

## GROWTH OF MIXED CULTURES OF *LISTERIA* AND *PSEUDOMONAS* SPP. IN BEEF MEAT AND THEIR PREDICTION USING POLYNOMIAL MODELS

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### Background

Among the psychrotrophic Gram-negative bacteria that can multiply in meat products, dairy products and vegetables, only a few have an influence on the quality and the shelf-life of these products and can be responsible for considerable financial losses. The principal flora responsible for such spoilage during aerobic storage are the pseudomonads. *Pseudomonas* spp. grew faster than other strains present in meat at temperatures between 2 and 15°C. Several pathogenic bacteria also grow within this temperature range and may cause foodborne outbreaks: *Listeria monocytogenes* has been shown to be responsible for outbreaks of food poisoning in France and in many countries (Rocourt and Bille, 1997).

### Objective

To predict the growth of both spoilage *Pseudomonas* spp. and pathogenic *Listeria* in meat products

### Methods

A first set of experiments were performed in decontaminated beef meat that was inoculated with *Listeria* or *Pseudomonas* strains. Strains were grown alone or together. The second set of experiments was performed in naturally contaminated meat that was purchased at the supermarket, meats were available in oxygen-permeable bags (experiments B and D) or in vacuum packs (experiments A and C) (Figure 1). The dominant *Pseudomonas* spp. were grown alone or in the presence of inoculated *Listeria*. Meat was stored under aerobic conditions at low temperatures. Bacterial counts were performed on: Tryptone Soja Agar (Difco, OSI, Maurepas, France) for non selective growth; on Palcam Agar (Merck, Nogent-sur-Marne, France) for *Listeria*; on *Pseudomonas* Agar (Oxoid, Unipath Ltd, Basingstocke, England) for *Pseudomonas*.

Growth parameters (A, logarithmic increase in the bacterial population, L, lag time, GT, generation time) were calculated by fitting the growth curves using a modified Gompertz equation (Zwietering et al., 1990). The *Listeria* model of Lebert et al. (1998) and the three *Pseudomonas* models of Robles (1999) were used as they took the growth variations into account and calculate a growth response interval.

### Results and discussion

Three models for one rapid and one slow growing strain of *P. fragi* and one slow growing strain of *P. fluorescens* were developed in a meat broth and tested in naturally-contaminated meat. TSA and CFC counts were similar indicating that *Pseudomonas* spp. were dominant. Table 1 shows that experimental L were longer than predicted L but that the observed GT were between the GT predicted by *Pfr162* and *PfrK1* model. The *Pseudomonas* models can be used to predict the growth of *Pseudomonas* spp. in meat.

In decontaminated meat inoculated with *Listeria* or *Pseudomonas* strains (Table 2), all the models provided satisfactory predictions. In mixed population experiments (Table 2), generation times for *Listeria* were similar when grown alone or with *P. fragi*, and were predicted satisfactory by the models. A comparison of the growth parameters of *Pfr162* alone and with *Lm14* showed that they were similar if account was taken of the confidence intervals. When co-inoculated, the growth of neither organism was either inhibited or stimulated. No interactions were observed between the strains, and their growth parameters were satisfactorily predicted by their respective models.

In naturally-contaminated meat inoculated with *Listeria*, Figure 1 shows that the experimental points for *Pseudomonas* were within the curves generated by the three models. In the four cases, observed GT were well predicted by the models. In experiments A and C, growth was inside the curves predicted by *PfrK1* and *Pfr158* models (slow models). In experiments B and D, growth was near the curves predicted by *Pfr162* models (fast model). A and D experiments (meat stored in vacuum packs) were characterised by longer lag times than those observed in experiments B and D (meat stored in oxygen-permeable bags): these differences can be due to the presence of different bacterial flora on the meat because of the way of storage. In meat stored in oxygen-permeable film, the microflora was already aerobic and mainly composed of *Pseudomonas* spp., that explains a small L. With vacuum-packaged meat, it took longer for the *Pseudomonas* to outgrow the anaerobic flora present in the meat. No *Listeria* growth was observed until *Pseudomonas* reached the stationary phase, slight increases (1 to 2 log(CFU)) were then observed. Predictions of *Listeria* growth were calculated for an A of 4.1 as found in Table 2 and *Lm14* model failed to predict the slow growth of *Listeria*.

### Conclusions

In experiments with mixed populations, three situations were observed: (1) in decontaminated meat, *L. monocytogenes* inoculated alone grew well at 6°C and the growth was correctly predicted by the model; (2) in decontaminated meat that was inoculated with *Listeria* and *Pseudomonas* strains, *L. monocytogenes* grew well and was not affected by the presence of the *Pseudomonas*, growth of both organisms were correctly predicted by the models; (3) in naturally contaminated meat inoculated with *L. innocua*, the strain did not grow until the *Pseudomonas* had reached the stationary phase. The models satisfactorily predicted the growth of *Pseudomonas* spp. but not that of *Listeria*.

In consequence, the *Lm14* model can not be used for refrigerated meat stored aerobically as it indicated a too 'fail-safe' prediction: meat had already reached a spoilage state even though no increase of *Listeria* was observed. The *Pseudomonas* models accurately predicted the growth of naturally occurring *Pseudomonas* spp.

## Pertinent literature

- Lebert, I., Bégot, C. and Lebert, A. (1998) Development of two *Listeria monocytogenes* growth models in a meat broth and their application to beef meat. Food Microbiol. 15, 499-509.
- Robles Olvera, V. (1999) Comparaison de la croissance de *Pseudomonas species* et *Listeria species* en milieu liquide et en viande de boeuf. Thèse de Docteur ès science, Université Blaise Pascal, Clermont-Ferrand II.
- Rocourt, J. and Bille, J. (1997) Foodborne Listeriosis. World Health statistics quartely 50, 67-73.
- Zwietering, M. H., Jongenburger, I., Rombouts, F. M. and Van't Riet, K. (1990) Modeling the bacterial growth curve. Appl. Environ. Microbiol. 56, 1875-1881.

Table 1: Observed and predicted values (lag time (L) and generation time (GT)) of *Pseudomonas* spp. grown on the surface of naturally contaminated beef muscle at a relative humidity of air  $\approx$  100% and pH 5.8. In small prints: lower and upper confidence intervals at 95%.

| T (°C) | logN <sub>0</sub> | A   | L obs. (h) | L pred. (h)          |                     |                     | GT obs. (h)   | GT pred. (h)         |                     |                     |
|--------|-------------------|-----|------------|----------------------|---------------------|---------------------|---------------|----------------------|---------------------|---------------------|
|        |                   |     |            | <i>Pfr</i> 162 model | <i>Pfr</i> K1 model | <i>Pf</i> 158 model |               | <i>Pfr</i> 162 model | <i>Pfr</i> K1 model | <i>Pf</i> 158 model |
| 6      | 4.4               | 5.8 | 19 (13-26) | 6.7 (3.9-11.6)       | 3.3 (0.8-13.5)      | 12 (5.8-24)         | 5.1 (4.5-5.6) | 4.3 (3.5-5.4)        | 6.1 (4.5-8.2)       | 6.5 (4.5-9.3)       |
| 6      | 3.0               | 7.5 | 17 (12-22) | 6.7 (3.9-11.6)       | 3.3 (0.8-13.5)      | 12 (5.8-24)         | 4.4 (4.1-4.8) | 4.3 (3.5-5.4)        | 6.1 (4.5-8.2)       | 6.5 (4.5-9.3)       |
| 4      | 3.0               | 7.4 | 18 (12-24) | 10.3 (5.5-19.3)      | 4.4 (0.9-22.3)      | 14 (6.4-32)         | 5.7 (5.2-6.2) | 6.2 (4.8-7.9)        | 8.8 (6.3-12.4)      | 10 (6.7-15)         |

Table 2: Observed and predicted lag times and generation times (h) for single strain or mixed cultures grown on the surface of decontaminated meat at 4°C, relative humidity  $\approx$  100%. In small prints: lower and upper confidence intervals at 95%.

*Pfr*162: *P. fragi* 162; *Lm*14: *L. monocytogenes* 14; *Li*CLIP: *L. innocua* CLIP 20595

| Experiments                          | Strains        | T (°C) | pH  | logN <sub>0</sub> | A   | L obs. (h) | L pred. (h)          |                    | GT obs. (h)   | GT pred. (h)         |                    |
|--------------------------------------|----------------|--------|-----|-------------------|-----|------------|----------------------|--------------------|---------------|----------------------|--------------------|
|                                      |                |        |     |                   |     |            | <i>Pfr</i> 162 model | <i>Lm</i> 14 model |               | <i>Pfr</i> 162 model | <i>Lm</i> 14 model |
| <i>Listeria</i> + <i>Pseudomonas</i> | <i>Pfr</i> 162 | 4      | 5.9 | 4.0               | 6.4 | 8 (4-12)   | 10 (6-16)            |                    | 5.2 (4.8-5.6) | 6.1 (5.0-7.3)        |                    |
|                                      | <i>Li</i> CLIP | 4      | 5.9 | 3.1               | 4.1 | 69 (41-97) |                      | 67 (10-446)        | 28 (23-33)    |                      | 26 (14-49)         |
| <i>Listeria</i>                      | <i>Lm</i> 14   | 4      | 6.3 | 4.1               | 4.8 | 36 (10-62) |                      | 56 (10-301)        | 33 (29-36)    |                      | 28 (13-39)         |
| <i>Pseudomonas</i>                   | <i>Pfr</i> 162 | 4      | 5.7 | 4.0               | 6.2 | 26 (20-31) | 11 (5-20)            |                    | 5.2 (4.5-5.8) | 6.3 (4.8-8.2)        |                    |

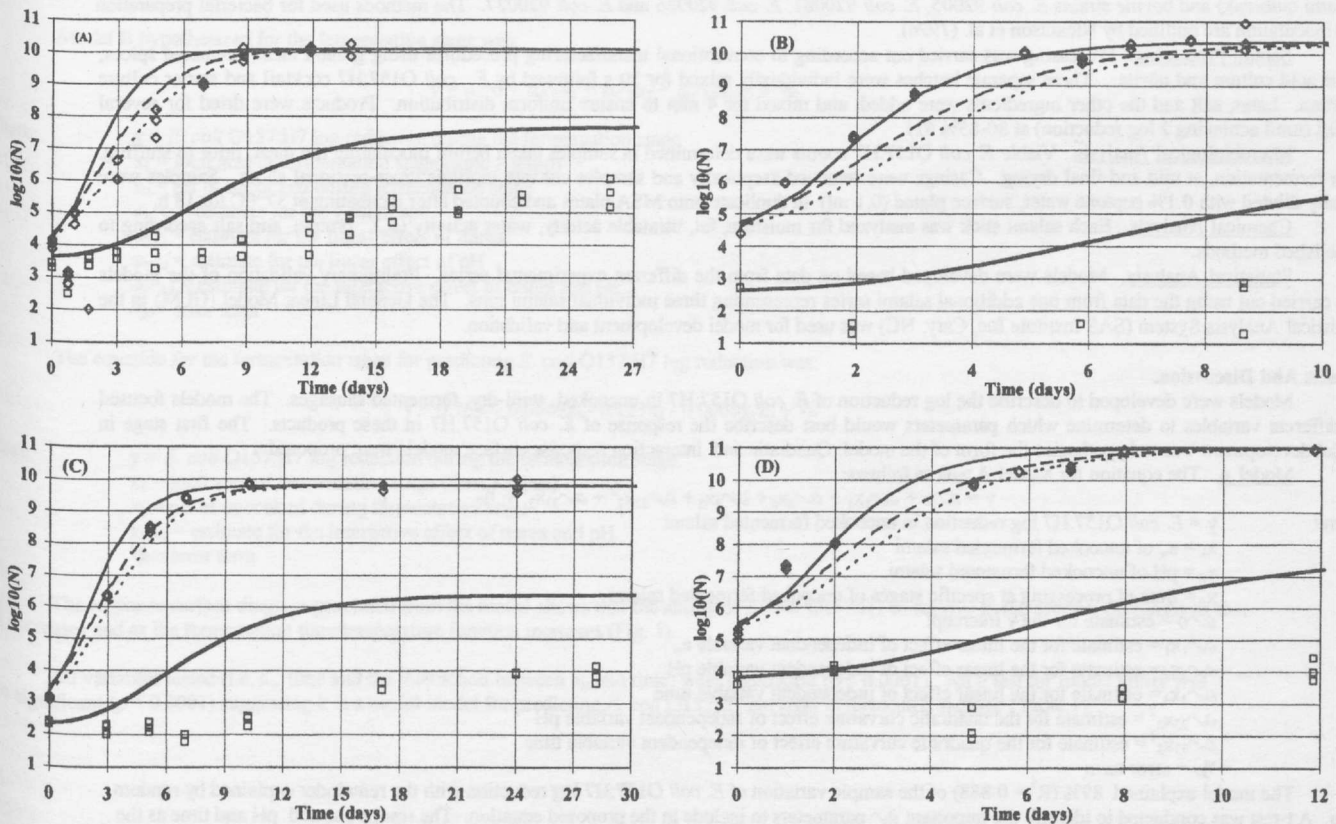


Figure 1: Growth of *Pseudomonas* spp. in naturally-contaminated meat re-inoculated with *Listeria* strains at 6°C, relative humidity of air  $\approx$  100%, pH 5.8. The meats were available in oxygen-permeable bags (B and D) or in vacuum packs (A and C).

Experimental data: *Pseudomonas* spp. ( $\diamond$ ), *Listeria* ( $\square$ ); Models: *Pfr*162 (—); *Pfr*K1 (---); *Pf*158 (.....); *Lm*14 (-.-)