# Effects of *Rhus verniciflua* Stokes Extract, Gallic acid, and Fisetin on the Lipid, Protein, and Myoglobin Oxidation in Hanwoo (Korean Cattle) Beef Model System

Sun Moon Kang<sup>1</sup>, Soohyun Cho<sup>1</sup>, Donghun Kim<sup>1</sup>, and Sung Ki Lee<sup>2,\*</sup>

<sup>1</sup> National Institute of Animal Science, Rural Development Administration, Suwon 441-706, Korea <sup>2</sup> Department of Animal Products of Food Science, Kangwon National University, Chuncheon 200-701, Korea

Abstract— This research was conducted to investigate the effects of RVS water extract (15 ppm), gallic acid (50  $\mu$ M), and fisetin (50  $\mu$ M) on the lipid, protein, and myoglobin oxidation in Hanwoo (Korean cattle) beef homogenate system. The experimental homogenates (M. longissimus dorsi) were incubated under oxidized condition by Fe (III)/ascorbic acid at 37°C for 7 hr. Trolox equivalent antioxidant capacity (TEAC), using ABTS<sup>+</sup> radical scavenging activity assay, and ferric reducing antioxidant power (FRAP) were increased significantly (P < 0.05) by all additions. Gallic acid and fisetin resulted in the highest (P < 0.05) FRAP and the highest (P < 0.05) TEAC, respectively. The inhibition of TBARS level was obtained by all antioxidants and was in following the order: fisetin > RVS extract > gallic acid. Carbonyl content was inhibited by RVS extract and fisetin. Fisetin was more effective than RVS extract for the inhibition of protein oxidation. Myoglobin oxidation was not inhibited by all antioxidants. Furthermore, gallic acid and fisetin accumulated the metmyoglobin formation. These results suggest that RVS extract is more advantageous than gallic acid and fisetin for improving the storage stability of beef products.

Keywords— Rhus verniciflua Stokes extract, gallic acid, fisetin, model system, Hanwoo beef.

# I. INTRODUCTION

From about 4,000 years ago, *Rhus verniciflua* Stokes (RVS) belonging to the *ivy* (Anacardiaceae) has been used for medicine in South Korea, China, and Japan [1]. The extract of RVS contains a variety of polyphenol compounds (butein, butin, gallic acid, fisetin, fustin, sulfuretin, and quercetin etc.) [2, 3, 4] and has beneficial effects, such as antioxidant, anticancer, antimutagenic, anti-inflammatory, antithrombotic, and anti-obecity [5, 6, 7]. Fisetin (3,7',3',4-tetrahydroxyflavone; 5-deoxyquercetin) and gallic acid (3,4,5-trihydroxybenzoic acid) are rich in various plants as well as in RVS [8, 9]. These

compounds have a powerful free radical scavenging activity and ferric iron chelating ability [10, 11], also pharmacological effects, such as anticancer and antiinflammatory [12, 13, 14, 15].

The antioxidant effect of RVS extract in liposome model system and meat has been reported by Liang et al. [16]. However, there is still little information on effect of extract and compounds of RVS on the radical scavenging activity, ferric iron reducing ability, and chemical oxidation in meat system. Therefore, this research was conducted to investigate the effects of RVS extract, gallic acid, and fisetin on the antioxidant status in beef homogenate model system oxidized by Fe (III) and ascorbate.

#### **II. MATERIALS AND METHODS**

#### A. Preparation of RVS extract

Thirty grams of RVS heartwood powder and 500 mL of deionized water were boiled at  $100^{\circ}$ C for 3 hr. After cooled in the room temperature, RVS extract was filterd with a Whatman filter paper No. 1 and a 0.45 µm syringe filter. The soluble matter content of RVS extract was 0.5% with a refractometer (PAL-03S, Atago Co., Ltd., Japan).

#### B. Preparation of meat homogenate model system

Fresh *M. longissimus dorsi* from 28 months-old-Hanwoo (Korean cattle) steer and 9 volumes of 0.12 M KCl-5 mM histidine buffer (pH 7.0) were homogenized at 15,000 rpm for 2 min. Meat homogenate was filtered with a cheese cloth, centrifuged at 2°C, 600 g for 10 min, and then incubated at 37°C under oxidized condition by 30  $\mu$ M FeCl<sub>3</sub> and 100  $\mu$ M sodium ascorbate (Final concentration). Before oxidation, deionized water (control), 50  $\mu$ M gallic acid, 50  $\mu$ M fisetin, and 15 ppm

2

RVS extract (Final concentration) in model system. At 0, 3, and 7 hr of incubation, meat homogenates were mixed with 0.02% BHT [Final concentration; 17] for stopping the oxidation and stored at  $-80^{\circ}$ C until measurement.

# *C. Trolox equivalent antioxidant capacity and ferric reducing antioxidant power*

Trolox equivalent antioxidant capacity (TEAC) was determined by ABTS<sup>+</sup> radical scavenging activity assay of Re et al. [18] and calculated as  $\mu$ M trolox using inhibition rate (%) of ABTS<sup>+</sup> radical and trolox (0~20  $\mu$ M) standard curve. Ferric reducing antioxidant power (FRAP) was performed by Benzie and Strain [19] and calculated as  $\mu$ M Fe (II) using standard (1000  $\mu$ M ascorbate) and curve of 100~1000  $\mu$ M FeSO<sub>4</sub>·7H<sub>2</sub>O.

#### D. TBARS and DNPH-carbonyl

TBARS (2-thiobarbituric acid reactive substances) was performed by described as Siu and Draper [20] and expressed as ng malondialdehyde (MDA) per mg protein. Protein concentration was performed by biuret method [21]. DNPH-carbonyl was determined by Oliver et al. [22] and calculated as nmol DNPH per mg protein using millimolar extinction coefficient of protein hydrazones [21.0 mM<sup>-1</sup>cm<sup>-1</sup>; 23] and BSA standard curve.

#### E. Oxymyoglobin and metmyoglobin concentrations

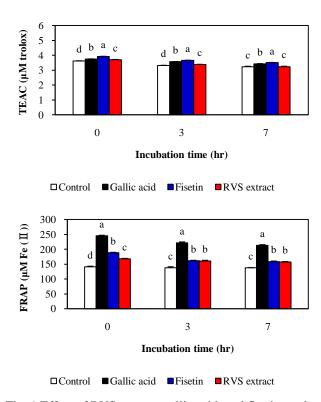
Oxymyoglobin (OxyMb) and metmyoglobin (MetMb) concentrations were performed by described as Krzywicki [24] and calculated as relative percentage (%) of myoglobin derivatives (OxyMb + MetMb + DeoxyMb).

## F. Statistical analysis

Data was analyzed by ANOVA (Analysis of variance) of SPSS [25]. Significant differences among means were determined by the Duncan's multiple range tests at P < 0.05.

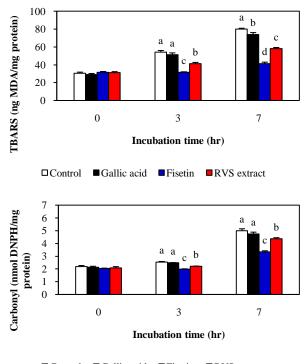
### **III. RESULTS AND DISCUSSION**

Fig. 1 shows the effect of RVS extract, gallic acid, and fisetin on the TEAC and FRAP in Hanwoo beef homogenate model system. Both TEAC and FRAP were significantly (P < 0.05) increased by all additives. Particularly, gallic acid- and fisetin-added beef homogenates had the highest (P < 0.05) TEAC and highest (P < 0.05) FRAP, respectively. But RVS extract-added beef homogenate showed the lowest (P < 0.05) effects in both TEAC and FRAP. Fisetin and gallic acid have antioxidant effects on free radicals and ferric iron [10, 11]. As well, fisetin is one of quercetin derivatives as 5-deoxyquercetin. According to Rice-Evans et al. [10], quercetins have higher radical scavenging activity compared with gallic acid.

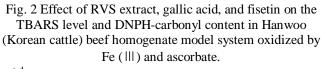


- Fig. 1 Effect of RVS extract, gallic acid, and fisetin on the TEAC and FRAP in Hanwoo (Korean cattle) beef homogenate model system oxidized by Fe (III) and ascorbate.
- <sup>a-d</sup>Means $\pm$ S.E. with different letters indicate significant differences between treatments at P < 0.05.

As shown in Fig. 2, TBARS level in beef homogenate was retarded by addition of all additives. Fisetin-added beef homogenate had the strongest antioxidant effect on TBARS development but gallic acid-added beef homogenate showed the lowest effect. DNPH-carbonyl content (Fig. 2) was inhibited by fisetin. Fisetin-added RVS extract and beef homogenate had the highest antioxidant effect on carbonyl accumulation. In experiments of liposome and beef homogenate model systems of Liang et al. [16], similar results, RVS extract strongly inhibited lipid oxidation, have been reported.

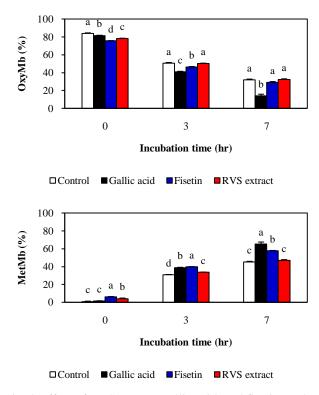






<sup>a-d</sup>Means $\pm$ S.E. with different letters indicate significant differences between treatments at P < 0.05.

Fig. 3 indicates the effect of RVS extract, fisetin, and gallic acid on the myoglobin oxidation in beef homogenate. The generation rate of MetMb was faster in RVS extract-, fisetin-, and gallic acid-added beef homogenates compared with the control. Thus, all additives decreased myoglobin oxidation stability in beef homogenate. Hayes et al. [26] also have suggested that sesamol, one of polyphenol compounds, promoted myoglobin oxidation in meat homogenate model system.



- Fig. 3 Effect of RVS extract, gallic acid, and fisetin on the myoglobin oxidation in Hanwoo (Korean cattle) beef homogenate model system oxidized by Fe (III) and ascorbate.
  <sup>a-d</sup>Means±S.E. with different letters indicate significant
  - differences between treatments at P < 0.05.

#### **IV. CONCLUSIONS**

Although addition of RVS extract, gallic acid, and fisetin increased total antioxidant activity in beef homogenate, gallic acid had small effect on lipid and protein oxidation. Moreover, all additives did not have antioxidant effect on myoglobin oxidation.

#### ACKNOWLEDGMENT

This work was carried out with the support of "Cooperative Research Program for Agriculture Science & Technology Development (Project No. PJ006218201002)" Rural Development Administration, Republic of Korea.

#### REFERENCES

- Kim M J, Hyun J O (1997) Genetic variation in urushiol components of *Rhus verniciflua* Stokes. Korean J Breed 29: 115-123
- Kim J S, Kwon Y S, Chun W J et al. (2010) *Rhus verniciflua* Stokes flavonoid extracts have anti-oxidant, anti-microbial and  $\alpha$ -glucosidase inhibitory effect. Food Chem 120:539-543
- Lee J D, Huh J E, Jeon G S et al. (2009) Flavonol-rich RVHxR from *Rhus verniciflua* Stokes and its major compound fisetin inhibits inflammation-related cytokines and angiogenic factor in rheumatoid arthritic fibroblast-like synovial cells and in vivo models. Int Immunopharm 9:268-276
- Son Y O, Lee K Y, Lee J C et al. (2005) Selective antiproliferative and apoptotic effects of flavonoids purified from *Rhus verniciflua* Stokes on normal versus transformed hepatic cell lines. Toxicol Let 155:115-125
- Jeon W K, Lee J H, Kim H K et al. (2006) Anti-platelet effects of bioactive compounds isolated from the bark of *Rhus verniciflua* Stokes. J Ethnopharm 106:62-69
- Lee D S, Jeong G S, Li B et al. (2010) Anti-inflammatory effects of sulfuretin from *Rhus verniciflua* Stokes via the induction of heme oxygenase-1 expression in murine macrophages. Int Immunopharm 10:850-858
- Park Y K, Jung G O, Lee K T et al. (2004) Antimutagenicactivity of flavonoids from the heartwood of *Rhus verniciflua*. J Ethnopharm 90:73-79
- Zheng L T, Ock J, Kwon B M et al. (2008) Suppressive effects of flavonoid fisetin on lipopolysaccharide-induced microglial activation and neurotoxicity. Int Immunopharm 8:484-494.
- Monach C, Scallbert A, Morand C et al. (2004) Polyphenol: food sources and bioavailability. Am J Clin Nutr 79:727-747
- Rice-Evans C A, Miller N J, Paganga G (1996) Structureantioxidant activity relationships of flavonoids and phenolic acids. Free Rad Biol & Med 20:933-956
- Jadon A, Bhadauria M, Shukla S (2007) Protective effect of Termanalia belerica Roxb. and gallic acid against carbon tetrachloride induced damage in albino rats. J Ethnopharmacol 109:214-218

- Faried A, Kurnia D, Faried L S et al. (2007) Anticancer effects of gallic acid isolated from Indonesian herbal medicine, Phaleria macrocarpa (Scheff.) Boerl, on human cancer cell lines. Int Oncol 30:605-613
- Prince P S M, Priscilla H, Devika P T (2009) Gallic acid prevents lysosomal damage in isoproterenol induced cardiotoxicity in Wistar rats. Eur J Pharmacol 139:139-143
- Sung B, Pandey M K, Aggarwal B B (2007) Fisetin, an inhibitor of cyclin-dependent kinase 6, down-regulates nuclear factorkappaB-regulated cell proliferation, antiapoptotic and metastatic gene products through the suppression of TAK-1 and receptor-interacting protein-regulated Ikappa Balpha kinase activation. Mol Pharmacol 71:1703-1714
- Liang C Y, Kang S M, Kim Y S et al. (2005) Antioxidant activity of *Rhus verniciflua* Stokes extract in model systems and cooked beef. Korean J Food Sci Ani Resour 25:189-195
- Mercier Y, Gatellier P, Renerre M (2004) Lipid and protein oxidation in vitro, and antioxidant potential in meat from Charolais cows finished on pasture or mixed diet. Meat Sci 66: 467-473
- Re R, Pellegrini N, Proteggente A et al. (1999) Antioxidant activity applying an improved ABTS radical cation decolorization assay. Free Radic Biol Med 26:1231-1237
- Benzie I F F, Strain J J (1999) Ferric reducing/antioxidant power assay: direct measure of total antioxidant activity of biological fluids and modified version for simultaneous measurement of total antioxidant power and ascorbic acid concentration. In: Methods in enzymology. Packer, L. (ed), vol. 299, Academic Press Inc., San Diego, CA, USA, pp 15-27
- Siu G M, Draper H H (1978) A survey of the malonaldehyde content of retail meats and fish. J. Food Sci 43:1147-1149
- Gornall A G, Bardawill C J, David M M (1948) Determination of serum protein by means of the biuret reaction. J Biol Chem 177:751-766
- Oliver C N, Ahn B W, Moerman E J et al. (1987) Age-related changes in oxidized proteins. J Biol Chem 262:5488-5491
- Jones L A, Holmes J C, Seligman R B (1956) Spectrophotometric studies of some 2,4-dinitrophenylhydrazones. Anal Chem 28: 191-198
- Krzywicki K (1982) The determination of haem pigments in meat. Meat Sci 7:29-36
- SPSS (2009) PASW Statistics 18, SPSS Inc., Illinois, USA
- Hayes J E, Stepanyan V, Allen P et al. (2009) The effect of lutein, sesamol, ellagic acid and olive leaf extract on lipid oxidation and oxymyoglobin oxidation in bovine and porcine muscle model systems. Meat Sci 83:201-208