

PROTEIN IDENTIFICATION AND *IN VITRO* DIGESTION OF FRACTIONS FROM *TENEBRIO MOLITOR*

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Abstract – Although amino acid composition of *Tenebrio molitor* has been studied before, limited knowledge is available, on which bulk proteins it consists of, and on its digestibility, being a determinant of the nutritional value of protein. The objective of this study was to investigate *in vitro* protein digestibility of whole *Tenebrio molitor* larvae, a water-soluble fraction (supernatant), and water-insoluble fractions (pellet and residue), and to identify which proteins were present in the fractions studied. The digestibility of the supernatant fraction (~80%) was much higher than that of pellet (~50%) and residue (~24%) after *in vitro* gastro-duodenal digestion as was determined using the o-phthalaldehyde (OPA) method. More proteins were digested after pepsin/pancreatin digestion than after only pepsin digestion. The most abundant proteins in the supernatant were hemolymph protein (~12 kDa) & putative allergens (e.g. alpha-amylase ~ 50 kDa), and muscle proteins (e.g. actin 30-50 kDa) in the pellet fraction as determined from LC-MS/MS and SDS-PAGE. In conclusion, the proteins in the soluble fraction that contained hemolymph proteins were more easily digestible than the insoluble, muscle protein containing fractions.

Key Words – Insect protein; *Tenebrio molitor*; *in vitro* digestion; Protein identification; LC-MS/MS

INTRODUCTION

Insects can be considered as an alternative protein source with less environmental impact [1]. [2] reported the protein content of *Tenebrio molitor* was comparable to that of conventional meat products, and the sum of essential amino acids of *T. molitor* was higher than that for the daily protein requirement.

The nutritional value of a food protein is evaluated not only in amino acid composition, but also in protein digestibility. No information from a food science point of view is available on protein

digestibility of *T. molitor* proteins. The objective of this experiment was to investigate protein digestibility (*in vitro*) of the whole insects and insect fractions (supernatant, pellet and residue) obtained from an aqueous extraction designed by [2].

I. MATERIALS AND METHODS

Tenebrio molitor larvae were purchased from a commercial supplier (Kreca V.O.F, Ermelo, the Netherlands).

In vitro gastric digestion with increasing digestion time was performed followed by *in vitro* duodenal digestion, based on [3]. The obtained digested fractions were characterized in terms of molecular weight (SDS-PAGE) and protein content (Dumas). Free amino acids of all digested fractions were determined using the o-phthalaldehyde (OPA) method.

Filter-aided sample preparation (FASP) was used to prepare protein samples from the three protein fractions obtained as described by [4] with some modifications. Peptide measurements were performed by nanoLC-LTQ-Orbitrap XL-MS/MS (Thermo electron, San Jose, CA, USA) as described by [4].

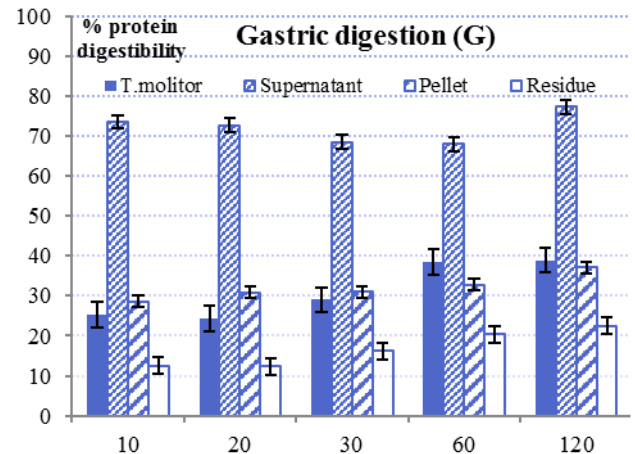
II. RESULTS AND DISCUSSION

Table 1. : Identified muscle proteins of defatted & ground whole *T. molitor*, supernatant and pellet fractions (UniProt: taxonomy 50557, *Insecta*) by LC-MS/MS; putative uncharacterized proteins identified based on family and domain databases from UniProt. Mol. Weight = molecular weight as calculated from the amino acid sequence. Log 10

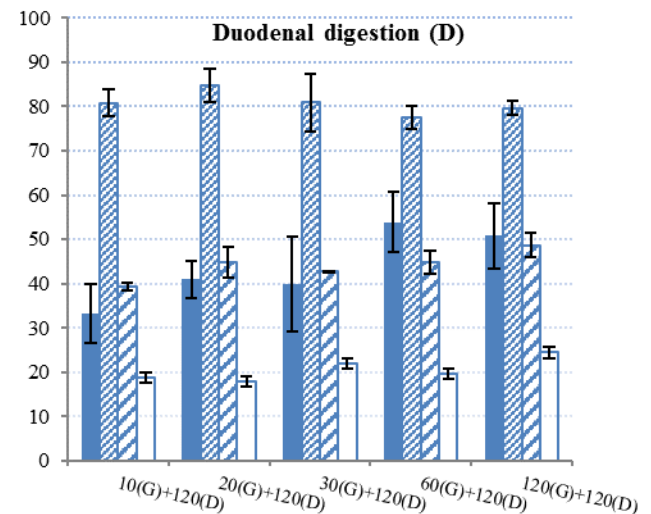
(iBAQ; intensity based absolute quantitation) was used for measuring protein intensity.

Muscle proteins	Mol. weight [kDa]	Log10 (iBAQ defatted <i>T. molitor</i>)	Log10 (iBAQ Pellet)	Log10 (iBAQ Supernatant)
1 Alpha-actinin-4	107	5.7	5.8	5.9
2 Actin like	42	6.4	7.2	4.8
3 Tropomyosin 1	75.2	6.5	7.2	5.4
4 Tropomyosin 2	32.5	6.9	8.2	5.5
5 Myosin heavy chain	262	5.8	6.8	3.5
6 Myosin-2 essential light chain	16.8		5.3	
7 Putative uncharacterized protein (Myosin_tail)	60.1	5.7	7.1	3.9
8 Calcium-transporting ATPase	72.9		4.8	
9 Calponin	20.3	6.9	6.7	7.4
10 Putative troponin C	18.4		7.0	
11 Troponin 1	23.8		6.7	
12 Troponin T	47.3	6.9	6.7	7.1

In our study, among all muscle proteins identified, protein tropomyosin was found as one of the most abundant proteins in pellet (LC-MS/MS). Corresponding to myofibril proteins found in *T. molitor*, myosin heavy chain had a molecular weight of 262 kDa, and myosin light chain of 16.8 kDa. Myosin heavy chain and light chains from sardines (*Sardinella longiceps*) showed molecular weights of 205, 31, 23, and 22 kDa [5]; myosin from white mackerel muscle had three light chain subunits with Mw of 26.5, 20, and 17.5 kDa [6].



A.



B.

Figure 1A and 1B: Protein digestibility (%) of the whole insect and insect fractions with increasing gastric digestion time (10, 20, 30, 60 and 120 min (n=2), and continue with duodenal digestion (120 min).

The digestibility of the water-soluble protein fraction (supernatant, about 80%) was higher than that of water-insoluble protein fraction (pellet 50% and residue 24%) after *in vitro* gastro-duodenal digestion as determined by the OPA essay. In comparison to water-soluble proteins of conventional meat, [7] mentioned on sarcoplasmic proteins, *e.g.* nitrogen content after digestion was 67% for chicken, 89% for beef, 88% for lamb and 87% for pork of total nitrogen after pepsin/pancreatin digestion.

III. CONCLUSION

This study gives insight in the bulk protein composition of *T. molitor* and the *in vitro* digestibility, thereby contributing to knowledge needed for future food applications of this insect species. The main conclusion is that proteins from insects do not behave very differently from conventional meat protein.

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REFERENCES

Paper:

1. Van Huis, A. (2013). Potential of Insects as Food and Feed in Assuring Food Security. *Annual Review of Entomology*, 58(1).
2. Yi, L., Lakemond, C. M. M., Sagis, L. M. C., Eisner-Schadler, V., van Huis, A., & van Boekel, M. A. J. S. (2013). Extraction and characterisation of protein fractions from five insect species. *Food Chemistry*, 141(4), 3341-3348.
3. Vreeburg, R. A. M., van Wezel, E. E., Ocaña-Calahorra, F., & Mes, J. J. (2012). Apple extract induces increased epithelial resistance and claudin 4 expression in Caco-2 cells. *Journal of the Science of Food and Agriculture*, 92(2), 439-444.
4. Lu, J., Boeren, S., de Vries, S. C., van Valenberg, H. J. F., Vervoort, J., & Hettinga, K. (2011). Filter-aided sample preparation with dimethyl labeling to identify and quantify milk fat globule membrane proteins. *Journal of Proteomics*, 75(1), 34-43.
5. Mathew, S., & Prakash, V. (2006). Effect of Calcium Salts on the Properties of Proteins from Oil Sardine (*Sardinella longiceps*)

During Frozen Storage. *Journal of Food Science*, 71(4), E178-E183.

6. Watabe, S., & Hashimoto, K. (1980). Myosins from White and Dark Muscles of Mackerel Some Physico-Chemical and Enzymatic Properties. *The Journal of Biochemistry*, 87(5), 1491-1499.
7. Storcksdieck, S., Bonsmann, G., & Hurrell, R. F. (2007). Iron-Binding Properties, Amino Acid Composition, and Structure of Muscle Tissue Peptides from *in vitro* Digestion of Different Meat Sources. *Journal of Food Science*, 72(1), S019-S029.