

EFFECTS OF ELECTRICAL STIMULATION ON μ -CALPAIN AND CATHEPSIN B+L ACTIVITIES IN POSTMORTEM BOVINE *LONGISSIMUS* MUSCLE

E. Veiseth^{*1}, K.I. Hildrum¹, L. Aass², and K. Hollung¹

¹Matforsk AS, Osloveien 1, N-1430 Ås, Norway; eva.veiseth@matforsk.no

²Department of Animal and Aquacultural Sciences, Norwegian University of Life Sciences, P.O. Box 5003, N-1432 Ås, Norway

Key Words: beef, calpain, cathepsin B+L, electrical stimulation, Warner-Bratzler shear force

Introduction

Consumers are aware of variability in beef product quality. According to one US study, 50% of consumers rated tenderness to be the single most important attribute of beef eating quality (Miller et al., 1995). High-voltage electrical stimulation (HV-ES) has been found to improve beef tenderness (Elgasim et al., 1981; Marsh et al., 1987), while the effect of low-voltage electrical stimulation (LV-ES) seems to be more variable (Hildrum et al., 1999). Despite concerns over its effectiveness, LV-ES of carcasses is in widespread use to enhance meat tenderness.

In a recent study we found a significant positive effect of LV-ES on Warner-Bratzler shear force (WBSF) in bovine *longissimus* (LD) muscle at 2 and 8 days *postmortem*, however large variation was observed in individual animals with regard to pH decline and WBSF (Hollung et al., 2007). This variation was not always correlated for the two responses; specifically some animals showed no response of LV-ES on pH decline but still displayed improved WBSF, and visa versa. Collectively, these results indicate that mechanisms other than accelerated pH decline and prevention of cold-shortening are affecting meat tenderness. Using samples from the Hollung study (Hollung et al., 2007), we sought to investigate the effects of LV-ES on proteases (μ -calpain and cathepsin B+L) in order to reveal whether altered activity of these enzymes could be a mechanism through which LV-ES improves meat tenderness.

Materials and methods

The ten Norwegian Red bulls (12-14 months) used in this study were slaughtered and treated as describe by Hollung et al. (2007). At 1, 3, 6, 10, and 24 h post-ES, samples for μ -calpain analysis were frozen in liquid nitrogen and stored at -80°C for later analysis. Casein zymography (Raser et al. 1995) was used for analysis of μ -calpain activity. Regions of transparent gel (indicating μ -calpain activity) were measured (Quantity One, Version 4.5.0, Bio-Rad Laboratories, Inc., Hercules, CA), and quantities were expressed as a percentage of the 1-h sample activity measured in the NES side of each animal. Analyses of cathepsin B+L activities were performed on fresh samples at 2 and 8 days *postmortem* from 4 animals. Extractions and subcellular fractionation was performed according to Erbjerg et al. (1999). Fractions were stored at -80°C until measured using the fluorometric procedure described by Kirschke et al. (1983). Total activity of cathepsin B+L was calculated as the sum of the activity in the different subcellular fractions.

Analysis of variance in μ -calpain and cathepsin B+L data was performed using MINITAB's general linear model (Minitab, version 14.2). The model for cathepsin B+L included the following factors: animal (fixed), level of ES treatment, ageing period, and all two- and three-factor interactions. The model for μ -calpain included the following factors: animal (fixed), level of ES treatment, time post-ES, and the interaction of ES and time post-ES.

Results and discussion

We observed significant effects of LV-ES on μ -calpain activity at 1 and 24 h post-ES (Figure 1A). Taken together, the increased μ -calpain activity at 1 h and reduced activity at 24 h post-ES indicate accelerated μ -calpain activity and autolysis. It is likely that these changes would lead to an accelerated and increased *postmortem* proteolysis of myofibrillar and cytoskeletal proteins, and thus improved meat tenderness. Indeed, several groups have previously found that ES can accelerate *postmortem* proteolysis through changes in the calpain system (Ferguson et al., 2000; Geesink et al., 2001).

Release of cathepsins from the lysosomes has previously been suggested to accelerate proteolysis in muscle (Dutson et al., 1980; Pommier, 1992). No significant changes in cathepsin B+L activities resulted from the LV-ES treatment in this study (Figure 1B), making it less likely that these proteases play a crucial role in the observed improvement of meat tenderness. However, there was a strong tendency ($P = 0.052$) towards an increase in their activity in the soluble fraction as a result of LV-ES, suggesting that their influence cannot be excluded altogether.

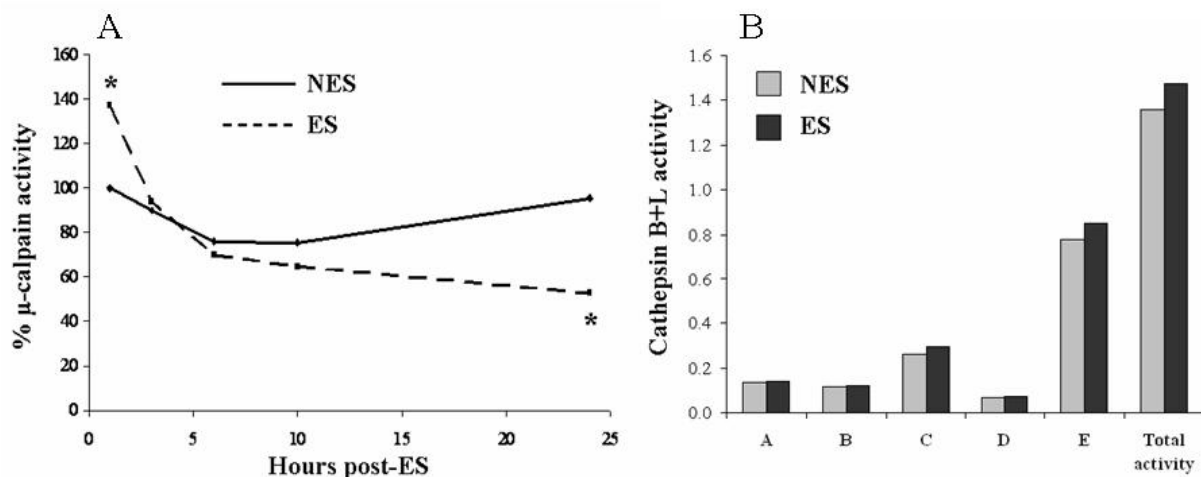


Figure 1. Protease activities in bovine LD muscle during *postmortem* storage. A. Activity of μ -calpain over time as a percentage of 1h-NES activity (n=10). B. Cathepsin B+L activities in different subcellular fractions (A=myofibrillar, B=heavy mitochondrial, C=lysosomal, D=microsomal, E=soluble) and total activity (n=4). *Significant ($P < 0.05$) differences between NES and ES

Conclusions

Hollung (et al. 2007) found a significant effect of LV-ES on pH at 1, 3, 6 and 10 h post-ES, but not at 24 and 48 h. Moreover, a significant positive effect of LV-ES on WBSF at both 2 and 8 days of ageing was observed. Since sarcomere lengths were unaltered following LV-ES, the reduced WBSF was not caused by a prevention of cold-shortening and must result from a different mechanism. The results of this study indicate that this mechanism is most likely increased μ -calpain activity. The accelerated rate of μ -calpain autolysis in the LV-ES samples indicates that the tenderisation process in these samples would be accelerated, and can explain the reduced WBSF seen as early as 2 days *postmortem*.

References

- Dutson, T.R., Smith, G.C., and Carpenter, Z.L. (1980), Lysosomal enzyme distribution in electrically stimulated ovine muscle, *Journal of Food Science*, 45, 1097-1098.
- Elgasim, E.A., Kennick, W.H., McGill, L.A., Rock, D.F., and Soeldner, A. (1981), Effects of electrical-stimulation and delayed chilling of beef carcasses on carcass and meat characteristics, *Journal of Food Science* 46, 340-343.
- Ertbjerg, P., Larsen, L.M., and Møller, A.J. (1999), Effect of prerigor lactic acid treatment on lysosomal enzyme release in bovine muscle, *Journal of the Science of Food and Agriculture*, 79, 95-100.
- Ferguson, D.M., Jiang, S.T., Hearnshaw, H., Rymill, S.R., and Thompson, J.M. (2000), Effect of electrical stimulation on protease activity and tenderness of M-longissimus from cattle with different proportions of *Bos indicus* content. *Meat Science* 55, 265-272.
- Geesink, G.H., Mareko, M.H.D., Morton, J.D., and Bickerstaffe, R. (2001), Electrical stimulation - when more is less. *Meat Science* 57, 145-151.
- Hildrum, K.I., Solvang, M., Nilsen, B.N., Frøystein, T., and Berg, J. (1999). Combined effects of chilling rate, low voltage electrical stimulation and freezing on sensory properties of bovine M-longissimus dorsi. *Meat Science* 52, 1-7.
- Hollung, K., Veiseth, E., Frøystein, T., Aass, L., Langsrud, Ø., Hildrum, K.I. (2007), Variation in the response to manipulation of post mortem glycolysis in beef muscles by low-voltage electrical stimulation and conditioning temperature, *Meat Science*, in press.
- Kirschke, H., Wood, L., Roisen, F.J., and Bird, J.W.C. (1983), Activity of lysosomal cysteine proteinase during differentiation of rat skeletal muscle, *Biochemical Journal*, 214, 871-877.
- Marsh, B.B., Ringkob, T.P., Russell, R.L., Swartz, D.R., and Pagel, L.A. (1987), Effects of early-postmortem glycolytic rate on beef tenderness. *Meat Science* 21, 241-248.
- Miller, M.F., Huffman, K.L., Gilbert, S.Y., Hamman, L.L., and Ramsey, C.B. (1995), Retail consumer acceptance of beef tenderized with calcium chloride. *Journal of Animal Science* 73, 2308-2314.
- Pommier, S.A. (1992), Vitamin-A, electrical-stimulation, and chilling rate effects on lysosomal-enzyme activity in aging bovine muscle. *Journal of Food Science* 57, 30-35.
- Raser, K. J., A. Posner, and K. K. W. Wang. 1995. Casein zymography: A method to study μ -calpain, m-calpain, and their inhibitory agents. *Archives of Biochemistry and Biophysics*, 319, 211-216.