TURKISH FERMENTED SUCUK: A SOURCE OF ANTIHYPERTENSIVE PEPTIDES

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Abstract: In search of new approaches against Angiotensin I Converting Enzyme (ACE) action of mechanism in Turkish sucuk, fermented by Lactobacillus sakei and Lb. plantarum used as material. Physicochemical and biochemical analysis were carried out at 0., 7. and 14. days. Sucuk samples were subjected to hydrolyzation process with pepsin and trypsin and total ACE inhibitory activity was evaluated. At 0th day ACE inhibitory activity (IC50) of hydrolysates was the lowest in control sample (3.46 mg/ml), while the highest inhibition was resulted in Lb. sakei (2.25 mg/ml) sample. Our results exposed that the hydrolysate of Turkish fermented sucuk by lactic acid bacteria especially by Lb. sakei could serve as a source of peptides with antihypertensive activity. Keywords: ACE, hypertension, Lb. sakei, Lb. *plantarum*, sucuk

I. INTRODUCTION

Recently, individuals started to consider their foods and its nutritional quality. There is a public demand for exploring new nutraceutical materials from certain and renowned foods to overcome some lifestyle related diseases (LSRD) such as diabetic, obesity, cardiovascular diseases (CVD) and hypertension [1]. Hypertension a dreadful disease which results a reduction in patients' life quality. Hypertension can be described as high blood pressure inside the blood vessels (Blood pressure >120/ 80 mmHg) [2]. While the number of hypertension patients consist %14.9 of World population in 2007, it is estimated to reach higher levels through the 2020 [3]. Level of blood pressure in human body is being regulated by an endocrine system which is called Rennin Angiotensin System (RAS). Angiotensin Iconverting enzyme (ACE), а dipeptidylcarboxypeptidase, most important fragment of this system plays a vital role in regulating blood pressure [4]. The ACE reacts on inactive Angiotensin I, and forms Angiotensin II, a vasoconstrictor, by cleaving His-Leu dipeptide [5]. Thus, Angiotensin II gives rise to constriction of blood vessels and as a result of it, causes hypertension. Consequently, ACE activity must be inhibited by different methods. Although there are pharmalogical drugs for curing hypertension, along with their ACE inhibition activity, drugs may also induce some side effects in human body [3]. With this justification, natural ACE inhibitors are requested to dominate activities associated with ACE. Hereby, during last decade much attention has been paid for safer applications to avoid from side effects of drugs and treat or minimize hypertension. Bioactive peptides from food sources, found to be promising approach to overcome hypertension.

Fermented Turkish Sucuk is a traditional meat product which is made of meat, fat, red and black pepper, cumin, pimento, salt and sugar. As a result of high amount of meat protein, sucuk is a great source of peptides. Throughout fermentation process proteins are being degraded into smaller peptides inside of sucuk [6]. Consequently, newly generated biologically active peptides can be realized and therefore nutritional value of sucuk has to be determined especially against ACE action of mechanism to value that delicious meat product. In present study we determined effects of different starter cultures (*Lb. sakei* and *Lb. plantarum*)on some physiological properties and total ACE inhibition of Turkish fermented sucuk.

II. MATERIALS AND METHODS

Materials: Four days vacuumed *Biceps femoris* muscle of 2-2.5 years old Montofon bovine was purchased from Saray Çiftliği Meat and Meat Product Company, Kayseri. Sheep tail fat and all spices were purchased from local markets in Kayseri.

Sucuk Production: Lb. sakei and Lb. plantarum, isolated from traditional Turkish sucuk, were used as starter culture for sucuk production. Four different types of sucuks such as; control (no inoculation), *Lb. sakei*, *Lb. plantarum* and combination type (inoculated with both *Lb. sakei* and *Lb. plantarum*) were produced and subjected to following analysis. Experimental sucuk dough was prepared according to Öz et al. [7]. Minced meat and fat along with the spices were mixed homogenously, and then starter cultures were inoculated into the dough separately at a ratio of 1 % (10^7 cfu/g). Sucuk dough mixtures were stuffed into synthetic collagen casings (36-mmØ) and ripened in fermentation cabinet (Nüve/Turkey) at 24±1°C, 90±2 % relative humidity (RH) for 5 days, and then at 22±1°C, 85±2 % RH for 5 days and finally 18±1°C, 80±2 % RH for 4 days.

Dry Matter and pH values: Dry matter and pH analysis were done according to AOAC [8].

Protein Extraction: Guba-Straub-Adenosine Triphosphate (GS-ATP) Soluble Proteins and Water Soluble Proteins (WSP) were extracted by adding 28 mL of solution to 2 g of the samples [9].

Sodium Dodecyl Sulfate Polyacrylamide Gel Electrophoresis (SDS-PAGE): GS-ATP and WSP extracts were separated according to their molecular weight (MW) by using SDS-PAGE technique. Samples were run on gradient gel made of 7.5%–20% at a constant current of 30 mA/gel with a Mini protein II unit (Bio-Rad Laboratories, Inc., Richmond, CA, USA) [10].

Hydrolysis of Sucuk Samples: Pepsin from porcine gastric mucosa (37 °C, pH=2.0, 2 hours and intestinal trypsin (37 °C, pH=7.0, 2 hours (Sigma–Aldrich, Inc. St. Louis, MO, USA) were applied on sucuk at an appropriate enzyme /substrate, respectively. Hydrolysates were filtrated trough A5 filter paper (Advantech Tokyo K. Ltd., Tokyo, Japan) and then 0.45 μ m filter and filtrates were collected for the ACE inhibitory assay.

Protein Concentration: Protein concentrations *were* determined by using the Biuret method [11]. *ACE inhibitory activity assay:* The ACE inhibitory activity of samples were determined according to the method of Cushman&Cheung [12], with slight modifications as described by Katayama et al. [13].

III. RESULTS AND DISCUSSION

Dry Matter and pH values: In the course of fermentation, there was a remarkable increase in dry matter (Table 1). No significant differences observed between the sucuk types at 0th day. However, at 7th days *Lb. sakei* sample was statistically different (p<0.05) compared to other 3 types and had the lowest dry matter. After 14days of ripening *Lb. sakei* and *Lb. sakei* + *Lb. plantarum* samples differed and had lower dry matter values (p<0.05). A significant decrease was observed in pH values between 0thday to 7th days. However a slight increase noted from 7th day to 14th day. Results suggested that due to higher microbial growth, accumulation of lactic acid was high during first seven days of ripening.

Table 1. Physicochemical properties of Sucuk samples in different time points.

Param eter	Control	LS	LP	LSP
Dry Matter				
0	44.86 ^{Ca} ±	44.86 ^{Ca} ±	$44.86^{Ca} \pm$	44.86 ^{Ca} ±
	0.18	0.18	0.18	0.18
7	$64.13^{\text{Bab}} \pm$	$62.80^{\text{Bb}} \pm$	$65.04^{\text{Bab}} \pm$	$65.51^{\text{Ba}}\pm$
	0.78	0.01	1.45	0.09
14	75.04 ^{Aab}	$73.80^{Ab} \pm$	$76.18^{Aa}\pm$	$74.31^{\text{Ab}}\pm$
	± 0.40	0.19	0.63	0.65
pН				
0	5.66 ^{Aa} ±0.	5.66 ^{Aa} ±0	5.66 ^{Aa} ±0.	5.66 ^{Aa} ±0
	00	.00	00	.00
7	4.66 ^{Ca} ±0.	$4.56^{Cb} \pm 0$	$4.50^{Cc} \pm 0.$	$4.54^{Bb}\pm0$
	01	.00	01	.03
14	4.69 ^{Ba} ±0.	$4.60^{Bc} \pm 0$	4.61 ^{Bc} ±0.	$4.65^{Bb}\pm0$
	02	.00	03	.00

Different capital letters in each column show statistically significant differences between the time points (P<0.05), and different small letters in each row show statistically significant differences between the sucuk types (P<0.05). LS: *Lb. sakei*, LP: *Lb. plantarum*, LSP: *Lb. sakei*+ *Lb. plantarum*.

SDS-PAGE: WSP extracts of 0th day sample, before inoculations and fermentation, showed 13 protein bands on SDS-PAGE gel (Figure 1). During the first 7 days, bands correspond to Bovine Serum Albumine (BSA) 66 kDa, Glutamic Dehydrogenase 55 kDa, Ovalbumin 45 kDa disappeared and caused generation of a new and intense band with an approximate molecular weight of 40 kDa (Figure 1). Apparently in samples at 7th and 14th days, 4 bands were observed.





LS: Lb. sakei, LP: Lb. plantarum, LSP: Lb. sakei+ Lb. plantarum

GS-ATP extract of 0th day exhibited 16 different bands (Fig. 2). High molecular weight bands such as Myosin Heacy Chain (MHC) -200 kDa, B-Galactosidase-117 kDa, Phosphorylase- B 97 kDa and also Actin-45 kDa were disappeared and smaller proteins or peptides were generated. Six bands were visible at 7th days and those bands maintained their existence during further fermentation period. SDS-PAGE images indicated that degradation of large proteins into smaller proteins occurred during first seven days. A few minor changes were observed in protein structure among the sucuk types perhaps due to similar degradation mechanism of the inoculated starter cultures.

Figure 2. SDS-PAGE image of GS-ATP soluble proteins in different time points.



LS: Lb. sakei, LP: Lb. plantarum, LSP: Lb. sakei+ Lb. plantarum

Protein Concentration (P.C.): P.C. values of sucuk hydrolysates ranged between 14.92-15.67 mg/ml. As it is expected, control 0th day sample had the lowest P.C. and other all samples had

higher P.C. than control 0th. By the time even in control sample spontaneous microflora became dominant, caused fermentation and degradation which was also approved by SDS-PAGE images (Fig. 3). At 14th day, even though *Lb. sakei* is more proteolytic [14], *Lb. plantarum* sample had higher P.C..



Figure 3. Protein concentration of Sucuk hydrolysates in different time points (mg/ml).

C: Control, S: Lb. sakei, P: Lb. plantarum, S+P: Lb. sakei+Lb. plantarum

ACE inhibitory activity assay: Results were expressed as IC_{50} value which is concentration of bioactive substance to inhibit 50% of an enzyme or a radical. Due to low degradation, control 0th day sample had higher IC_{50} value (3.46 mg/ml). Between 14th day samples *Lb. sakei* had the lowest IC_{50} (2.25 mg/ml) (Fig. 4). Results suggested that inoculation of *Lb. sakei* into sucuk may cause generation of more antihypertensive peptides that may help inhibition of ACE action of mechanism to avoid formation of Angiotensin II, a vasoconstrictor in RAS.





C: Control, S: *Lb. sakei*, P: *Lb. plantarum*, S+P: *Lb. sakei*+ *Lb. plantarum*

IV. CONCLUSION

Antihypertensive bioactive peptides is a promising approach for replacing pharmacological drugs to deal with hypertension. Because of degradation during the course of fermentation, Sucuk becomes a valuable source of different peptide sequences which may act like bioactive peptides. A purification process is currently ongoing by HPLC as an attempt to reach and identify the bioactive peptides.

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REFERENCES

1. Jiménez-Colmenero, F., Carballo, J., and Cofrades, S. (2001). Healthier meat and meat products: their role as functional foods. Meat Science 591: 5-13.

2. Katayama, K., et al. (2008). Porcine Skeletal Muscle Troponin Is a Good Source of Peptides with Angiotensin-I Converting Enzyme Inhibitory Activity and Antihypertensive Effects in Spontaneously Hypertensive Rats. Journal of Agricultural and Food Chemistry 562: 355-360.

3. Ahhmed, A. M. and Muguruma, M. (2010). A review of meat protein hydrolysates and hypertension. Meat Science 861: 110-118.

4. Skeggs, L., Khan, E., and Shumaay, P. (1965). The preparation and function of the hypertensin-converting enzyme. Journal of Experimental Medicine 103: 95-299.5. Ryan, J. T., et al. (2011). Bioactive Peptides From

Muscle Sources: Meat and Fish. Nutrients 39: 765-791. 6. Dalmış, Ü. and Soyer, A. (2008). Effect of processing methods and starter culture (Staphylococcus xylosus and Pediococcus pentosaceus) on proteolytic changes in

Turkish sausages (sucuk) during ripening and storage. Meat Science 802: 345-354.

7. Öz, F., Kaya, M., and Aksu, M. İ. (2002). The effect of different nitrite doses and starter culture usage on the growth of Escherichia coli O157:H7 in the sucuk (Turkish style dry sausage) processing. Turkish Journal of Veterinary and Animal Sciences 263: 651-657.

8. AOAC, *Official Methods of Analysis (17th Ed.)* Chemists, A.O.O.A., Editor. 2000: Arlington, VA.

9. Abdulatef, M. A. (2014). Traditional cured meatmaking process degrades the proteins of M. latissimus dorsi of bovine. International Food Research Journal 211: 139-148. 10. Laemmli, U. K. (1970). Cleavage of structural proteins during the assembly of the head of the bacteriophage T4. Nature 227: 680-685.

11. Gornall, G. A., Baradawill, J. C., and David, M. M. (1949). Determination of serum protein by means of the biuret reaction. Journal of Biological Chemistry 177: 751-766.

12. Cushman, D. W. and Cheung, H. S. (1971). Spectrophotometric assay and properties of the angiotensin-converting enzyme of rabbit lung. Biochemical Pharmacology 20: 1637–1648.

13. Katayama, K., et al. (2004). Inhibitory Profile of Nonapeptide Derived from Porcine Troponin C against Angiotensin I–Converting Enzyme. Journal of Agricultural and Food Chemistry 524: 771-775.

14. Sanz, Y., et al. (1999). Hydrolysis of muscle myofibrillar proteins by Lactobacillus curvatus and Lactobacillus sake. International Journal of Food Microbiology 532–3: 115-125.