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The prevalence of Campylobacter and Arcobacter in turkey and poultry meat

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Introduction

Raw poultry meat is known to be an important vehicle in the transmission of Campylobacter enteritis in humans. The most likely mechanisms of transmission are contamination from raw poultry meat to other ready-to-eat foods via the unwashed hands of the cook or via kitchen utensils, direct hand-to-mouth infection after handling of raw poultry, or insufficient cooking of poultry so that the organisms can survive (PARK et al., 1991).

Campylobacter jejuni subsp. jejuni and related thermophilic Campylobacters are common inhabitants of the intestinal tract of poultry and other food animals. BRYAN and DOYLE (1995) reported that the majority of raw poultry products are contaminated with thermophilic Campylobacter spp. and are therefore a potential hazard for humans.

The clinical importance of Arcobacter (A.) spp. is at present limited. Both A. butzleri and A. cryaerophila have been isolated from the faeces of humans with gastroenterits and stomach cramps (KISS and CSORIAN, 1995). Arcobacter spp. are morphologically similar to Campylobacter spp. but differ particularly in being able to grow in air rather than under micro-aerophilic conditions. Unlike the thermophilic Campylobacters (C. jejuni, C. coli, C. lari) Arcobacters are only able to grow at temperatures below 30°C.

Material and Methods

Turkey and poultry samples

Chilled and individually packed turkey carcasses from different slaughterhouses were acquired as was chilled poultry meat from different EU producers and different processing plants. All samples were kept refrigerated. Microbiological culture methods

Atmosphere:

The gas replacement method (BOLTON et al. 1992) was used with anaerobic gas mixture to provide micro-aerobic atmosphere with hydrogen.

Isolation media (ATABAY et al. 1996):

Plating media: mCCDA (Oxoid) plus SR155 was prepared according to manufacturer's instructions. CAT agar (ASPINALL et al. 1993) consisted of mCCDA basal medium (Oxoid) with CAT supplement (cefoperazone-amphotericin-teicoplanin) prepared by adding sterile solutions of the antibiotics (cefoperazone 8 mg/l⁻¹, amphotericin B 10 mg/l⁻¹, teicoplanin 4 mg/l⁻¹). Blood agai comprised 5% (v/v) defibrinated sheep blood in blood agar base No. 2 (Oxoid).

Enrichment broths: CAT broth was prepared using Campylobacter enrichment medium with 5% (v/v) laked horse blood (Oxoid) incorporating the CAT selective supplement described above. Acrobacter enrichment broth (AEB) was prepared as described by LAMMERDING et al. (1996).

In this protocol the isolation method used for poultry and turkey meat was modified to include direct plating of carcass rinse onto mCCDA (10 µl) and onto blood agar using the STEELE and MCDERMOTT membrane filter method.

Other modifications were: (a) omisson of CAT agar at 37°C after enrichment because the plates had been overgrown with competitive flora; (b) omission of Arcobacter enrichment broth followed by mCCDA at 30°C, because it offered no advantage over CAT broth followed by CAT agar (at 30°C) for isolating Arcobacter spp.

Results

380 samples of poultry meat were examined and Campylobacter could be detected in 131 samples. Of the 120 examined turkey samples 25 were found to be infected with Campylobacter. Differentiation of the Campylobacter isolates was done using the api Campy[®] system and computer program (bioMérieux).

Arcobacter could be isolated from 20 samples of turkey meat and from 81 samples of poultry meat. Arcobacter spp. were also differentiated using the api Campy[®] system (bioMérieux) and in addition also according to the scheme of BOLTON et al (1992).

The following Campylobacter strains were detected:

58,5% Campylobacter jejuni biotype 1

8% C. coli

18% C. upsaliensis

16% other Campylobacters (C. sputorum, C. fetus, C. hyointestinalis, C. jejuni var. doylei, C. lari)

The Arcobacter isolates were mainly found in skin samples and could be positively identified as A. cryaerophila and A. butzleri. In addition to that 4% of all examined samples contained Helicobacter cinaedi.

Discussion

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In Germany Campylobacter spp. are the second most important causative organisms of human enteritis. Only salmonellosis is diagnosed more often. However, it is very likely that Campylobacter is only considered to be of less importance than Salmonella because there is no official requirement for examination of samples for Campylobacter as there is for Salmonella. Furthermore cultural methods and differentiation of Campylobacter is far more complicated and is thus carried out less often. Campylobacter cells derived from food or environmental samples may be difficult to cultivate since they might be in a so called "viable but non-culturable" phase. Different methods for the detection of Campylobacter spp. and Arcobacter spp from food are described in literature.

In this study different cultural methods for the detection of Campylobacters and Arcobacters were tested.

Campylobacter spp were found in 34,4% of the examined poultry meat. This percentage seems to be rather low when one considers that in poultry flocks up to 90% of the animals are carriers of these organisms, nevertheless since the meat samples were kept refrigerated all the time this percentage is alarming and shows a potential risk for human infection.

Turkey meat is only rarely examined for contamination with Campylobacter. In 20,8% of the samples in this study Campylobacter could be detected.

Whereas C. jejuni biotype 1 was isolated most often from poultry, C. fetus subsp fetus. was the most common serovar found in turkey meat

The detection of Arcobacter spp., especially A. butzleri, may well be of importance for food hygiene and the food producing industry, since Arcobacter spp. are able to grow at temperatures around 15°C. This means that Arcobacter can multiply in freshly merchantised broilers and thus potentially causes a risk for human health.

References

ASPINALL, S.T., WAREING, D.R.A., HAYWARD, P.G., HUTCHINSON, D.N. (1993). Selective medium for thermophilic Campylobacters including Campylobacter upsaliensis. Journal of Clinical Pathology 45, 829-831.

ATABAY, H.I., CORRY, J.E.L., POST, D.E. (1996). Comparison of the productivity of a variety of selective media for *Campylobacter* and *Arcobacter* species. In: *Campylobacter VIII*. Proceedings of the 8th International Workshop on *Campylobacters*, *Helicobacters* and Related Organisms. ed. Newell, D.G., Ketley, J., Feldman, R.A., pp. 151-161. New York: Plenum Publishing Corporation

BOLTON, F.J., WAREING, D.R.A., SKIRROW, M.B., HUTCHINSON, D.N. (1992). Identification and biotyping of Campylobacters. In: Identification Methods in Applied and Environmental Microbiology; ed. Board, R.G., Jones, D., Skinner, F.A., PP. 151-161. London: Academic Press.

BRYAN, F.L. and DOYLE, M.P. (1995). Health risks and consequences of Salmonella and Campylobacter jejuni in raw poultry. Journal of Food Protection 58, 326-344.

KISS, R. and CSORIAN, E.S. (1995). Isolation of *Campylobacter*, *Helicobacter*, *Arcobacter* species. Food, human and environmental origin - *Campylobacter* surveillance in Hungary. In: *Campylobacter VIII*. Proceedings of the 8th International Workshop on *Campylobacters*, *Helicobacters* and Related Organisms. ed. Newell, D.G., Ketley, J., Feldman, R.A., p.39. New York: Plenum Publishing Corporation.

LAMMERDING, A.M., HARRIS, J.E., LIOR, H., WOODWARD, D.E., COLE, L., MUCKLE, C.A. (1996). Isolation method for recovery of Arcobacter butzleri from fresh poultry and poultry products. In: Campylobacter VIII. Proceedings of the 8th International Workshop on Campylobacters, Helicobacters and Related Organisms. ed. Newell, D.G., Ketley, J., Feldman, R.A., pp. 329-334. New York: Plenum Publishing Corporation.

PARK, R.W.A., GRIFFITHS, P.L., MORENO, G.S. (1991). Sources and survival of Campylobacters: relevance to enteritis and food industry. Journal of Applied Bacteriology 70, 97S-106S.

STEELE, T.W., MCDERMOTT, S.N. (1984). The use of membrane filters applied directly to the surface of agar plates for the isolation of Campylobacter jejuni from facees. Pathology 16, 263-265.