### WATER MOBILITY AND DISTRIBUTION IN PORCINE MUSCLE POST MORTEM AND THE EFFECT OF CHILLING RATE

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### Background

Water-holding capacity (WHC) is a major quality attribute of fresh meat (Andersen, 2000). However, the exact mechanisms determining the WHC of meat are far from understood. Especially, characteristics about proposed water populations in the meat and how they are interrelated with drip loss and how water populations are affected by intrinsic and extrinsic factors need to be studied further. NMR relation measurements provide information about the compartmentalisation and mobility of water in muscle and meat (Hazlewood et al., 1974). Moreover, NMR relation measurements are also shown to be able to determine potential drip loss in meat (Tornberg et al., 1993; Bertram et al., 2001). Consequently, this method seems appropriate to use for further elucidation of water mobility and distribution during the conversion of muscle to meat and how interior and exterior factors affect these.

#### Objective

The objective of the present study was to use Low-Field (LF) NMR relaxation measurements to characterize the proposed water populations in pork and investigate how the distribution and mobility of the water changes *post mortem* as influenced by the halothane gene (Heterozygote carriers (Hal-Nn) vs. wildtype) and chilling regime.

#### Methods

The macro- and microstructures of porcine meat of different technological quality (high to low WHC) were controlled using either chemical modulators (DMSO, Urea, NaCl) or physical treatments (mincing, homogenization, centrifugation). Subsequently, LF-NMR relaxation measurements (23.2 MHz Maran Benchtop Pulsed NMR Analyser, Resonance Instruments, Witney, UK) were carried out on well-characterised meat samples, and compared with microscopic investigations to further reveal the results in relation to physical features of the pork. Moreover, LF-NMR relaxation was continuously measured during the *post mortem* period of 12 porcine muscle samples from DPLY cross-breed slaughter pigs, which were carriers (Hal-Nn) and non-carriers (wildtype) of the halothane gene exposed to two commercially used cooling profiles.

#### **Results and discussion**

LF-NMR relaxation revealed the presence of three distinct water populations (T<sub>2b</sub>, T<sub>21</sub>, T<sub>22</sub>) in meat (Figure 1a), which could be described due to the physical structure of the meat (Figure 1b). T<sub>2b</sub> constitutes water tightly associated with macromolecules,  $T_{21}^{21}$  constitutes water located within highly organized protein structures, e.g. water in tertiary or/and quaternary protein structures and spatials with high myofibrillar protein densities including actin and myosin filament structures, and T<sub>22</sub> constitutes the extra-myofibrillar water containing the sarcoplasmatic protein fraction.





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**Figure 1a.** Distribution of T<sub>2</sub> relaxation times in pork. T<sub>2</sub> data were performed with a  $\tau$ -value (time between 90° pulse and 180° pulse) of 150 µs. Measurements were performed at 25°C, and data were acquired from 4096 echoes as 16 scans repetitions with a repetition time of 2 sec between two succeeding scans.

Figure 1b. Confocal microscopy of pork 24 h post mortem  $(10^{\times} \text{magnification})$  (TCS NT, Leica Microsystems, Wetzlar, Germany). Before examination the sample was covered with coverslips whereupon Nile red dissolved in acetone was settled, and the acetone was evaporated before mounting.

<sup>C</sup>omparison of WHC, determined by Honikel's method (Honikel, 1998), and the water populations determined by NMR revealed high <sup>c</sup>orrelation between the T<sub>22</sub> population and WHC (Figure 3).

Moreover, changes in water mobility and distribution were followed *post mortem* using continuous LF-NMR measurements. Both chilling <sup>re</sup>gime and genotype significantly affected changes in the characteristics (rate constants and populations) of T<sub>2b</sub>, T<sub>21</sub> and T<sub>22</sub>, and hereby the WHC of the meat. Figure 4 displays the found changes in the populations of T<sub>22</sub> (extramyofibrillar water) by time.



Figure 3. Relationship between T22 population and drip loss, determined by Honikel's method (Honikel, 1998).



# Conclusions

The present study reveals that drip loss originates mainly from the extra-myofibrillar water population containing the <sup>sarcoplasmatic</sup> protein fraction. Moreover during the conversion of muscle to meat, the intrinsic water distribution is heavily affected <sup>by</sup> genotype and chilling regime through their effect on membrane stability.

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