquire A., van Peteghen C., 1991, J. Chromatogr., 2, 562; 4. Daeseleire Guesquire A., van Peteghen C., 1992, J. Chromatogr., Oct 30(10), 400; 5. Degand G., Schmite P., Maghuin-Rogister G., Chromatogr., 489, 235; 6. Gerken C.L., Tatum J.D., Morgan J.B., Smith G.C., 1995, J. Anim. Sci., 73(11), 3317; 7. Henricks D.M., 1981, Proc. Intern. Symp. "Steroids in animal production", april 1980, Ed. H. Jasiorowski, Warsaw Agricultural University, 161; 8. Hewit Blanchflower W.J., Mc Caughey W.J., Elliot C.T., Kennedy D.G., 1993, J. Chromatogr., Jun 11 639(2), 185; 9. Karg H., Meyer H.H.D., K., Landwehr M., Hoffmann B., Schopper D., 1984, In book: Manipulation of growth in farm animals, Ed. by J.F. Roche Callaghan, Martinns Nijhoff Publischers, Boston; 10. Lagana A. and Marino A., 1991, J. Chromatogr., Dec 27, 558(1-2), 89; 11. Jailed Gaspar P., Bello I., 1978, J. Chromatogr., 147, 537.

