

## P-02-37

***Yersinia enterocolitica* grows and remains active in pork packaged under high-O<sub>2</sub> modified atmosphere (#597)**Elina J. Säde<sup>1</sup>, Ilhan C. Duru<sup>2</sup>, Jenni Hultman<sup>3</sup>, Petri Auvinen<sup>2</sup>, Katri J. Björkroth<sup>3</sup><sup>1</sup> University of Helsinki, Dept. of Food Hygiene and Environmental Health, Helsinki, Finland; <sup>2</sup> University of Helsinki, Institute of Biotechnology, Helsinki, Finland; <sup>3</sup> University of Helsinki, Dept. of Microbiology, Helsinki, Finland**Introduction**

*Yersinia enterocolitica* 4/O:3 is a frequent cause of sporadic human yersiniosis in Finland, and contaminated pork is considered its main vector. It is widely agreed that the contamination of meat with *Y. enterocolitica* takes mainly place during slaughtering and processing of carcasses. However, less is known what happens to *Y. enterocolitica* in the distribution chain and how the storage conditions and interaction with other bacteria growing in meat affect *Y. enterocolitica*. Thus, the purpose of this study was to assess if *Y. enterocolitica* is able to survive, remain active and grow as part of developing spoilage microbiome in high-O<sub>2</sub> modified atmosphere packaged pork steaks.

**Methods**

Fresh pork neck steaks (ca. 150-180 g) were obtained from a Finnish meat cutting plant. Steaks were cut to half, and the other half was spiked with *Yersinia enterocolitica* strain 4/O:3 (spiked samples) at levels of ca 6000 CFU/g, and the other half was left untreated (control). Steaks were packaged under modified atmosphere containing 70% O<sub>2</sub> and 30% CO<sub>2</sub> and stored at 5 °C. At days 2 to 9 and at the day 12, 3 spiked and 3 control packages were opened and sampled (25 g) for the following microbiological determinations: *Y. enterocolitica*, *Enterobacteriaceae*, aerobic microbes and lactic acid bacteria on CIN, VRBG, PCA and MRS medium, respectively. Metatranscriptomic analyses were conducted at day 3, 5, 7, 9 and 12: cells were collected from 10 g samples for RNA extraction followed by cDNA amplification, library construction, sequencing, quality filtering and adapter trimming of the reads before mapping of reads other than rRNA.

**Results**

**Microbial numbers and pH:** During the 12-day trial, *Y. enterocolitica* numbers increased from  $2.6 \times 10^4$  to  $3.3 \times 10^6$  CFU/g in spiked samples, whereas in control steaks, the numbers remained below the detection limit

(100 CFU/g) throughout the trial. In most sample, also the numbers of *Enterobacteriaceae*, aerobic microbes and lactic acid bacteria increased during the trial, and at the day 12, colony counts of  $2.0 \times 10^7$ ,  $8.4 \times 10^8$ , and  $6.3 \times 10^8$ , respectively, were recorded. The pH values ranged from 5.74 to 6.59 being lowest at the days 5 and 6 (mean pH 5.91), and highest at the day 12 (mean pH 6.46)

**Metatranscriptomic analysis:** For the spiked samples, the proportion of RNA reads mapped to *Y. enterocolitica* ranged from 0% to 1.86%, whereas most of the reads obtained were mapped to lactic acid bacteria, mainly to species *Leuconostoc gelidum*, *Lactococcus piscium*, *Carnobacterium divergens* and *Carnobacterium maltaromaticum*, and to *Brochothrix thermosphacta*. For the control samples, 12 to 67% of the reads obtained were mapped to the above mentioned species of lactic acid bacteria, and 24 to 48% to *B. thermosphacta*. In addition, a varying proportion (0 to 42%) of RNA reads mapped to *Acinetobacter* spp., with highest (24-42%) proportion obtained in control steaks analyzed at the end of the trial.

**Conclusion**

Our work suggests that *Y. enterocolitica* 4/O:3 is well-adapted to grow in chilled pork packaged under a modified atmosphere with 70% O<sub>2</sub> and 30% CO<sub>2</sub>: *Y. enterocolitica* 4/O:3 was able to grow and reach high numbers ( $>10^6$  CFU/g) in pork and remained metabolically within the developing spoilage bacterial community until the end of the 12-day trial. Furthermore, we showed that *B. thermosphacta*, *L. gelidum*, *L. piscium*, *C. maltaromaticum*, *C. divergens* and *Acinetobacter* spp. were dominating the bacterial communities and played a key role in the microbiomes of spoiled samples. Whereas *L. gelidum*, *L. piscium*, *Carnobacterium* spp. and *B. thermosphacta* are all known meat spoilers, *Acinetobacter* spp. has been less frequently reported in spoiling meat, and thus, its role in the spoilage process deserves further studies.

## Notes