



QUALITY OF MEAT FROM STEERS OF TWO DIFFERENT FRAME SIZES GRAZING HIGH QUALITY PASTURES SUPPLEMENTED WITH HIGH MOISTURE MAIZE GRAIN OR WHOLE PLANT MAIZE SILAGE

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Background

Beef produced from grazing cattle results in higher levels of unsaturated fatty acids (Miller, *et al.*, 1987 and García and Casal, 1992), lower n-6:n-3 fatty acids (Enser, *et al.* 2001) and higher levels of conjugated linoleic acid (CLA) in body fat (French, *et al.* 2000) as opposed to meat from animals fattened in high concentrate diets. However, short term supplementation with maize grain or soybean harvest by-products in the finishing stage did not affect fatty acid profile (Grigera Naón, *et al.*, 2000 and Grigera Naón, *et al.*, 2003).; longer supplementation periods may be necessary to overcome seasonal grass production, thus the widespread use of other supplements such as high moisture maize grain and maize silage (Abdelhadi, 2000), which in turn may affect meat quality. Frame size of animals can have decisive bearings on the length of the fattening stage (Di Marco, 1998) and on some aspects of quality such as tenderness and cooking loss (Muir *et al.*, 1998 and Camfield *et al.* 1999).

Objectives

The objective was to evaluate the effect of type of energy fall-winter supplementation with high moisture maize grain or whole plant maize silage on meat quality of Aberdeen Angus steers with contrasting mature body weight.

Materials and methods

Over 177 days, covering autumn and winter, 32 male calves of six months of age, grazing pastures were assigned to the following treatments: LM, animals of low (L) mature body weight supplemented with high moisture maize grain (M); LS, animals of low (L) mature body weight supplemented with whole plant maize silage (S); HM, high (H) mature body weight animals supplemented with high moisture maize grain (M) and HS, high (H) mature body weight animals supplemented with whole plant maize silage (S). Individuals were weighed every 21 days and fat depth between ribs 12th and 13th was measured by ultrasound using a 3.5 MHz transducer. At the end of the experiment, eight animals from each treatment were slaughtered. Colour was assessed on the *Longissimus* muscle exposed between the 12th and 13th rib, blooming time was 60 minutes (Wulf and Wise, 1999), readings were taken in L* a* b* colour space, using a Minolta CR-300 (Minolta Co. Ltd., Japan) colorimeter. A Testo 230 pH-meter with a puncture type combination electrode (Testo GmbH & Co., Germany) was used to measure muscle ultimate pH (pHu). Fatty acids were extracted according to Folch *et al.* (1957) and analyzed as methyl esters by gas chromatography. Tenderness was measured with an Instron 4442 Universal Testing Machine (Canton, MA, USA) with a Warner-Bratzler shearing attachment on samples cooked in a water bath at 70 °C for 50 minutes. Data were analyzed using GLM procedure SAS (SAS Inst. Inc., Cary N.C.).

Results and discussion

Animal performance data is shown in Table 1, H calves had higher (P < 0.01) initial liveweight as expected. Initial fat depth was the same across treatments. At the end of the trial L steers had a deeper (P < 0.01)



subcutaneous fat layer, lower liveweight gain ($P < 0.01$) and were lighter ($P < 0.01$) than H reflecting that smaller animals were more mature. Steers on M gained more ($P < 0.01$) in autumn than those supplemented with S.

Among treatments, the proportion of ether extract (EE), saturated fatty acids (SFA), polyunsaturated fatty acids (PUFA), n-6 fatty acids, n-3 fatty acids and the ratio between the latter two (Table 2) in meat was similar ($P > 0.1$). The n-6: n:3 ratio was below 4:1, therefore considered healthy for human beings (Holman, 1995). Concentration of monounsaturated fatty acids (MUFA) was higher in L steers, in accordance to their higher degree of fatness. Animal size influenced the proportion of conjugated linoleic acid (CLA) for LS and LM was 1.08 g/100 g and 0.82 g/100 g respectively ($P < 0.05$), whereas for HS was 0.93 g/100 g and 1.06 g/100g for HM ($P > 0.10$). In smaller animals the effect of the different feeding regime became apparent, the rich forage diet enhanced CLA body fat levels as reported elsewhere (French *et al.*, 2000 and Grigera Naón *et al.*, 2003). In case of larger steers this difference was not detected, which can be associated with the fact that they were thinner animals. Ultimate pH (Table 3) was higher for H ($P < 0.05$) than for L and similar for both supplements. There was a significant ($P < 0.05$) interaction between animal size and diet for a^* and b^* , LM showed higher values in comparison to LS, both parameters were similar for large frame steers. Grigera Naón *et al.* (2001), reported that a^* and b^* were affected when maize grain was fed to grazing steers. Tenderness was not affected neither by type of animal nor by the supplement fed. However, there was a trend for those animals on S to produce somewhat more tender meat. Cooking loss was higher in H as compared to L ($P < 0.05$), Camfield *et al.* (1999), reported comparable results from three muscles of steers differing in frame size. Such higher cooking loss was recorded in spite of higher pHu in H ($P < 0.05$), but such value (pHu = 5.8) can seldom affect juiciness (Lawrie, 1998). Cooking losses were similar ($P > 0.1$), when comparisons were made between supplements, Geay *et al.* (2000) consider that juiciness cannot be modified by feeding regimes.

Conclusions

During the supplementation period, meat from large animals showed lower MUFA and higher pHu than meat from smaller animals; supplements had some effect on CLA concentration and colour parameters a^* and b^* in small frame steers.

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Table 1. Animal performance

Variable	Factors				se	Contrasts, <i>P</i> <		
	H	L	M	S		H vs.L	M vs. S	B*D
Initial liveweight, kg	185	153	171	166	6.4	<0.01	0.47	0.21
Final liveweight, kg	296.1	260.1	283.9	272.3	7.8	<0.01	0.15	0.23
Daily gain , g/d								
-Autumn	356	426	457	325	43	0.10	<0.01	0.64
-Winter	1103	924	1016	1011	36	<0.01	0.86	0.88
Initial fat depth, mm	2,2	2,2	2,2	2,2	0,11	0,71	0,97	0,21
Final fat depth, mm	2.88	4.46	4.01	3.34	0.47	<0.01	0.03	0.34



Table 2. Ether extract and fatty acid composition (%) of muscle lipids

Variable	Factors				se	Contrasts, <i>P</i> <		
	H	L	M	S		H vs.L	M vs. S	B*D
CLA	0.99	0.95	0.94	1.00	0.08	0.55	0.38	0.02
EE, %*	8.12	8.70	8.02	8.79	1.08	0.58	0.46	0.25
SFA	50.0	45.9	48.0	47.9	2.41	0.07	0.98	0.98
MUFA	41.6	47.9	43.7	44.7	1.59	<0.01	0.56	0.30
PUFA	8.34	7.34	8.27	7.41	1.18	0.34	0.41	0.13
n-3	1.96	1.62	2.03	1.55	0.40	0.34	0.18	0.06
n-6	5.31	4.63	5.19	4.76	0.36	0.52	0.58	0.12
Ratio n-6:n-3	3.36	3.27	2.99	3.64	0.52	0.85	0.21	0.39

* on a dry matter basis

Table 3. Cooking loss, pH, colour and tenderness

Variable	Factors				se	Contrasts, <i>P</i> <		
	H	L	M	S		H vs.L	M vs. S	B*D
Cooking loss, g/g	0.17	0.14	0.16	0.14	0.01	0.04	0.12	0.43
pH	5.80	5.64	5.66	5.77	0.08	0.04	0.14	0.15
a*	18.6	20.2	20.3	18.5	1.55	0.27	0.22	0.04
b*	6.65	7.00	7.27	6.39	0.55	0.53	0.13	0.03
L	36.0	36.6	36.8	35.8	1.1	0.55	0.36	0.20
Shear force, kg	6.79	6.65	7.30	6.14	0.59	0.82	0.07	0.58