

Pre- and postnatal development of adipose depots in meat animals with a specific focus on the pig

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Outline

- ☐ **Introduction**
- ☐ **Structure, location and functions of adipose tissues**
- ☐ **Fetal and postnatal development and growth of adipose tissues**
- ☐ **Factors affecting body adiposity and intramuscular fat deposition**
- ☐ **Conclusion**

Adipose tissue : a tissue of great interest for meat producing animals

- ❑ **A significant compartment in the body in term of**
 - mass
 - physiological functions

- ❑ **An influence on production efficiency**
 -  - Selection for lean growth to increase the lean/fat ratio
 - Reduction in body fat

Adipose tissue : a tissue of great interest for meat producing animals

- ❑ **A role in the sensory, nutritional and technological qualities of meat and processed products**

Two parameters closely connected with the quality of meat

Intramuscular fat content (IMF)

- Sensory and technological qualities
- Important for high quality products

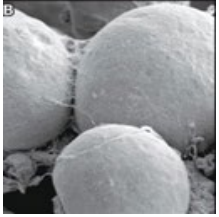
Fatty acid composition

Nutritional and technological qualities

Outline

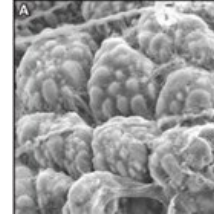
- Introduction
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Two main types of adipose tissue with differences in morphology and functions



White adipose tissue (WAT)

- ❑ Predominates after birth
- ❑ Contains white adipocytes
- ❑ Important for the storage and release of energy



Brown adipose tissue (BAT)

- ❑ Abundant in newborns and hibernating animals
- ❑ Contains brown adipocytes
- ❑ A thermogenic function



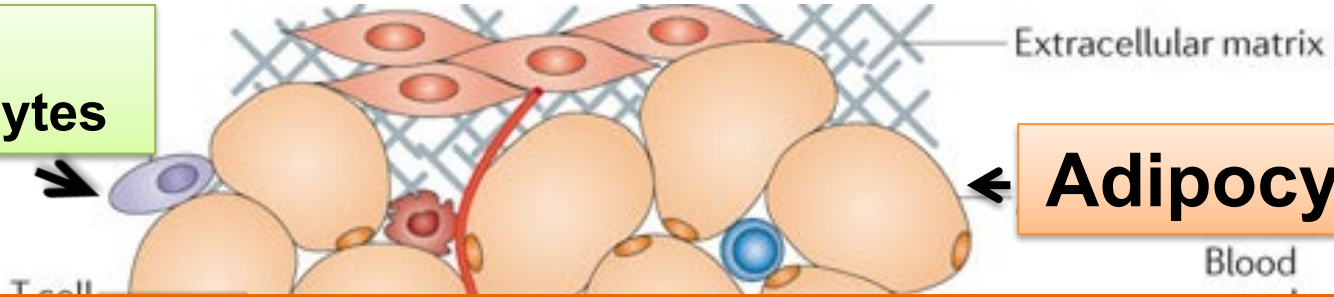
Absent in pigs
(Lack of functional UCP1)

Recent identification of a third type of cells in WAT: beige/brite adipocytes

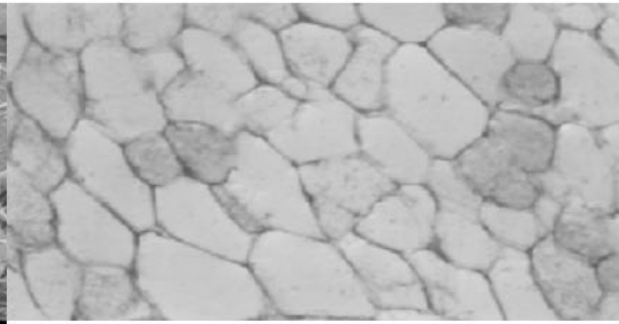
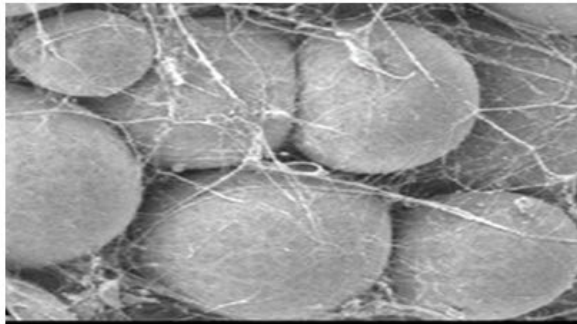
- ❑ **Detected in mice** (Wu *et al.*, 2012), **cattle** (Asano *et al.*, 2013) and **sheep** (Pope *et al.*, 2014)
- ❑ **Emerge in WAT depots in response to appropriate stimuli**
- ❑ **Store energy with the potential to express the mitochondrial membrane uncoupling protein 1 (UCP1)**
- ❑ **Beige adipocytes in pigs?**
 - still a controversy
 - cold-tolerant pigs (Tibetan) can maintain their body temperature through the “browning” of their WAT with an overexpression of UCP3 in the absence of UCP1 (Lin *et al.*, 2017)

WAT contains many cell types

Stem cells
Pre-adipocytes

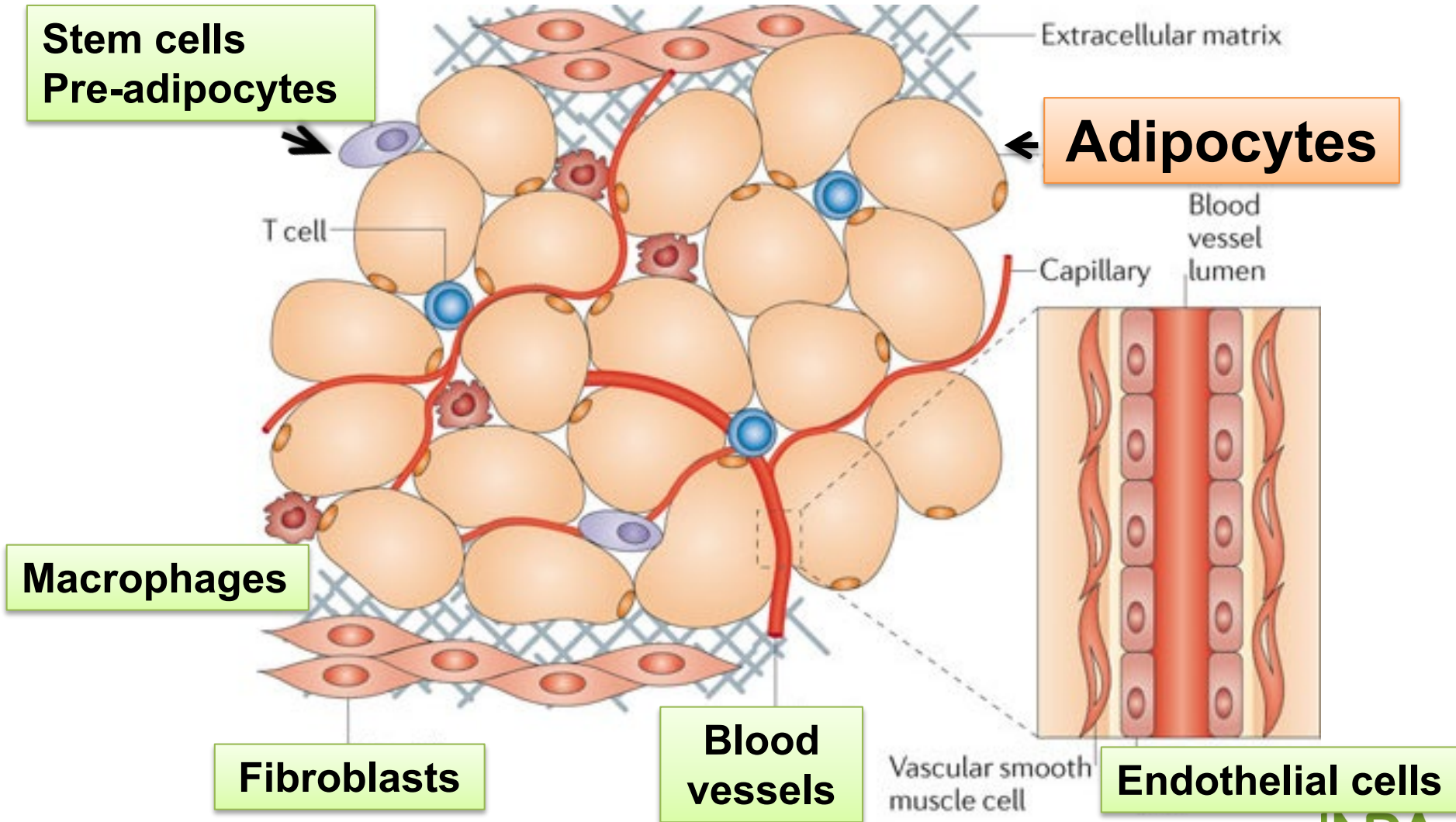


Adipocytes





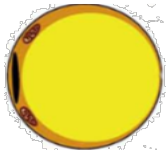
- ☐ Predominating cells in WAT (40-50% of total cells)
- ☐ Spherical cells when isolated with a wide range of diameters (20-120 μm)
- ☐ Cells detected in several tissues

WAT contains many cell types

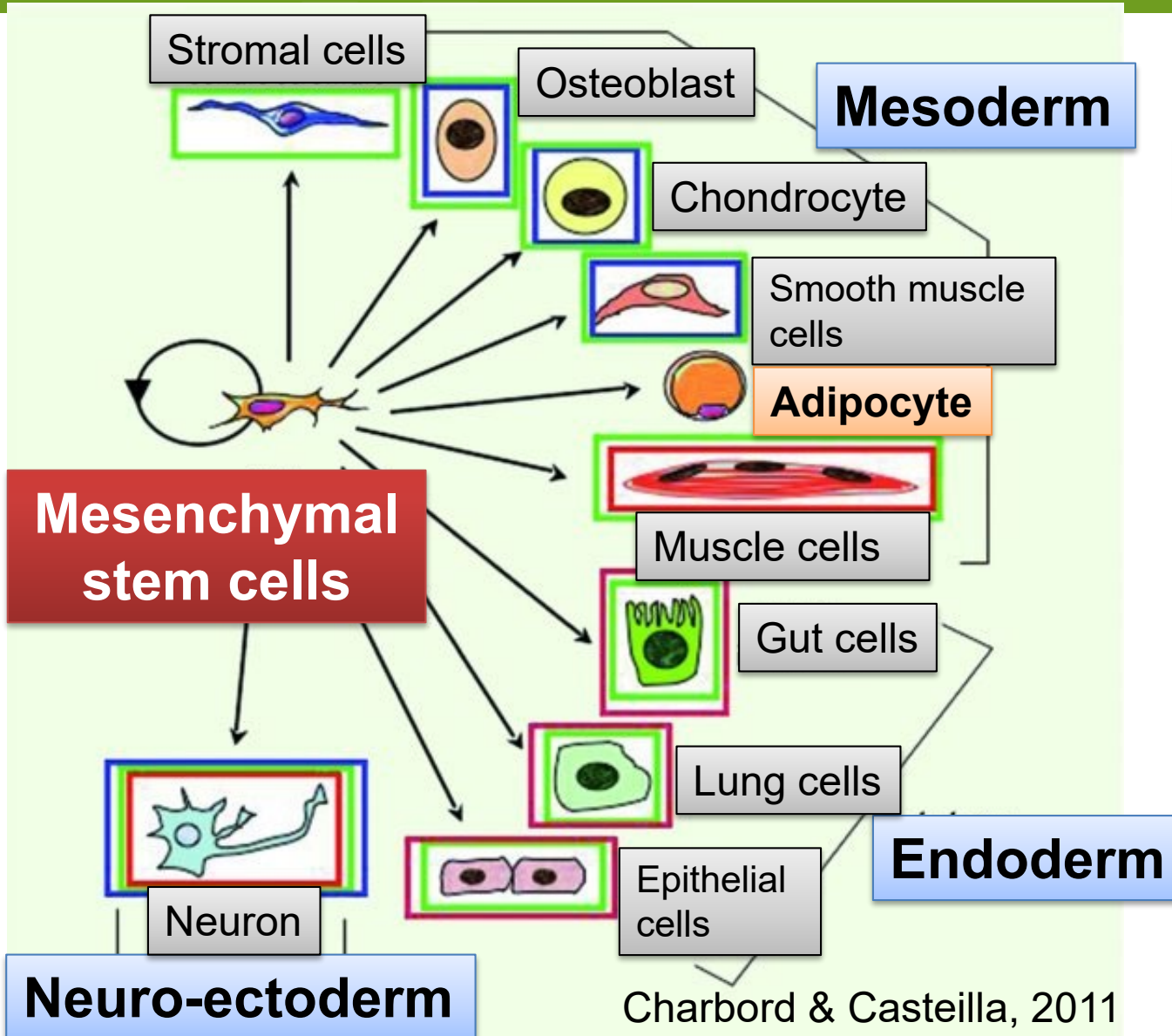


Adapted from Ouchi *et al.*, 2011

Features of brown, beige and white adipocytes

	Brown		Beige/Brite	White
			< 40 μm 	
Shape of lipid droplets	Multiple, small droplets			Single, large lipid droplet
Mitochondria	+++++			++
UCP1	High expression	Expression after cold exposure		Not detected
Function	Heat production with energy dissipation			Energy storage (triglycerides)

Origin of adipocytes?

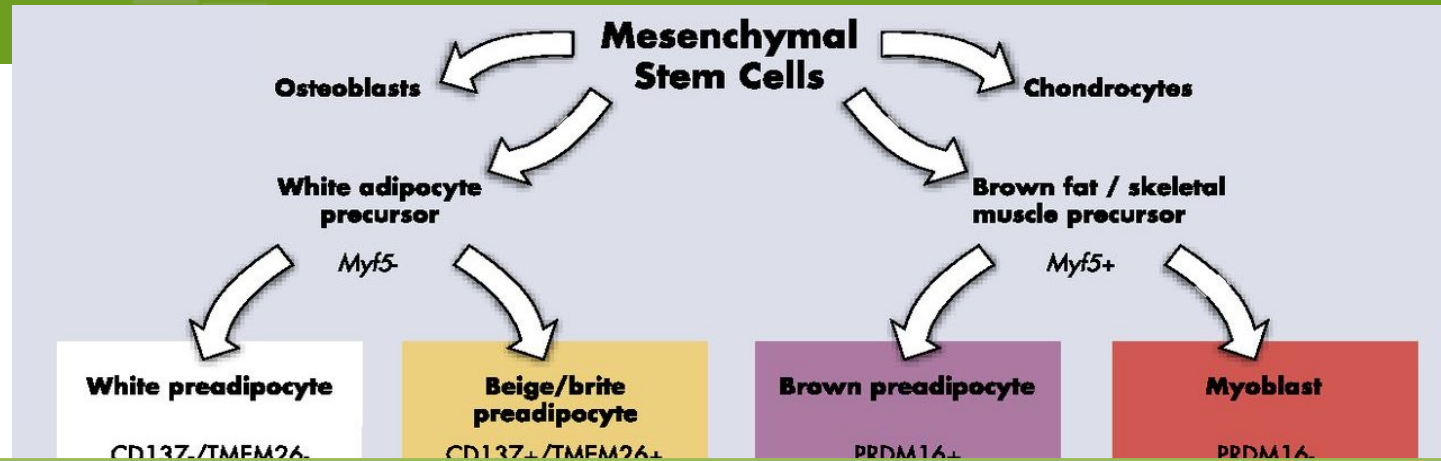


Adipocyte



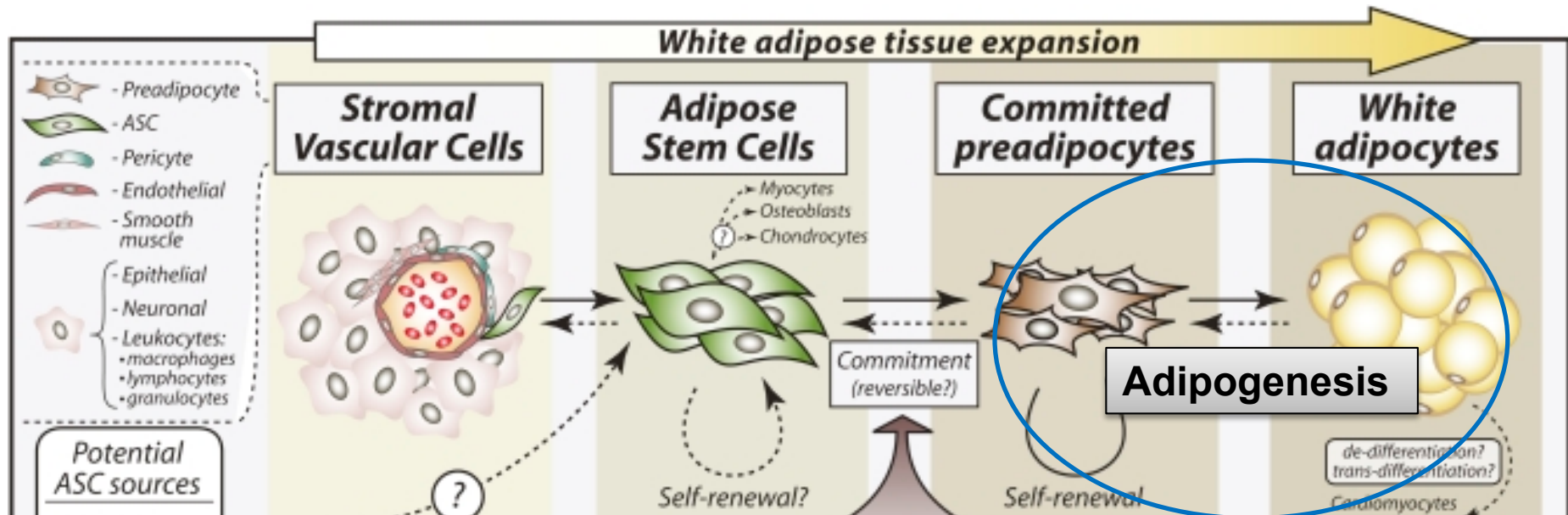
**Mesoderm
+
Neural
crest (?)**

Origins of white, beige and brown adipocytes



- ❑ In contrast to other tissues, the embryonic origin of adipose cells remains the subject of debate
- ❑ Available data support the idea that progenitor cells are heterogeneous and may have different embryonic origin

From stem cells to white adipocytes



Proliferation/Multi lineage capacity
↓ DLK1, IGF2

Specific function
↑ specific adipocyte genes (LPL, FASN, FABP)

Regulators that affect the differentiation of white, beige and brown adipocytes



- ❑ A large field of investigation including in livestock
- ❑ Some recent studies are investigating the role of long non coding RNA on adipocyte differentiation (Wei *et al.*, 2019)
- ❑ Many studies investigate the transcriptome of WAT with the aim to identify molecules that may be useful to better control variation in fat content but the biological interpretation remain a challenge (Dalrymple & Guo, 2017; Baik *et al.*, 2017)

Physiological functions of adipose tissue



Protection

- ☐ An insulating layer (reduction of heat loss through the skin)
- ☐ A protective function (providing mechanical protection and support around the major organs)

Physiological functions of adipose tissue

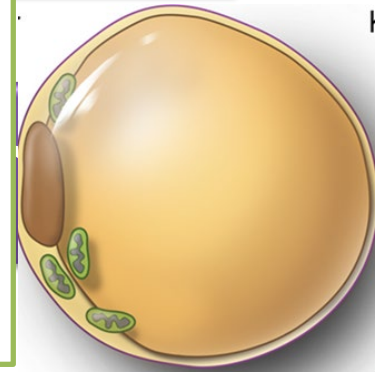


Protection

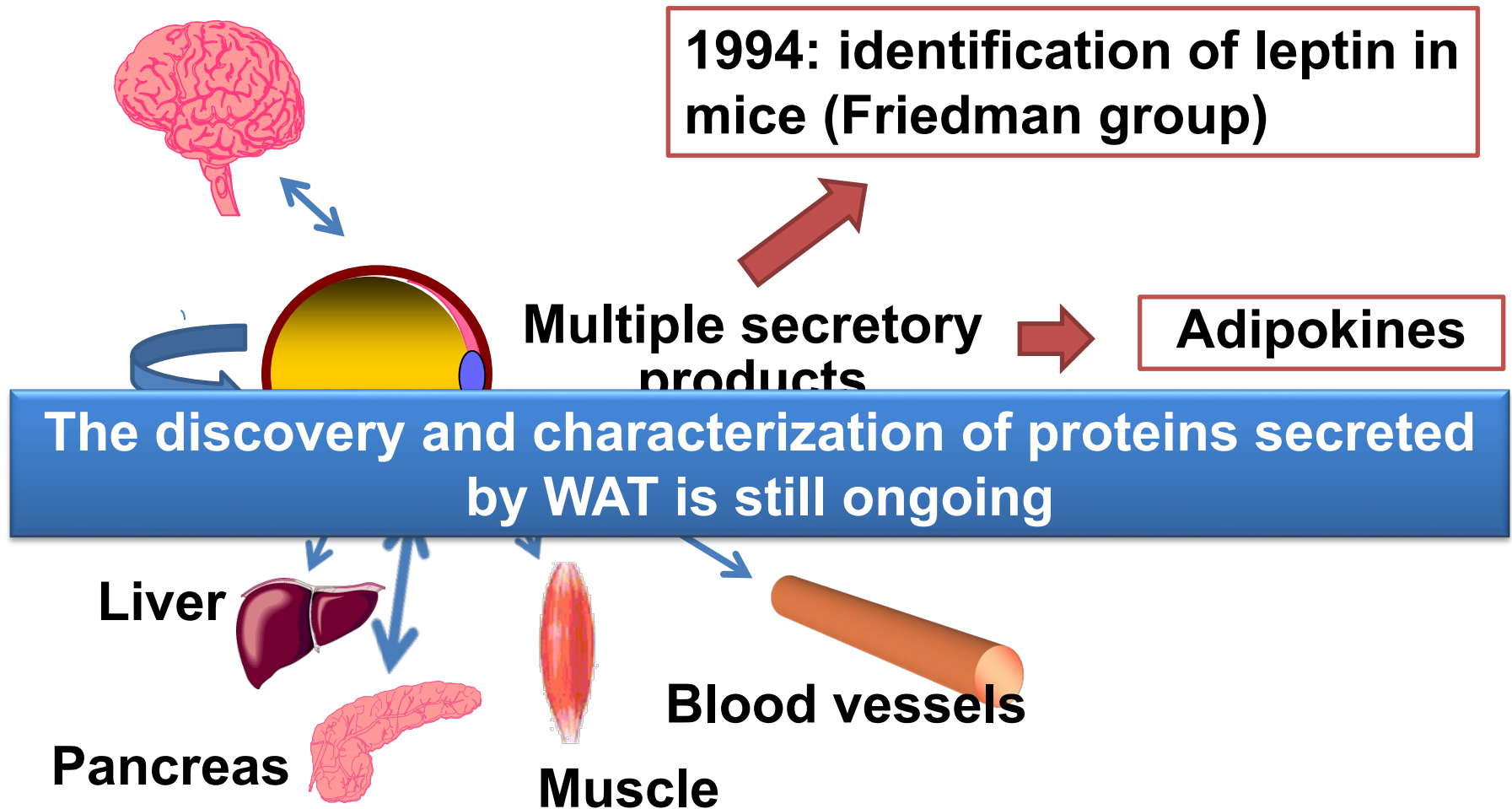
Energy storage

Storage and release of fatty acids and glycerol

Adipocyte: a central player in the control of energy balance and whole body lipid homeostasis

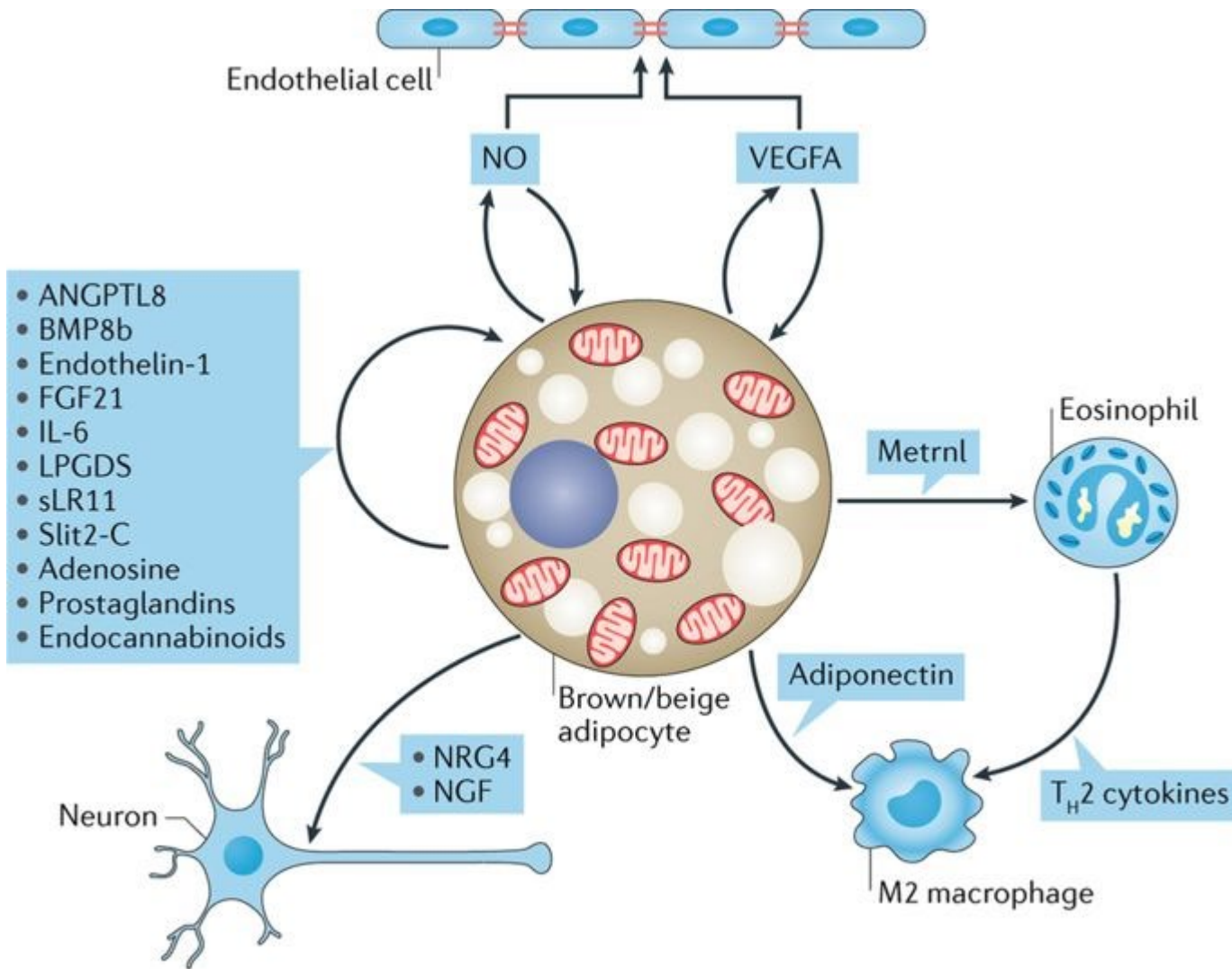


WAT: a secretory/endocrine organ



Lafontan, 2005; Haugen & Dreven, 2007; Rodríguez *et al.*, 2015, Giralt *et al.*, 2016

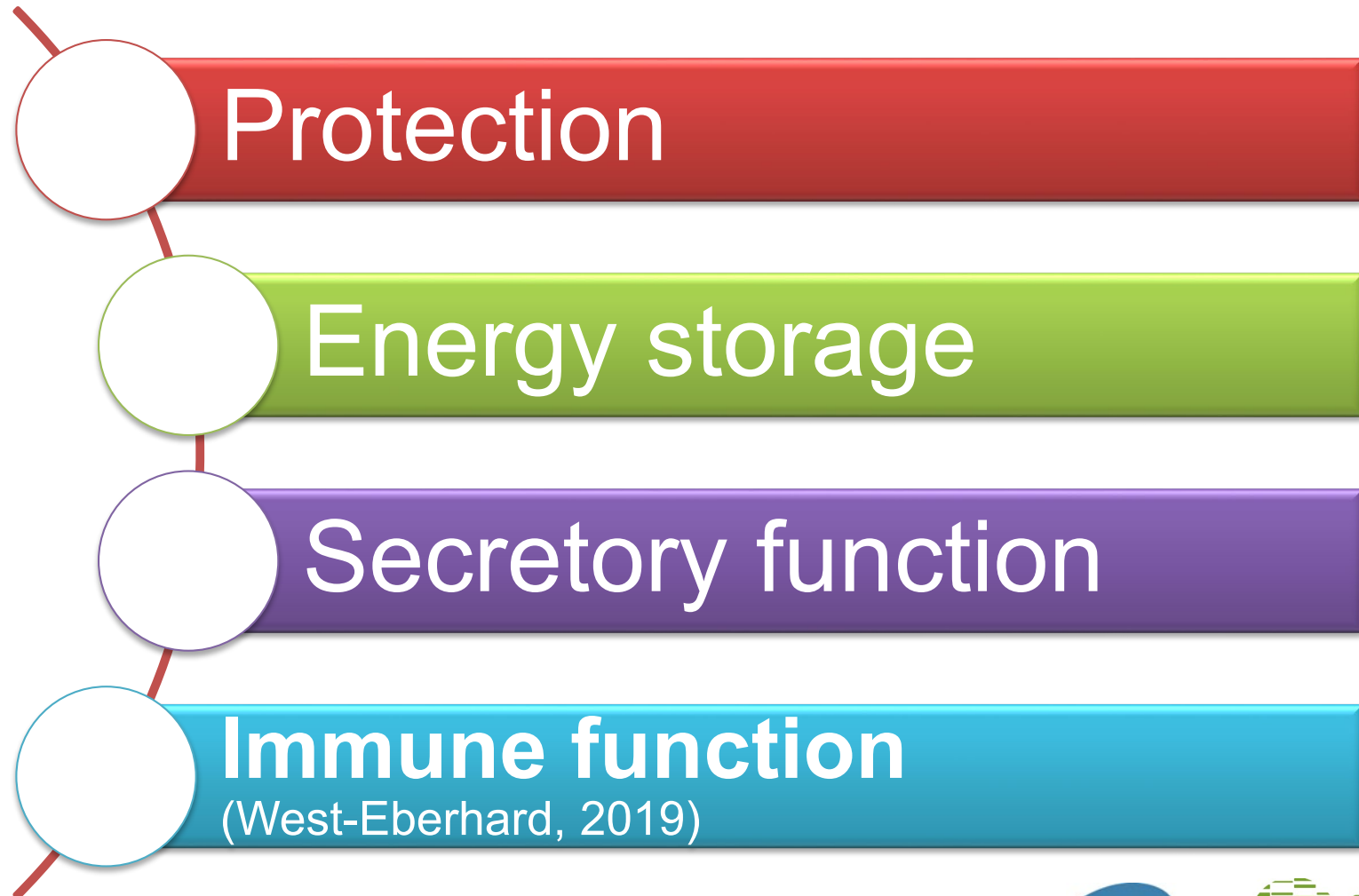
The secretory function of BAT



Villaroya *et al.*, 2017

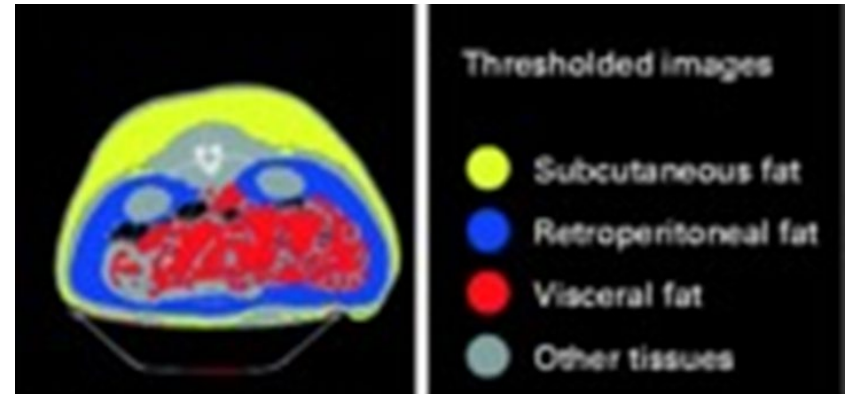
Nature Reviews | Endocrinology

Physiological functions of adipose tissue



WAT: : a number of individual depots in the body

Adiposity distribution (CT scan) (lumbar vertebra, L2, Val-Laillet *et al.*, 2010)



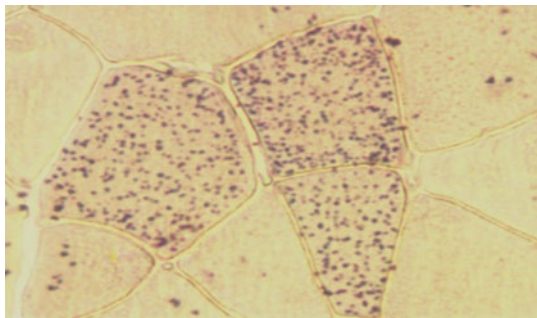
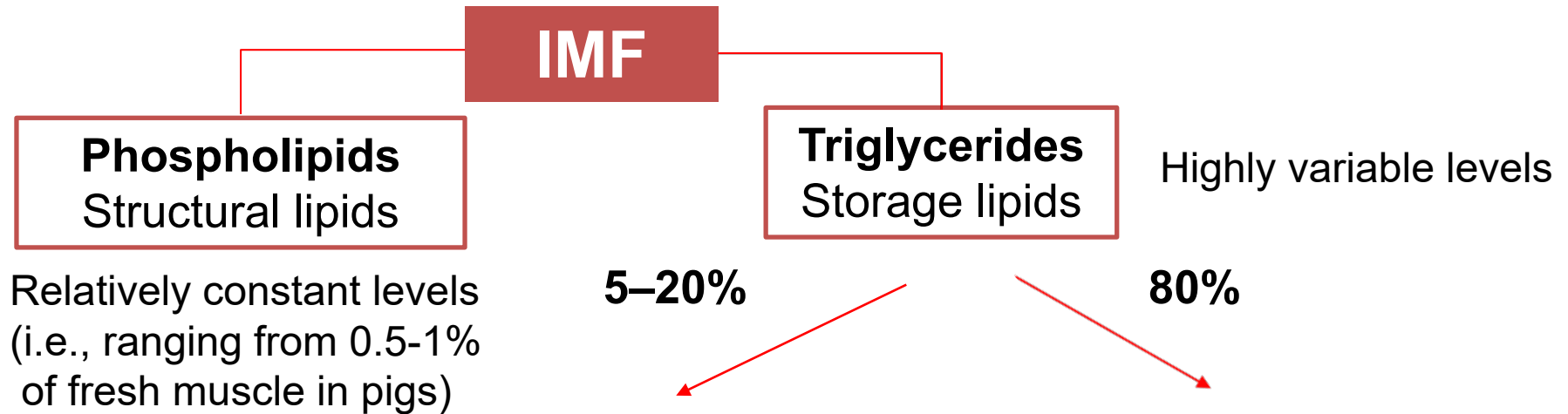
- Under the skin

- In the abdominal cavity
(surrounding viscera such as mesenteric and perirenal fat depots, around epididymis, etc.)

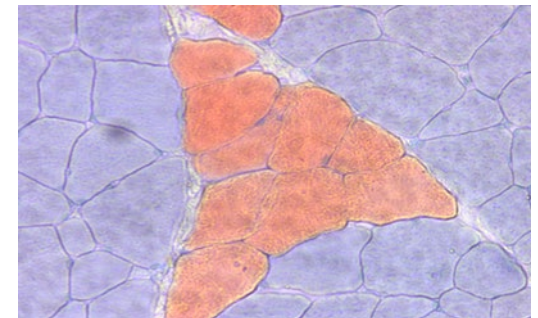
- Within the musculature
Inter- and intra-muscular depots

- Within the bone marrow

WAT: features of IMF depots



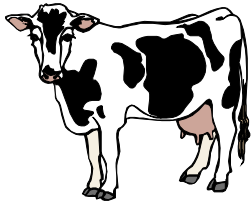
Lipid droplets
in the cytoplasm
within myofibers



Small groups of cells
located between muscle
fiber and fiber bundles

The relative proportions of WAT depots

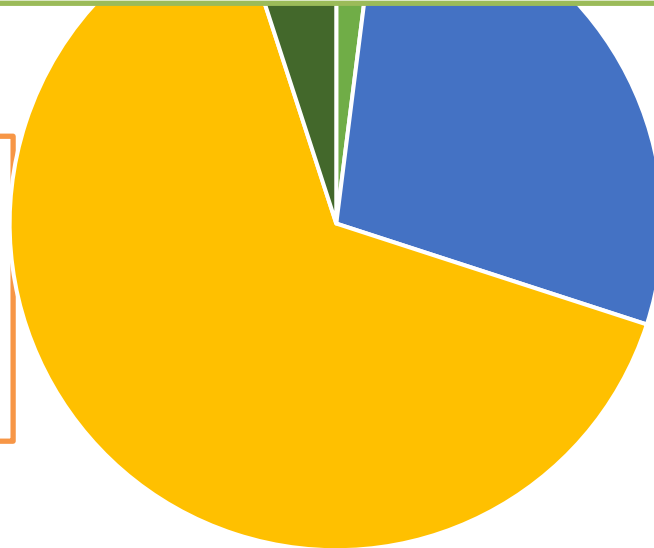
The relative size of these depots differs between species



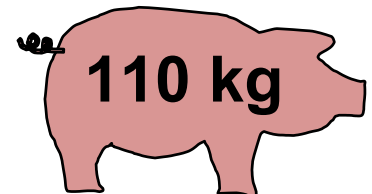
700 kg

SCAT: 12%
Intermuscular AT: 43%

**Subcutaneous AT
(65-75%)**



**Intermuscular AT
(25-33%)**



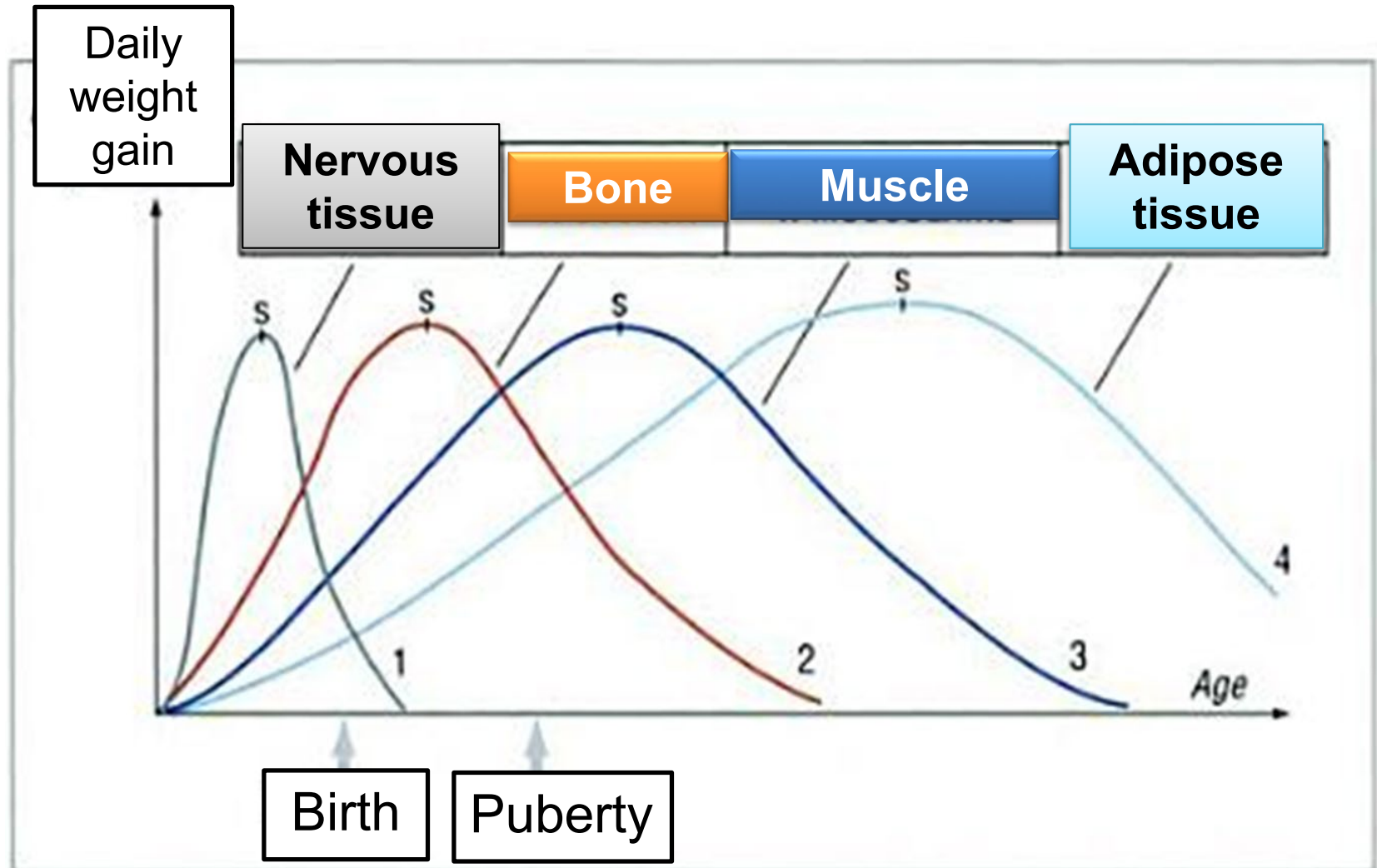
110 kg

Dumont & Février, 1957; Girard *et al.*,
1998; Monziols *et al.*, 2005

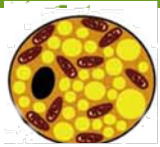
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Tissue accretion according to age

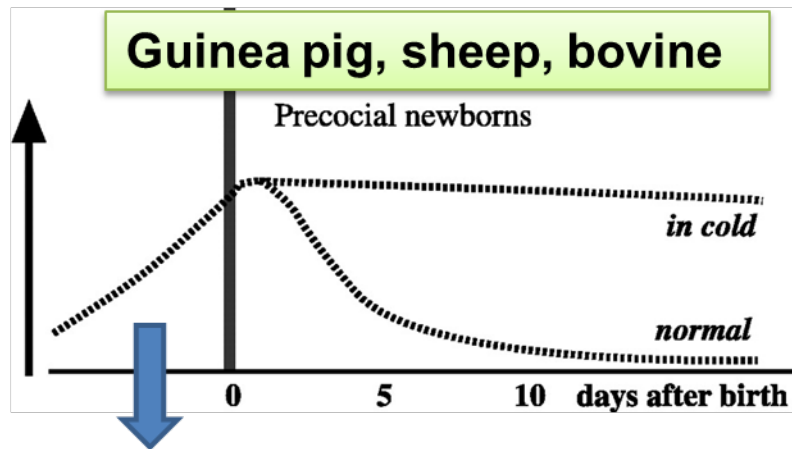


Age-related changes in BAT

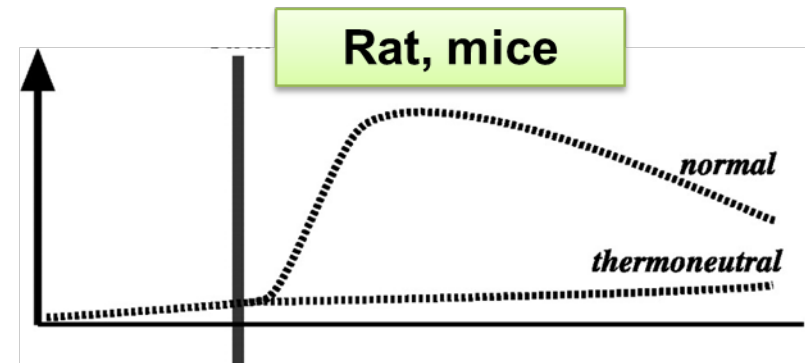


Found in mammals with exceptions: not detected in pigs

Birth



Birth

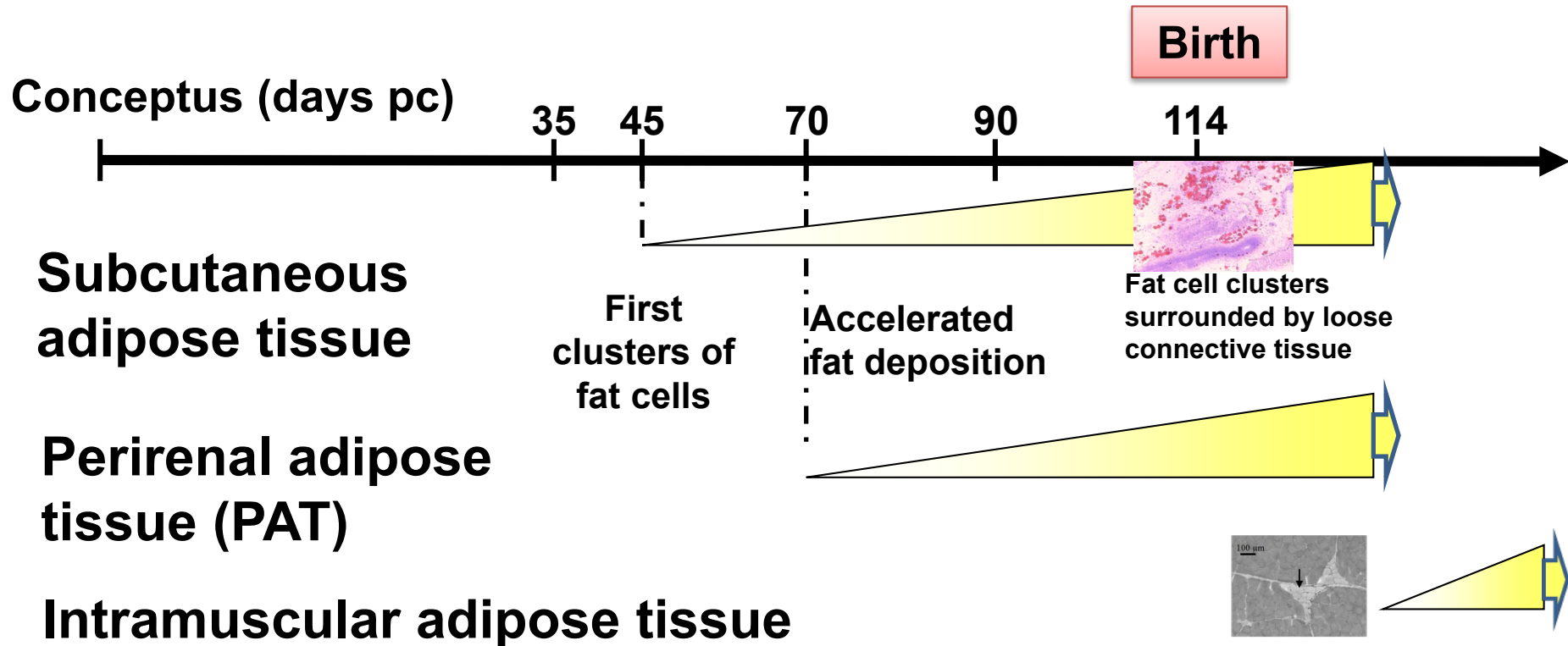


180 and 260 dpc fetuses

↪ perirenal adipose tissue is a mix of white and brown adipocytes

- ☐ present in adults: hibernating animals, rodents, humans +cow...
- ☐ BAT → WAT in large mammals ?

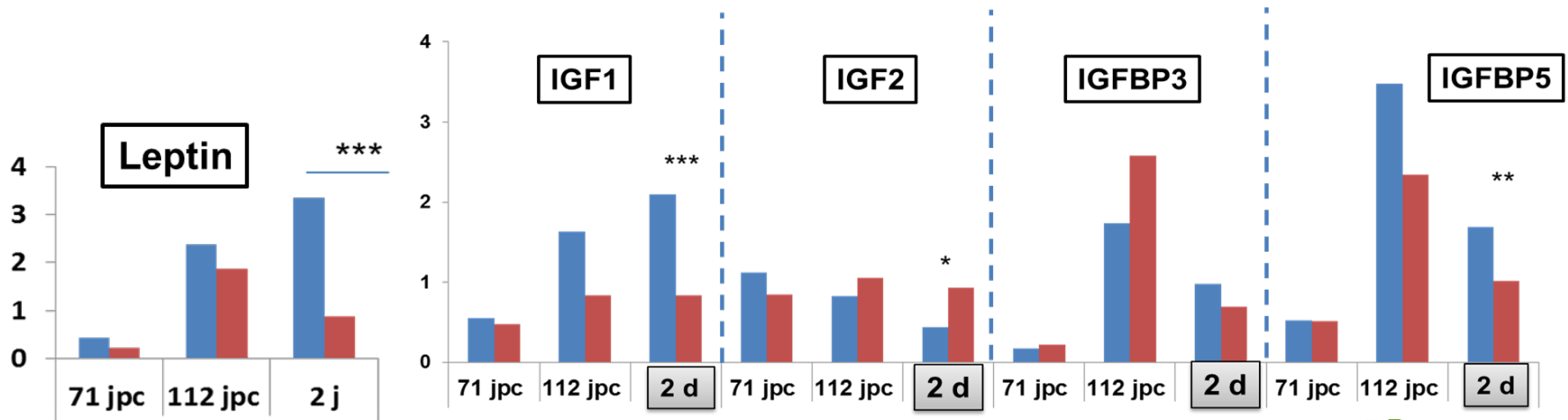
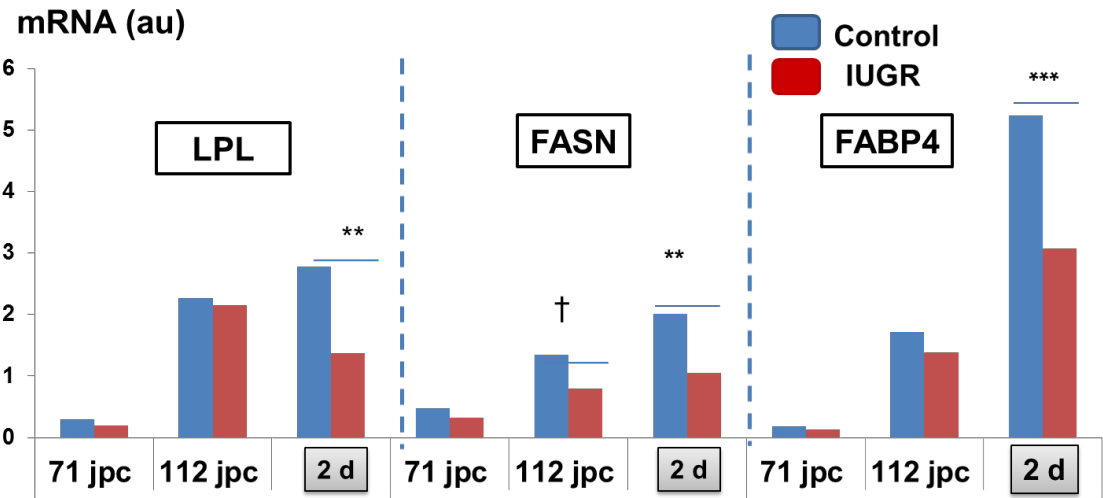
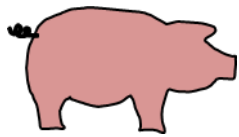
Prenatal and neonatal development of WAT



Adipose tissue development differs according to depot and according to species (PAT: the first detected in cattle)

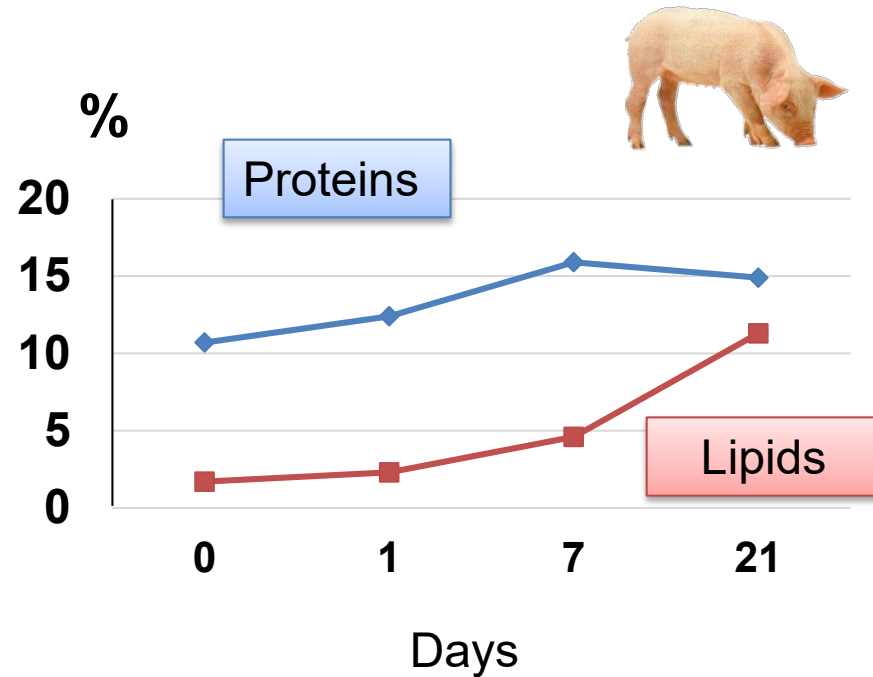
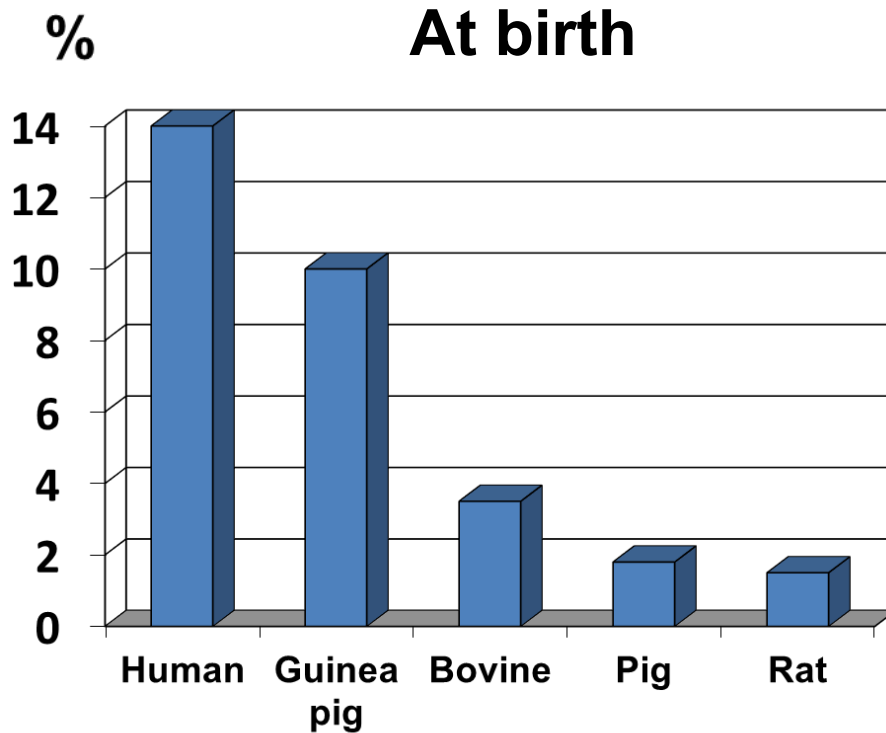
↑ adipose mass throughout life

Many changes occurs in the expression of genes during the prenatal and neonatal period



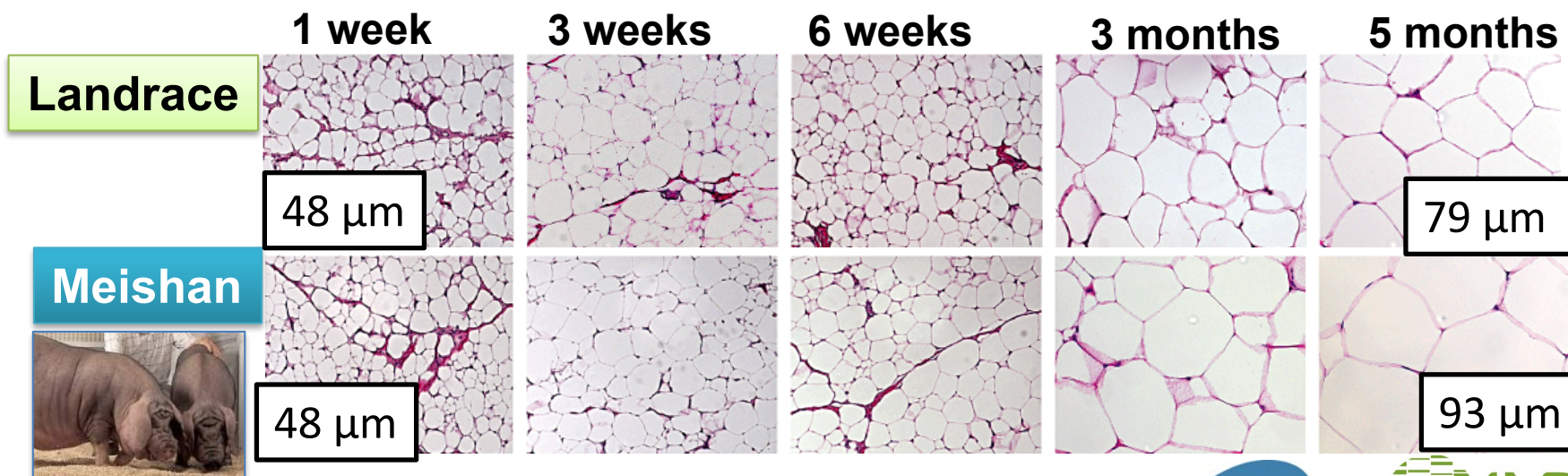
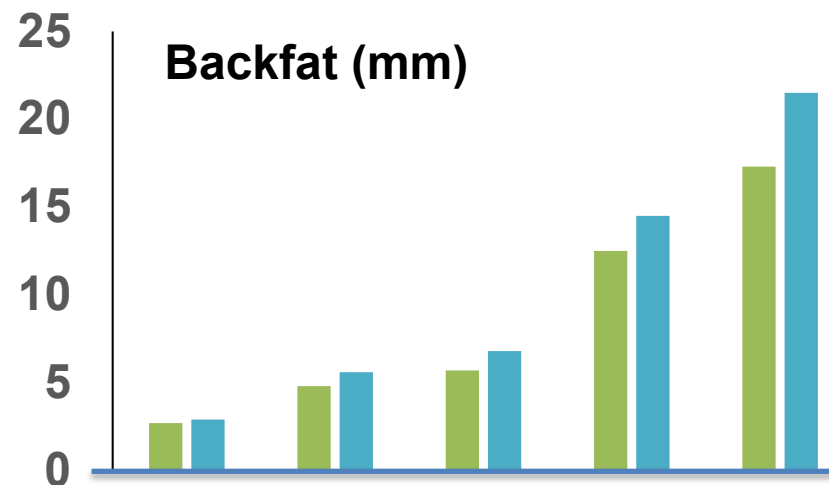
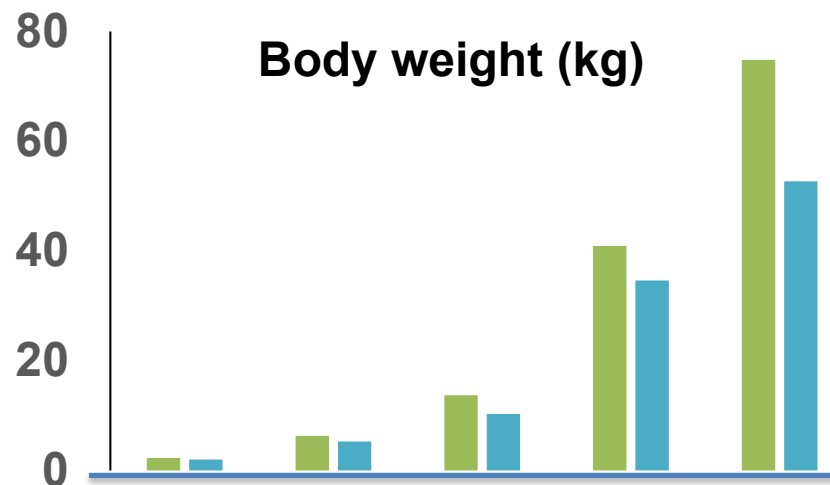
Gondret *et al.*, 2013

Body fat mass (%) in the neonatal period



➔ **Large differences between species**

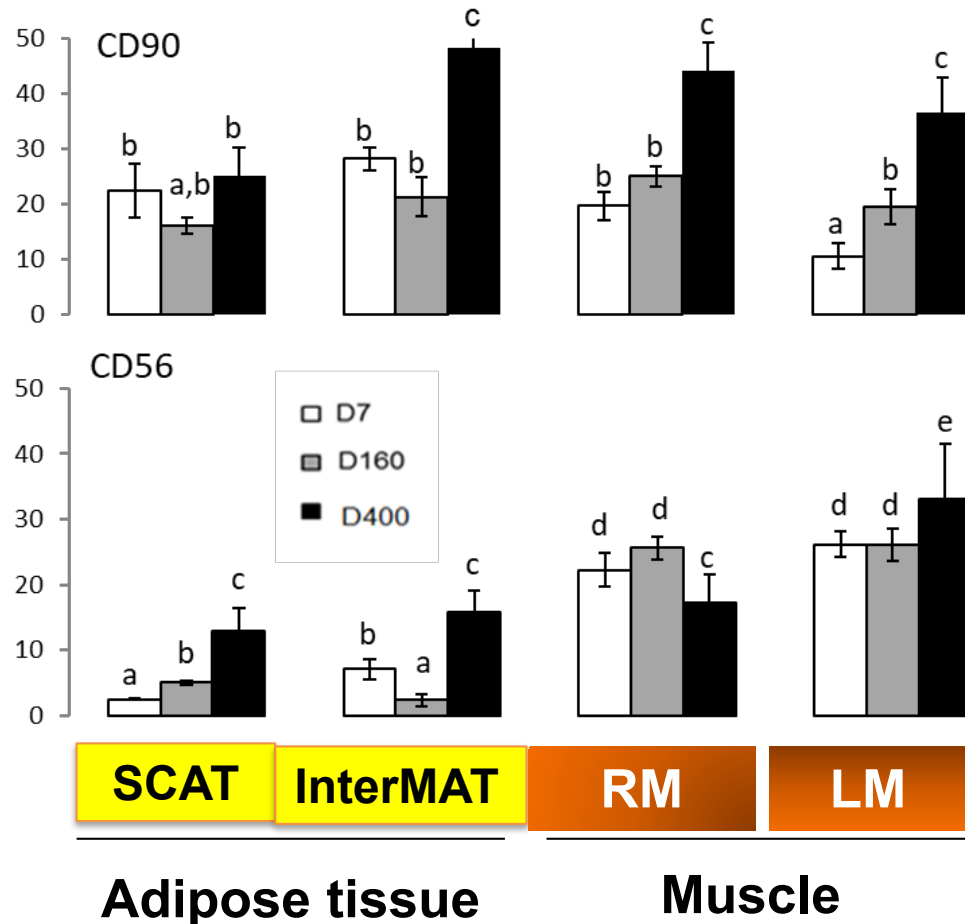
Cellular development of sc adipose tissue



Nakajima *et al.*, 2011

Adult stem cells in porcine WAT and skeletal muscle

Positive cells with adipogenic properties (%)

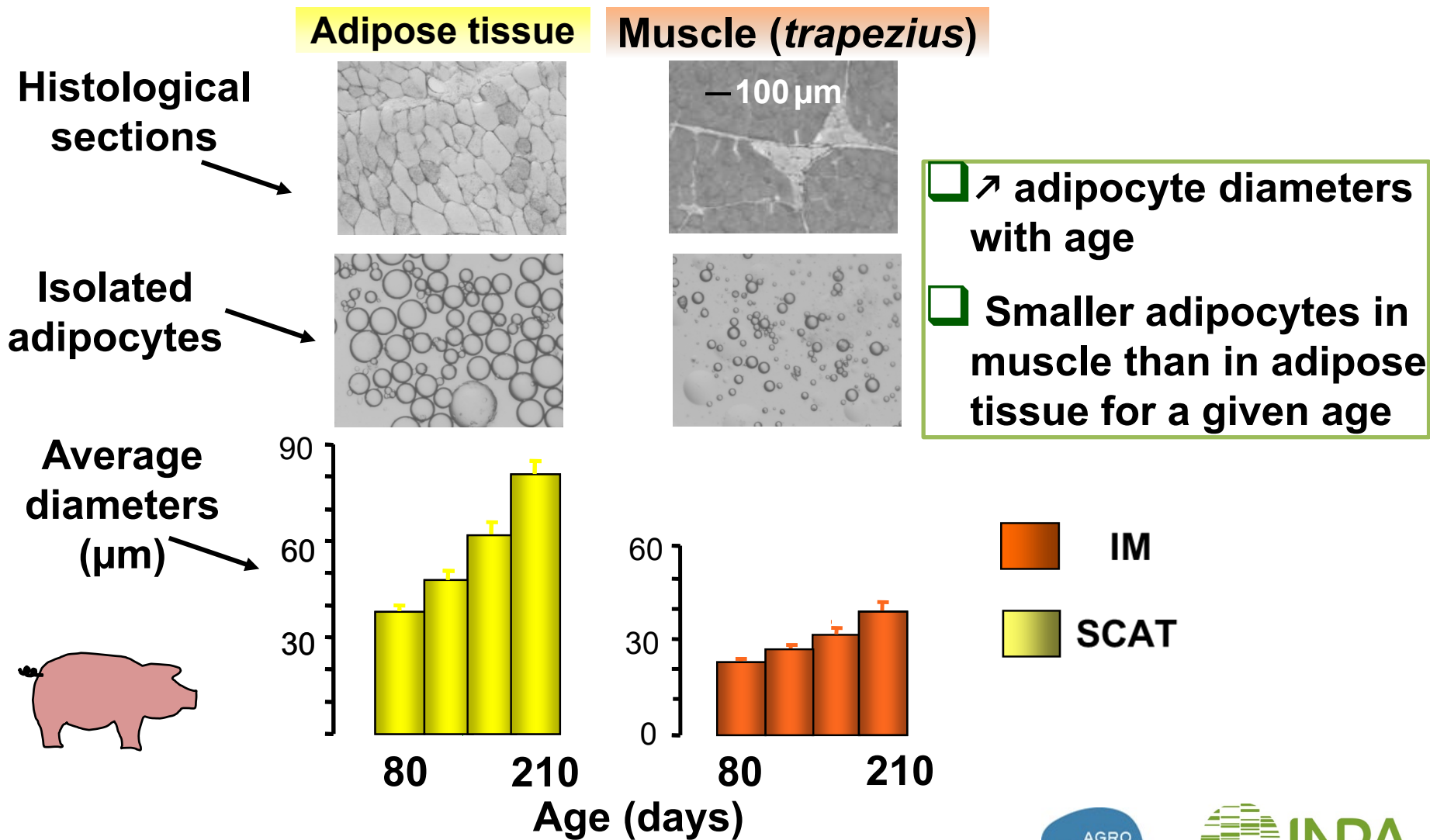


Age-related changes in stem cell populations with the ability to differentiate into adipocytes



A potential for increasing the number of adipocytes during the postnatal period

Site-specific development of adipocytes

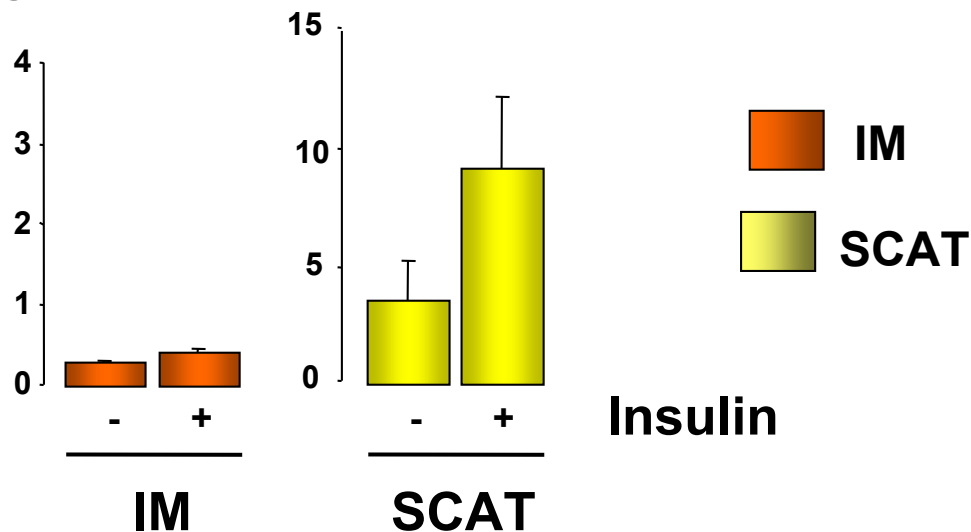


Differences in the physiology of adipocytes according to adipose depots

Lipogenesis

(adipocytes from 80-d-old pigs)

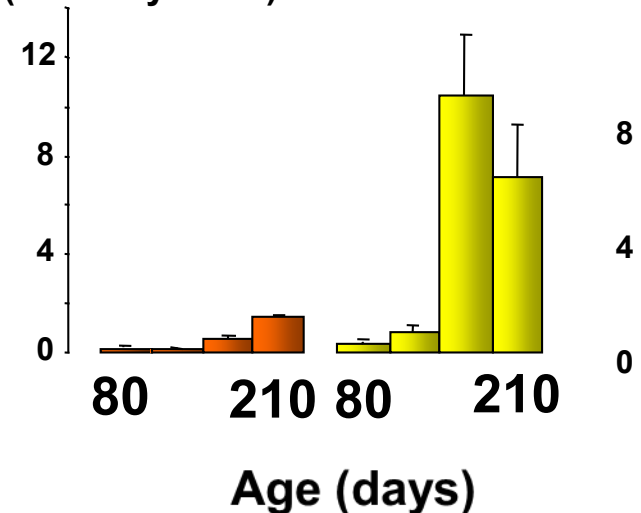
nmol glucose/4 h/ 10^5 cells



Leptin

mRNA

(arbitrary units)

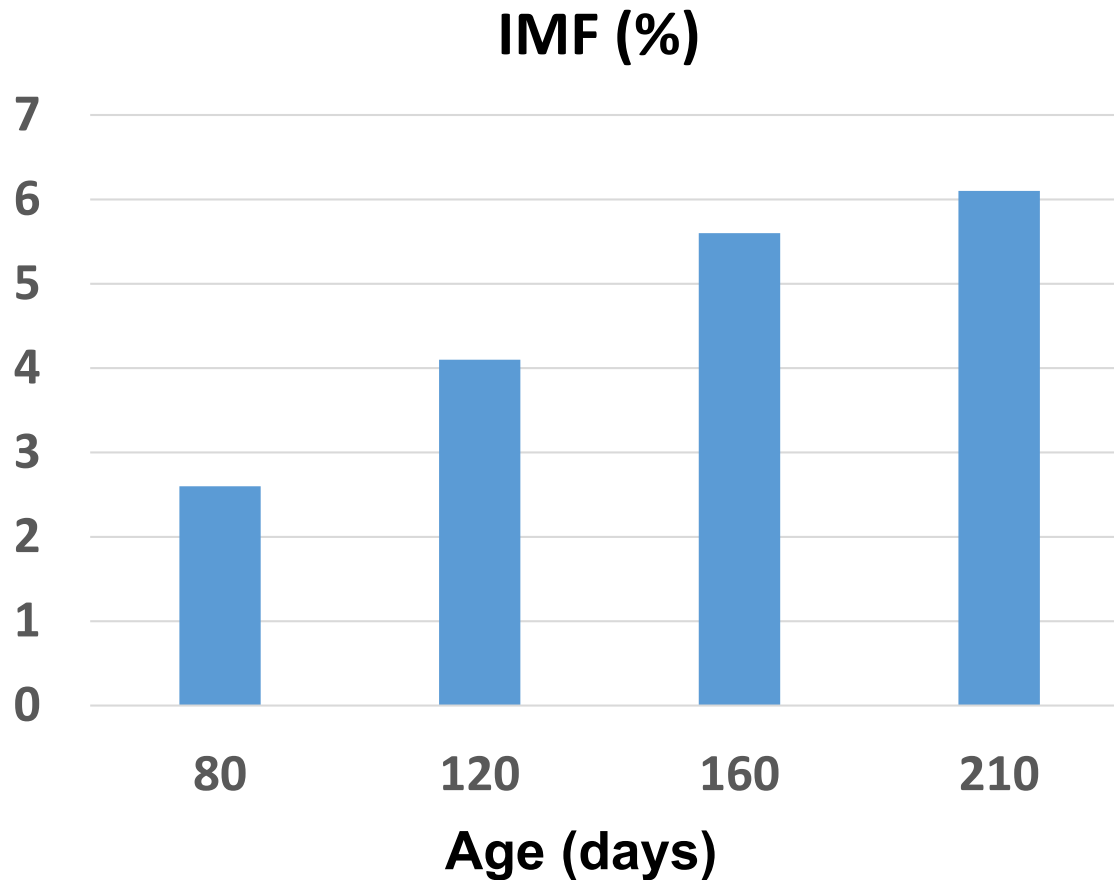
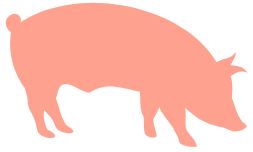


- Basal rate: IM << SC
- insulin-stimulated rate: IM << SC

SCAT >> IM

Gardan *et al.*, 2006

IMF content increases with age in the trapezius muscle

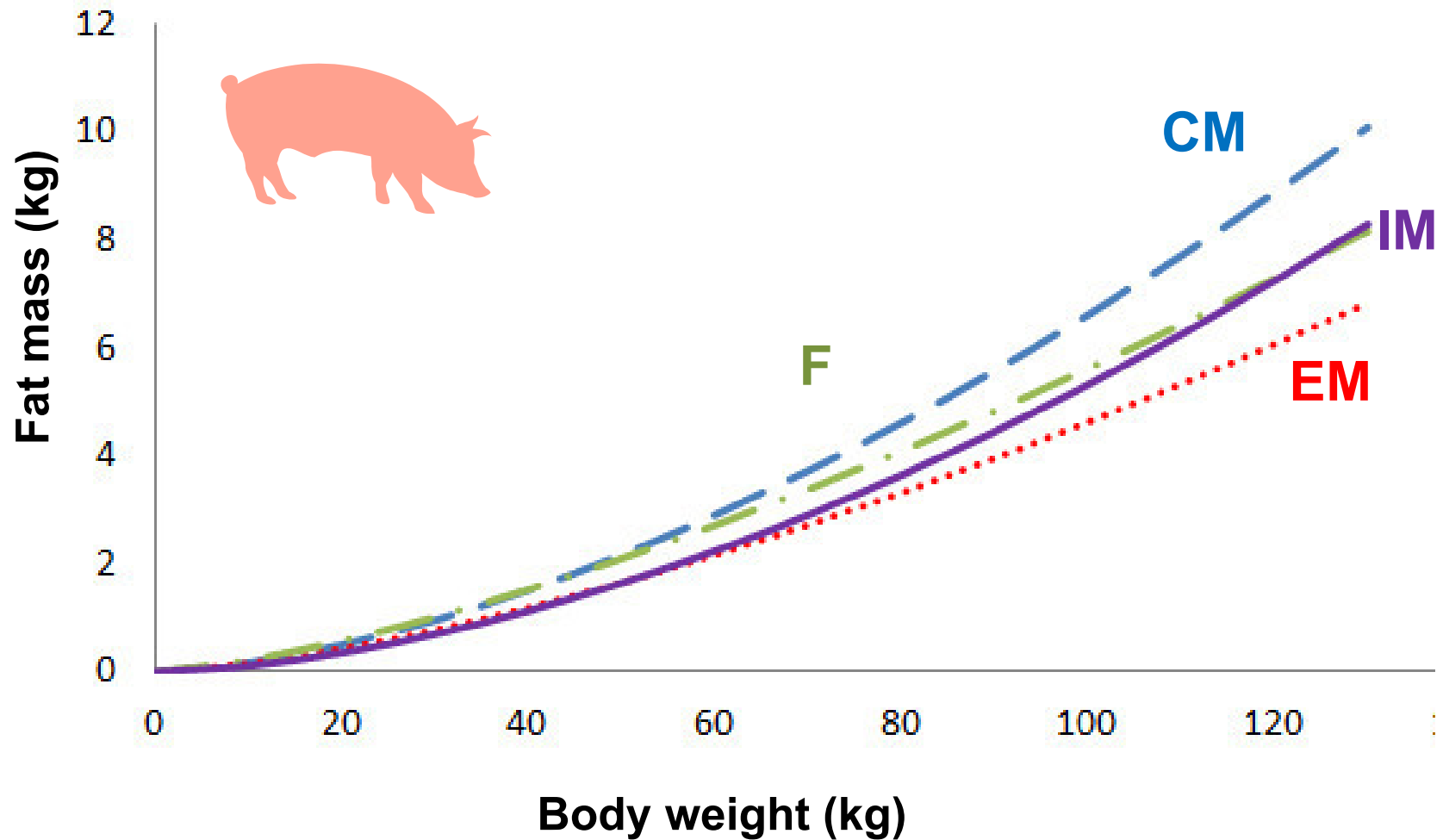


Gardan *et al.*, 2006

Outline

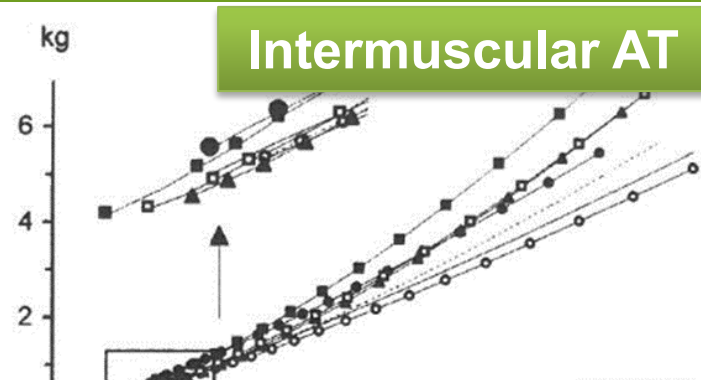
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Influence of age and sex on body fat mass



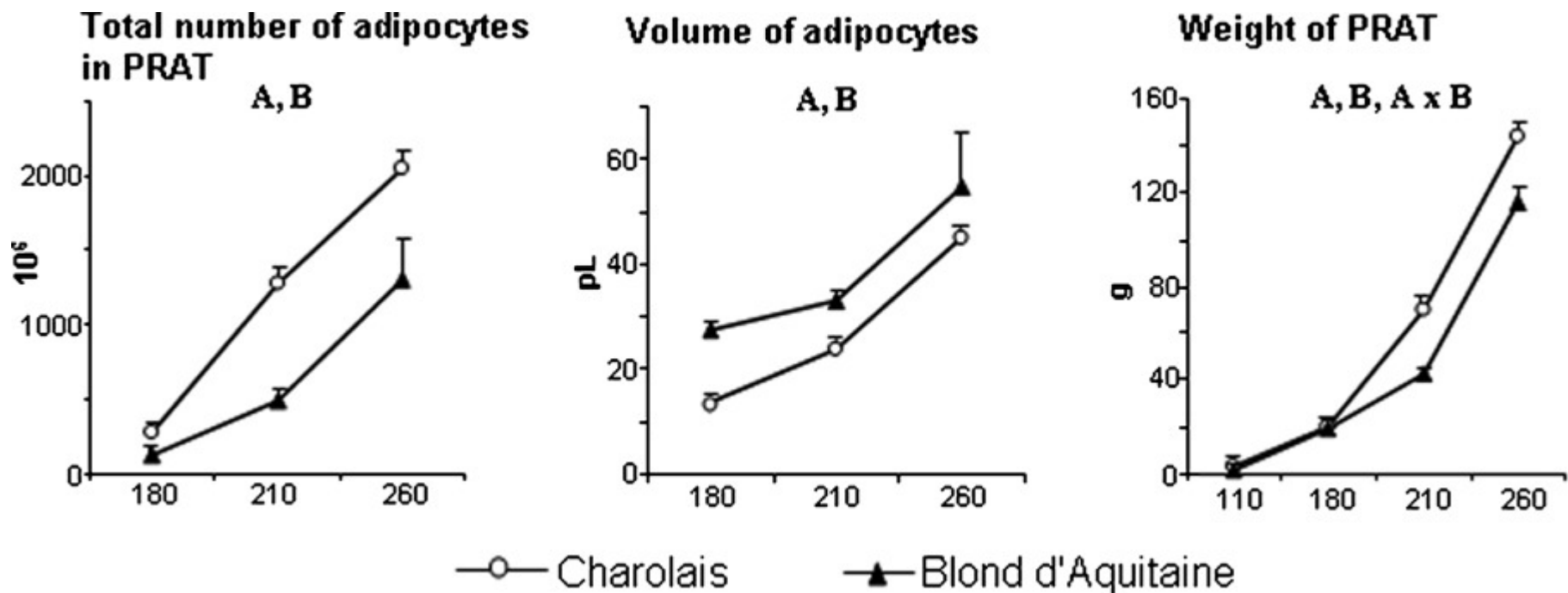
Carabús *et al.*, 2017

Mass of WAT depots in different pig breeds



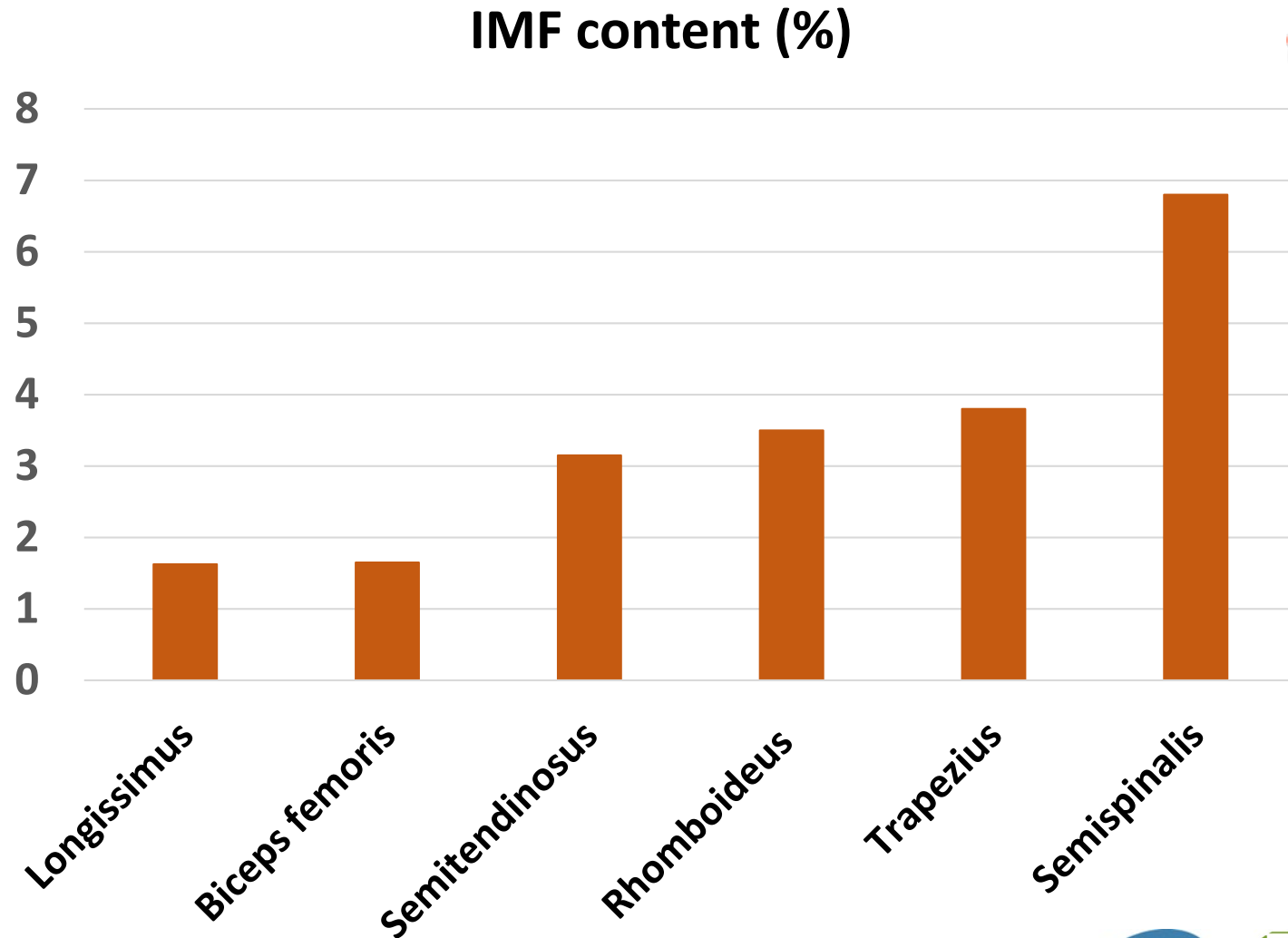
- ☐ The difference between breeds increases with body weight
- ☐ Differences between breeds are less important in intermuscular adipose tissue than in other depots

Perirenal WAT at 110, 180, 210, and 260 days post-conception in Charolais and Blond d'Aquitaine



Taga *et al.*, 2011

IMF content varies between muscles

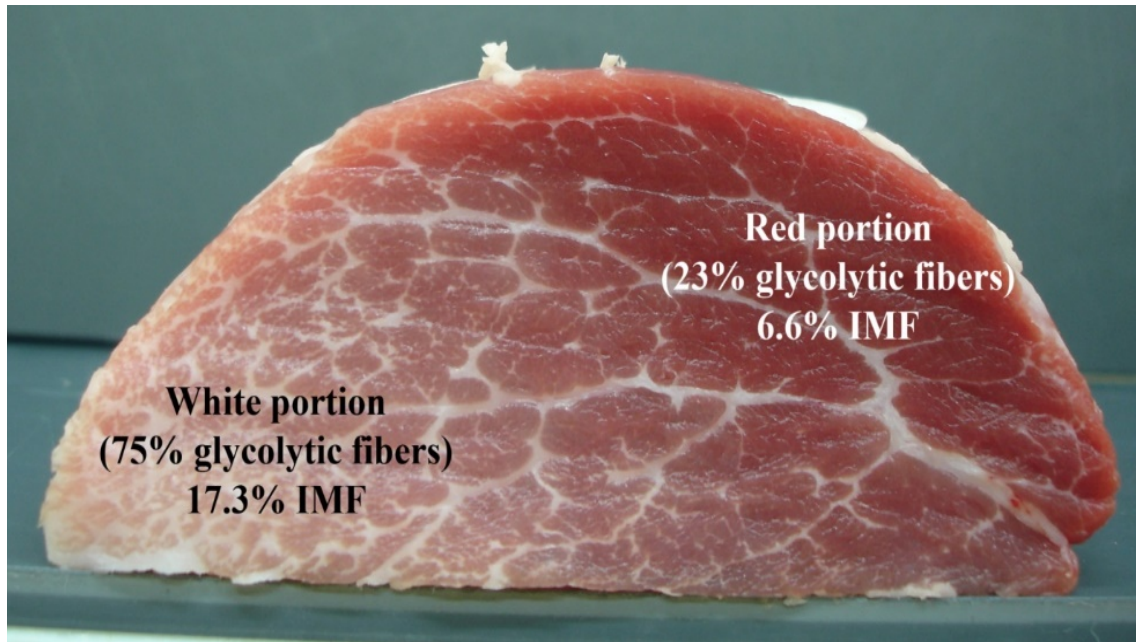


Maignel *et al.*, 2013; Tyra *et al.*, 2013; Font-i-Furnols *et al.*, 2018



IMF content depends on anatomical location within some muscles

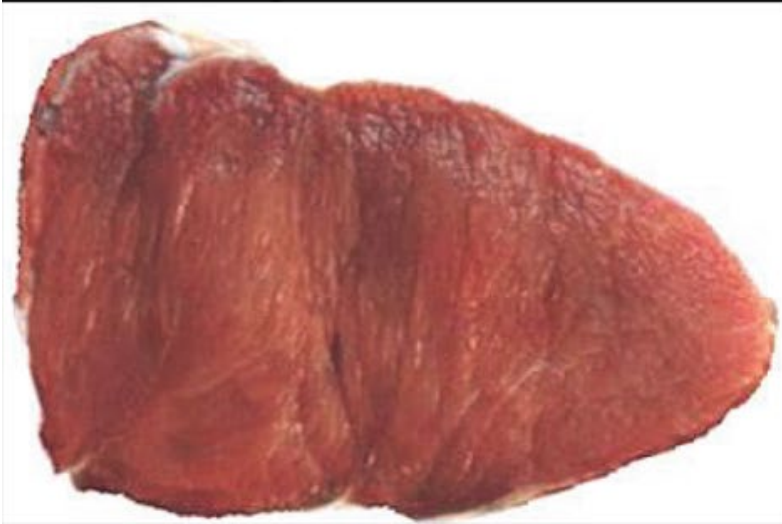
Semitendinosus muscle cross section from a Basque pig (145 kg live weight)



The IMF content is about three times greater in the white glycolytic than in the red oxidative portion of muscle

IMF content depends on breeds

Muscle cross-sections of LM



Belgian Blue bull
24 mo of age
IMF content: 0.5%



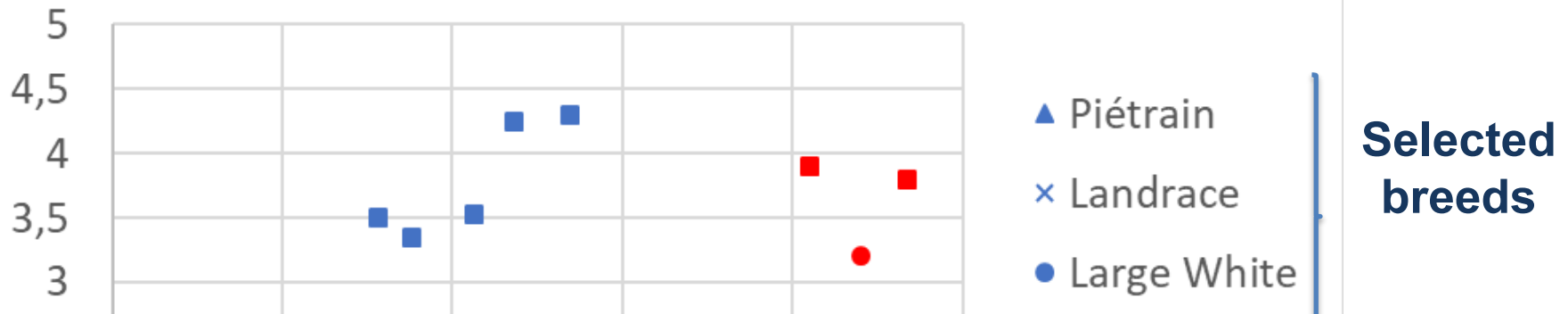
Japanese Black steer
26 mo of age
IMF content: 33% IMF

Albrecht *et al.*, 2017



IMF content and backfat thickness in different pig breeds

IMF in longissimus (%)



- ☐ The European conventional breeds (Large White, Landrace or Pietrain) have a low general adiposity
- ☐ Duroc pigs have limited BFT and IMF content varies according to studies
- ☐ Meishan pigs have moderate IMF content and high BFT
- ☐ French local breeds have a high overall adiposity (BFT and IMF content)

Influence of nutrition on fat mass/fatty acid composition

Nutrition and fat mass

- ❑ Many studies in this area of research
- ❑ Studies dealing with the impact of early nutrition on later adipose tissue development

Relationship between the nature of the ingested fat and meat fatty acid composition

- ❑ Monogastric animals: it is possible to change the FA composition of meat through the feed (Pork and chicken may be enriched with long-chain n-3 by inclusion of fish oil in the diet)
- ❑ Enrichment of beef and lamb is more challenging due to the extensive modification of dietary lipids by the rumen microbiome.

(Wood *et al.*, 1999; Hocquette *et al.*, 2010; Kouba & Mouro, 2011; Lebret *et al.*, 2015; Scollan *et al.*, 2017)

In summary

- ❑ **Several types of adipose tissue**
- ❑ **A number of individual depots in the body with**
 - **variable sizes**
 - **specific physiological properties**
- ❑ **AT development occurs later than skeletal muscle development: it is initiated prenatally except for intramuscular adipose tissue**
- ❑ **Body fat mass: low at birth (<2% in pigs) but a large increase after birth**

In summary

❑ The postnatal fat expansion

- is associated with changes in cell populations of adult stem cells
- involves the hypertrophy of adipocytes (accumulation of TG in the adipocyte lipid droplets)
- involves the recruitment of adult stem cells to generate new adipocytes

❑ WAT development differs according to depots, species, sex, breeds

❑ IMF content varies according to anatomical muscle origin, age, breeds

❑ Developmental differences in the physiology of adipocytes according to adipose depots (SCAT vs IMF)

Challenges / Future research directions

❑ Challenges for meat quality improvement

- **Sensory quality:** a need to increase IMF content but a need of invisible fat
- **Nutritional quality:** a search of healthy fat (PUFA)
- **Technological qualities:** the requested fat depends on the product

❑ A need to increase our knowledge on

- **adult stem cells:** an emerging field of research for livestock species
- **the mechanisms involved in the control of adipogenesis in different depots**

❑ A search of strategies to better control IMF content to meet the diverse needs of producers and consumers

Thank you for your attention

