EFFECT OF PRESLAUGHTER FEEDING SYSTEM ON GUT BACTERIA IN CATTLE

NG Gregory formerly at MIRINZ, PO Box 617, Hamilton, New Zealand. Now at SARDI, PO Box 1571, SA 5153, Australia LH Jacobson, TA Nagle and GJ LeRoux formerly at MIRINZ, PO Box 617, Hamilton, New Zealand. Now at AgResearch, Private Bag 3123, Hamilton, New Zealand.

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Background

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The New Zealand meat processing industry is putting greater effort into ensuring that meat is *E. coli*-free. There have been two reasons for the move towards zero tolerance for this group of bacteria. Firstly, the presence of *E. coli* in foods of animal origin indicates that there is a moderate to high risk that the food has been contaminated at some stage with faeces, non-potable water or the type and number of *E. coli* that are present. Secondly, certain types of *E. coli*, and in particular *E. coli* 0157:H7, have emerged as organism is not likely to be present in a food, is to control or exclude all *E. coli*, using a zero tolerance policy. Live cattle have been reticulum and omasum (Brown et al. 1997; Chapman et al. 1997).

It is well-recognised that the counts of *E. coli* in facces at the time of slaughter can be highly variable (Davidson and Taylor 1978). For example, in some cattle there is less than 10^2 counts g⁻¹ facces whereas others have more than 10^7 g⁻¹. One reason for this sizeable variation is difference in the time off feed. Fasting before slaughter encourages the growth of *E. coli* in the rumen, and this could lead to more *E. coli* in the facces (Jordan and McEwen 1998). In New Zealand, when cattle are despatched for slaughter, they Little is known about the effect that these preslaughter feeding schedules have on gut microflora.

This study aimed at comparing transport directly off pasture with either feeding hay for two days before transport, or fasting for 2⁴ hours before transport. Comparisons were made in terms of liveweight loss, surface soiling of the skin, gut microflora and digest^a

Methods

Forty five Angus steers were divided into three preslaughter treatments: hay feeding for 48 hours before transport to the processing plant (48h hay), fasting for the last day before transport (Fasted), and allowing to feed on pasture up to the time of transport to the slaughter. They were not fed during this period but had free access to water. Just before slaughter, they were scored for surface plus abomasum, small intestine, caecum and large intestine were tied off to ensure that there was no mixing between these regions of viscera within 1 hour of slaughter. These samples were transferred directly to the microbiology laboratory for estimation of bacteria numbers, and they were also used for dry matter, and physical consistency assessment. Each gut region was weighed separately before and after emptying its digesta. At 24 hours after slaughter, pH and stickiness were measured in the *longissimus dorsi* muscle.

Microbiological measurements made on the digesta and faces included the number of *E. coli, Enterobacteriaceae, Enterococci* and facultative anaerobes. Runniness of the faces or digesta was measured using a splatter test with planimetry. The physical phases of the digesta or faces were assessed using a press test. In this test, the digesta or faces separated into three distinct phases of opaque fluid (sludge ring), and the outermost ring was an almost translucent watery ring (free-water ring).

The effect of three of the preslaughter feeding systems on the counts of *E. coli* g⁻¹ of faeces are shown in Table 1. The corresponding total counts of *E. coli* in the entire gastrointestinal tract are shown in the lower half of Table 1. The fasted group had significantly higher concentrations and total burden of *E. coli* than the pasture-fed and the 48h hay-fed animals (p < 0.001). The total burden of *E. coli* in the large intestine and the counts g⁻¹ of *E. coli* in the faeces were lowest in the 48h hay treatment. The difference in faecal *E. coli* counts g⁻¹ between the fasted and the 48h hay fed treatments was substantial: about 10³.

The number of facultative anaerobes g^{-1} of faces was higher in the fasted animals in comparison with the pasture and 48h hay-fed steers. The total burden of facultative anaerobes in the entire alimentary tract was higher in the fasted steers in comparison with the pasture-fed animals (p < 0.05).

In general, the differences between the treatments in faecal *E. coli* counts were present along the whole length of the gut except for the duodenum. Within the rumen, it appeared that moist conditions and a relatively high pH favoured the growth and survival of *E. coli*, as counts of *E. coli* in the rumen digesta were negatively correlated with rumen digesta dry matter percent (r = -0.75, p < 0.001), and were positively correlated with rumen contents pH (r=0.52, p < 0.001). *Enterococci* and *E. coli* counts g^{-1} were negatively correlated in the rumen (r=-0.47, p < 0.001)

Faeces which were runny, as measured by the splatter test, had a low dry matter (r=-0.82, p<0.001) and a relatively large sludge phase (r=0.86, p<0.001). This association was particularly noticeable for the pasture-fed steers which had a larger sludge phase and of the faeces than the other groups (p<0.05). In many respects, the preslaughter feeding treatment differences in the sludge phase influenced this aspect of digesta consistency throughout the digestive tract. Whereas, this did not apply to runniness. The fasted surface soiling in the pasture-fed steers after overnight holding at the processing plant was 24% whereas in the other treatments it was less than 7% (p<0.05).

An important implication from the findings in this study is that differences in *E. coli* counts between animals in the rumen will probably persist further down the tract. The pattern in the treatment differences in *E. coli* numbers persisted along the gut after the

digesta had passed from the rumen and through the duodenum. This could indicate that the entry rate of E. coli from the rumen into the small intestine was an important factor influencing subsequent E. coli numbers. It implies that manipulating the growth and survival of E. coli in the rumen before transport could be an effective way of influencing the counts of E. coli in the faeces during the critical preslaughter period. Several factors could be influencing the numbers of E. coli in the rumen. A low dry matter material with a near-neutral pH favoured E. coli. Fasting the animals before transport favoured the formation of a high pH-low dry matter ruman environment. In practical terms, the most effective way of manipulating gastro-intestinal counts of E. coli was to feed hay. However, hay feeding did not have much effect on rumen contents pH and dry matter, and so it is suspected that some other factors mediated the E. coli-suppressing effect provided by the hay. These could include the presence of inhibitory compounds that may have been present in the hay, but to a lesser extent in the pasture. Duncan et al. (1998) observed that the coumarins, esculetin, umbelliferone and scopoletin, which are normal constituents in some swards, inhibited the growth of E. coli 0157 in vitro. These components were not monitored in the present study, but the hay that was used did include species such as Anthoxanthum odoratum, Trifolium repens, Trifolium pratense, Polygonum aviculare and Taraxacum officinale which normally contain some of those compounds (Murray et al. 1982). Other possible inhibitors of E. coli, that could have been present in hay but not in fresh pasture, include products from fungi that could have been growing in the hay. An alternative explanation is that the lower counts of E. coli in the hay-fed animals could have been due to greater competition from other bacteria in the gastro-intestinal tract. The hay-fed animals had more Enterococci in their alimentary tracts, and there was a negative relationship between Enterococci and E. coli in the rumen. Some Enterococci are known to inhibit the growth of E. coli, Salmonella, Shigella and Enterobacter species, and have been used in other

^{species} as probiotics because of their beneficial effect on the gut microflora (Franz et al. 1999). The cattle that were pasture-fed up to the time of trucking had more runny faeces. The faeces from the pasture-fed cattle contained a higher proportion of the sludge phase, relative to the fibre and free-water phases. In addition, the pasture-fed animals were dirtier, in terms of fresh faecal soiling on the hide. In summary, feeding pasture on the farm up to the time of transport has only a small effect on the composition of gut contents but it has an adverse affect on faeces consistency and stock cleanliness.

Conclusions. Pasture-fed cattle were carrying approximately 75 billion aerobic bacteria and 86 billion facultative anaerobes in their gastro-intestinal tracts when they were slaughtered. The number and type of bacteria were strongly influenced by the preslaughter feeding system. Fasted animals had less acidic and more moist rumen contents, and they had more *E. coli, Enterobacter* and facultative anaerobes throughout the gut. Cattle fed hay for 48 hours before transport to slaughter had higher numbers of *Enterococci* and fewer *E. coli*. The pasture groups had runnier facees, and were dirtier after overnight holding at the processing plant. It was concluded that feeding cattle hay for 48 hours before despatch for slaughter provided several advantages over other preslaughter feeding systems.

Pertinent Literature

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Table 1. Effect of preslaughter feeding system on bacteria in the faeces and the entire alimentary tract in cattle. No SEDs are given for *E. coli* numbers as non-parametric Kruskal Wallis one-way ANOVA on ranks was used for analysis due to unequal variance of the treatments. Means in a row without a common superscript letter were significantly different at P < 0.05.

	48h Hay	Pasture	Fasted	SED
Counts (logio) g ⁻¹ of fa	eces	dula a bi-ba-holeti		000
Aerobic plate count	6.109 ^a	6.274 ^a	7.358 ^b	0.173
E. coli	3.716 ^a	5.002 ^b	6.650 °	_
Enterobacteriaceae	4.634 ^a	5.163 ^a	6.968 ^b	0.385
Enterococci	5.001	4.600	4.382	0.348
Anaerobes	6.016 ^a	6.410 ^a	7.439 ^b	0.226
lotal burden in the ali	mentary tract, le	0210		
Aerobic plate count	11.889 ^a	10.875 ^b	10.255 ^b	0.111
c. coli	7.609 ^a	8.779 ^a	10.480 ^b	-
enterobacteriaceae	8.142 ^a	9.161 ^a	10.700 ^b	0.288
^c nterococci	9.901	9.383	8.982	0.329
Anaerobes	11.867 ^a	10.932 ^b	11.189 ^b	0.157
Weweight loss before	slaughter, g kg	empty body we	eight	
Ouring transport	22 ^a	23 ^a	12 ^b	2
op to slaughter	77	95	102	8